







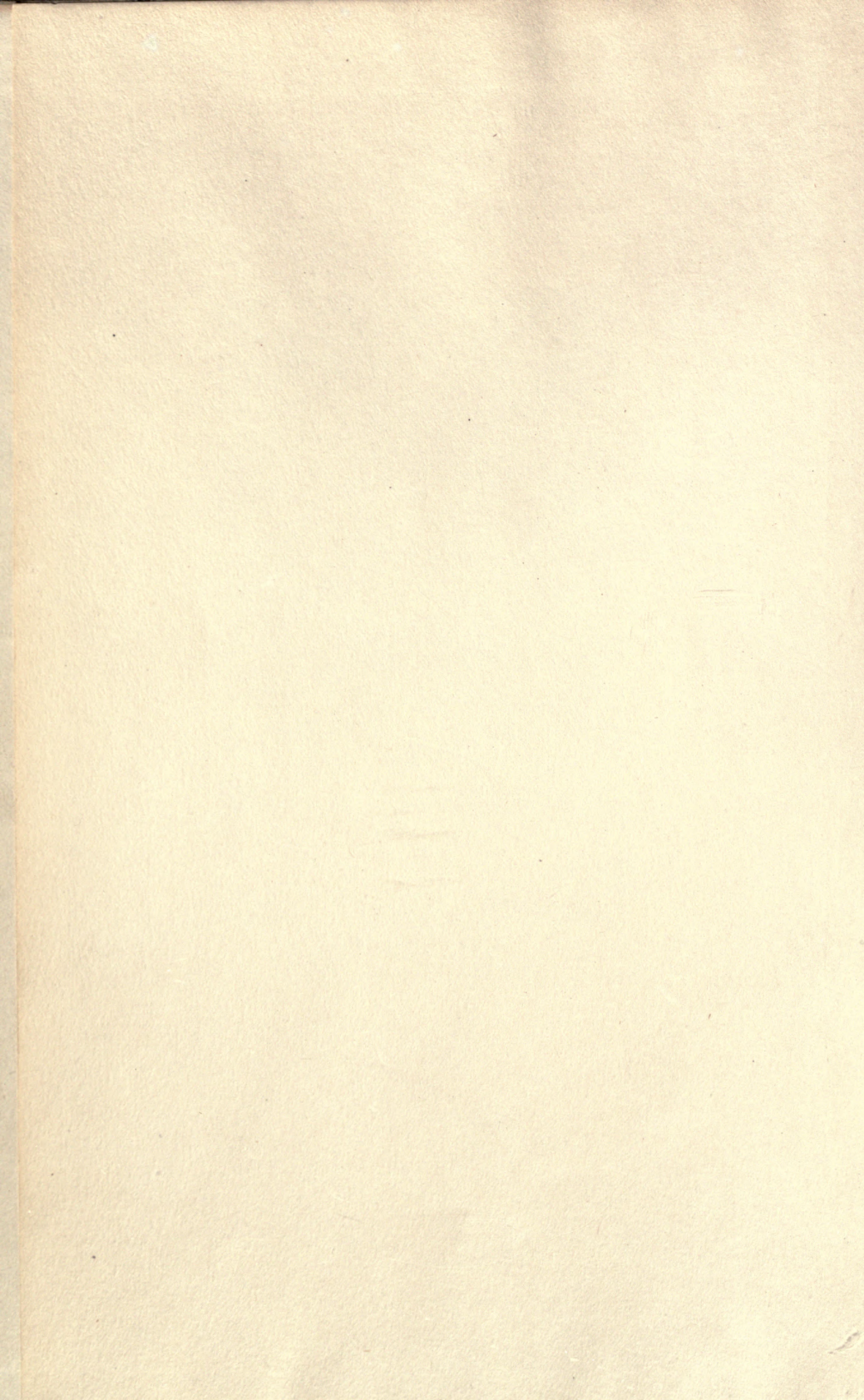
EX LIBRIS

BIOLOGY  
LIBRARY  
G





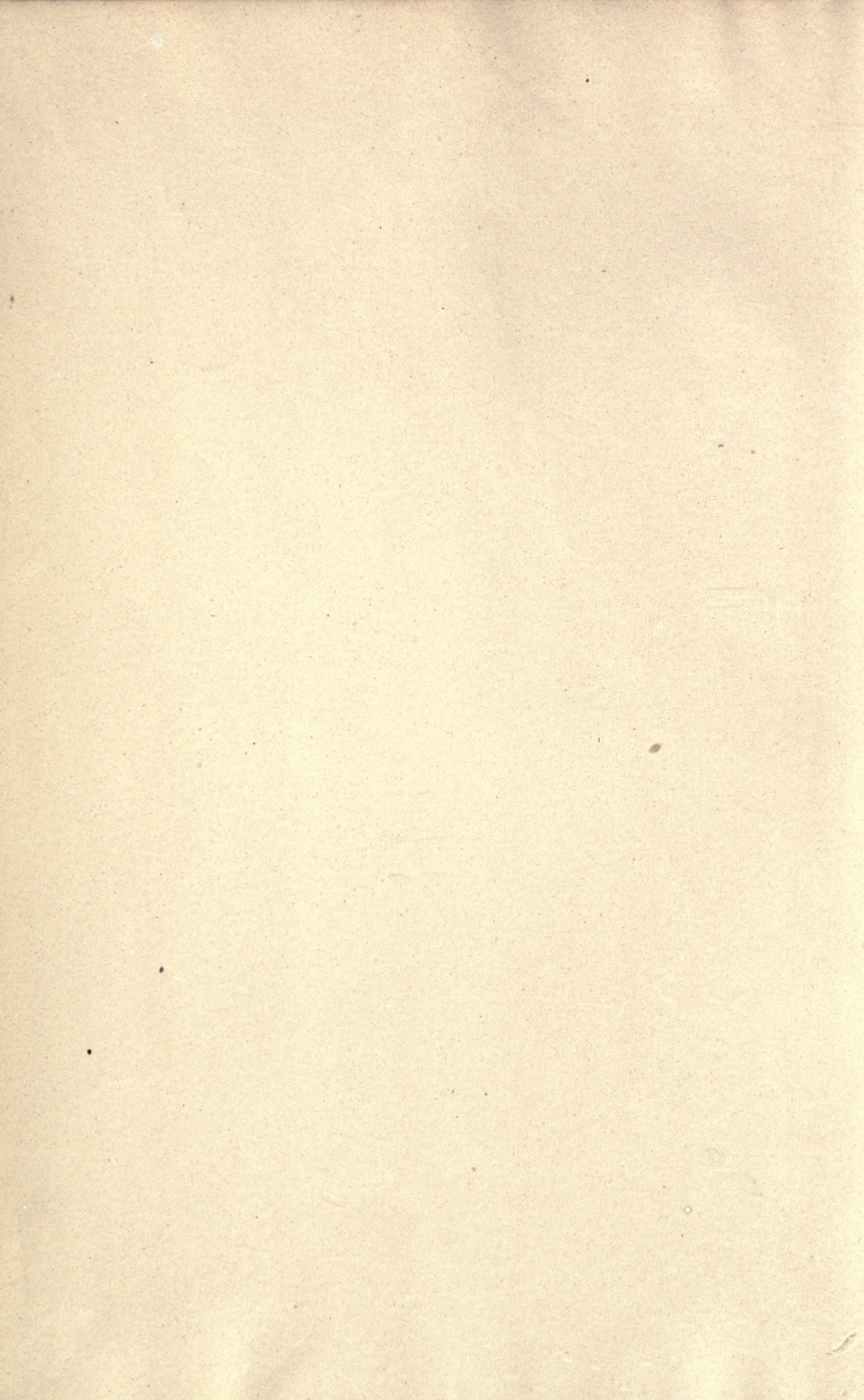














IMMUNITY  
METHODS OF DIAGNOSIS AND THERAPY  
AND  
THEIR PRACTICAL APPLICATION

---

CITRON

---

GARBAT





# IMMUNITY

METHODS OF DIAGNOSIS AND THERAPY

AND

THEIR PRACTICAL APPLICATION

BY

DR. JULIUS CITRON

ASSISTANT AT THE UNIVERSITY CLINIC OF BERLIN, II MEDICAL DIVISION

TRANSLATED FROM THE GERMAN AND EDITED

BY

A. L. GARBAT, M. D.

ASSISTANT PATHOLOGIST AND ADJUNCT VISITING PHYSICIAN,  
GERMAN HOSPITAL, NEW YORK

SECOND EDITION, REVISED AND ENLARGED

30 ILLUSTRATIONS

2 COLORED PLATES AND 8 CHARTS

UNIV. OF  
CALIFORNIA

PHILADELPHIA  
P. BLAKISTON'S SON & CO.  
1012 WALNUT STREET

OR 181  
C-6  
1914  
cop. 2

BIOLOGY  
LIBRARY  
G

Replacing 305417.

COPYRIGHT, 1914, BY P. BLAKISTON'S SON & CO.

TO WHOM  
ADDRESSED



TO  
PROFESSOR FRIEDRICH KRAUS  
AS EVIDENCE OF DUE HONOR AND THANKFULNESS  
THIS BOOK IS DEDICATED  
BY THE AUTHOR  
ON THE OCCASION OF THE OPENING OF THE NEW  
II MEDICAL DIVISION

300085





## PREFACE TO THE SECOND GERMAN EDITION.

---

The first edition of "Immunity, Methods of Diagnosis and Therapy," was, I am glad to say, favorably received. The book apparently fulfilled a demand, as not quite two years have elapsed and a second edition is required. An Italian translation by G. Volpino and an English edition by A. L. Garbat have in the meantime appeared. A Spanish and a Russian publication are in preparation.

During the last two years certain problems in this field have gained greatly in importance, and therefore require more detailed consideration. Special chapters have been allotted to tumor studies and to anaphylaxis. Probably, even this treatise on anaphylaxis will by many be considered too short. Without underestimating the distinct scientific value of the very numerous articles being written on this subject, I feel that the fundamental principles of anaphylaxis are still too theoretical and the practical application still too limited, to warrant a more exhaustive review in a practical book such as this.

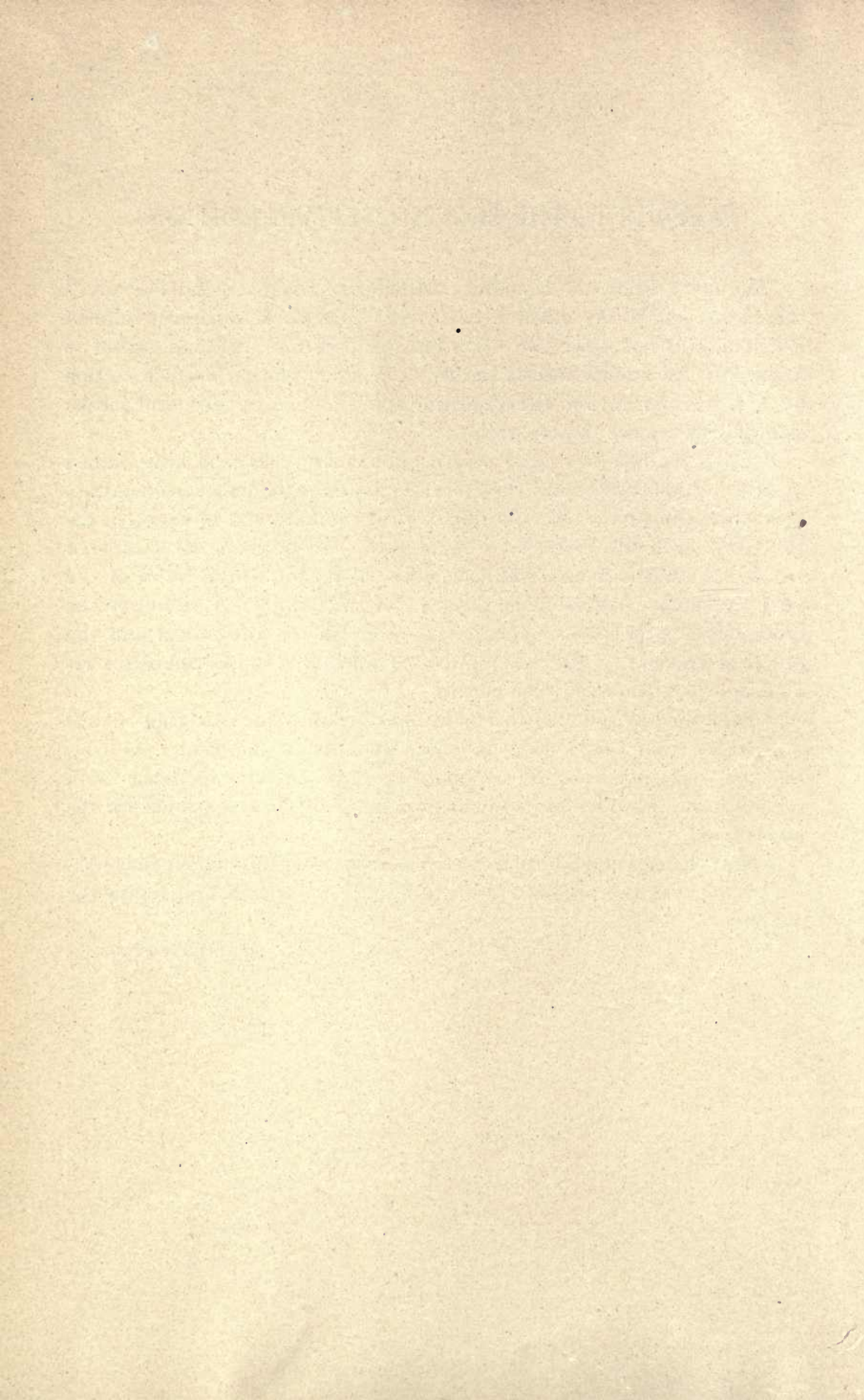
Chemotherapy, on the other hand, has attained such striking clinical importance, that I have dwelt in length upon its development. As ever, the practical elements are emphasized. My broad experience with salvarsan has aided me in distinguishing the points of importance for the practitioner.

Slight changes and additions have been scattered throughout the book.

I hope that this second edition will add new friends and retain the old ones.

JULIUS CITRON.





## PREFACE TO THE SECOND ENGLISH EDITION.

---

The necessity for a second English edition has arisen after an interval of a year. It is a source of satisfaction to note the apparent general approval and demand which this small book on "Immunity" has fulfilled. The credit to Dr. Citron is well deserving. The text of the new English publication has been carefully reviewed and minor changes made, but it follows closely that of the German with its important additions. Like in the first English edition, however, so also in the second, topics of more recent development, treated only very slightly or not at all in the German, have been enlarged or inserted: gonococcus and typhoid complement fixation tests, agglutination and hemolysis tests for transfusion, prophylactic typhoid inoculation, etc. Friendly criticism to discuss some subjects with more detail has been gladly met and corrected, so that a beginner may have no greater difficulty than is only natural. It is hoped that this English edition will meet with the same approbation which I am happy to say has been accorded the former one.

A. L. GARBAT.





## PREFACE TO THE GERMAN EDITION.

---

This book is to serve a purely practical purpose. The methods of serum diagnosis, on account of their growing clinical significance, are constantly stimulating greater interest in all branches of medical science. While giving instruction in this subject, I realized that it would be of great help to both the medical student and physician if they possessed a short text-book which would review in a purely critical form the various methods of immunity diagnosis, especially those relating to tuberculosis and syphilis.

The two systems of *Kolle* and *Wassermann*, and *R. Kraus* and *Levaditi* are doubtless the standards on the subject in German medical literature. On account of their size and price, however, these volumes come to be sought only by the specialist.

It was therefore my aim in this book so to present the subject of immunity that the general medical man, who is even slightly acquainted with laboratory work, can learn the details of the various reactions and their significance. In selecting the different methods, I have taken up those which are used in the clinic for diagnostic, therapeutic, or prophylactic purposes. In addition there are herein included certain fundamental considerations of questions on immunity which for the present are only of theoretical interest, but which owing to the rapid development of the subject, may soon become of practical importance.

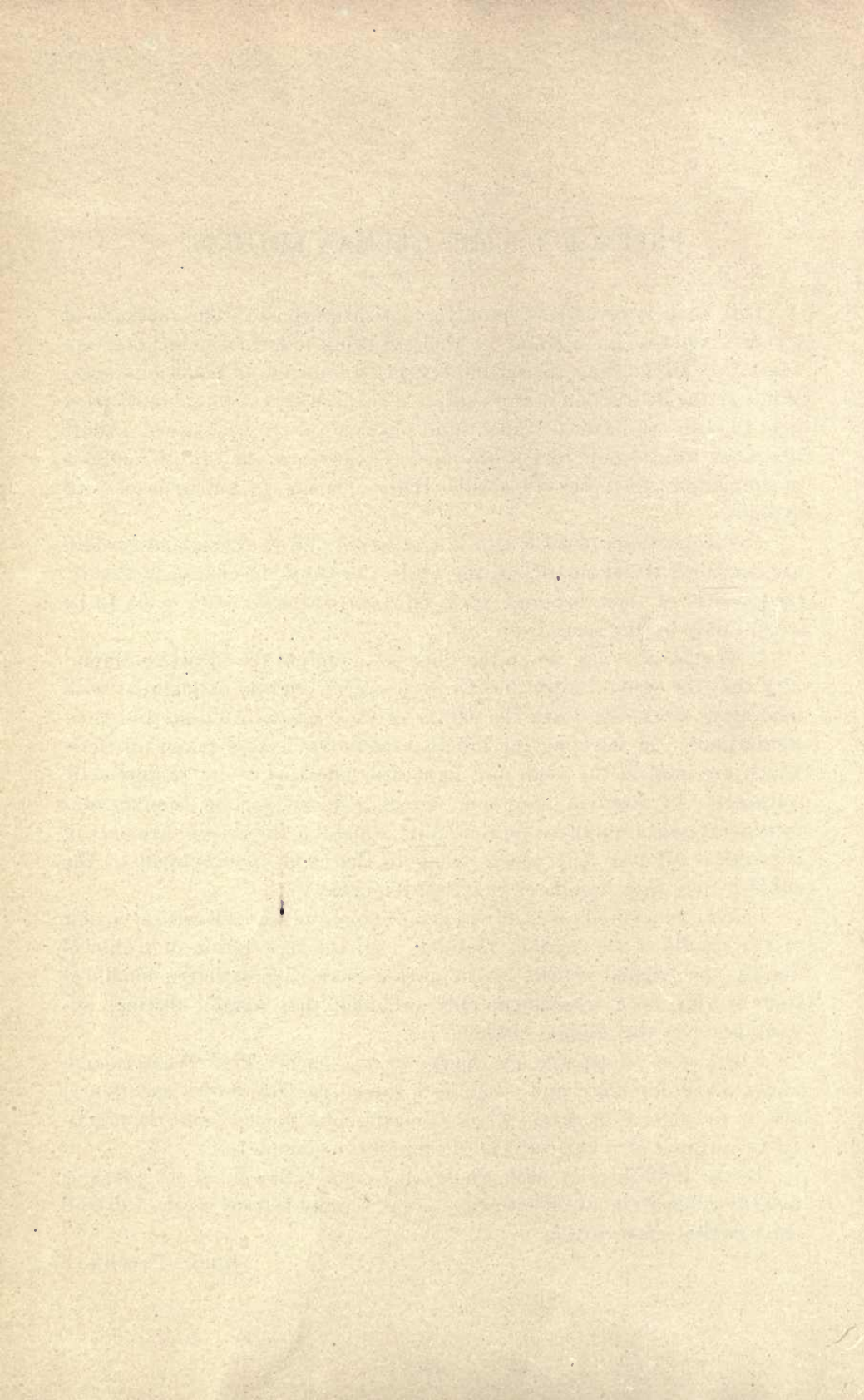
I have endeavored especially to place before the reader a critical review of the results of the various methods. In the description of technical details, the original articles of the author have been selected; modifications having been considered, only provided they exhibit distinct advantages over the original method.

I here wish to express my thanks to my teacher Prof. Wassermann, under whose guidance and stimulus I gained my laboratory experience; also to my chief Prof. Kraus whose clinical genius proved to me the practical importance that this subject of immunity commands.

To the publishers as well, whose kind coöperation in all my plans as regards publication and illustration, greatly simplified my work, I extend my heartiest appreciation.

JULIUS CITRON.





## NOTE BY THE AMERICAN EDITOR.

---

The study of "Immunity," once of merely theoretical interest and purely scientific importance, is to-day no longer such. A realm of practical considerations, considerations which are constantly coming up and enlisting the attention of the busy practitioner have little by little supplanted those phenomena at one time vaguely understood and mostly taken for granted. Gradually have the uncertainties so long dominating and obscuring an intelligent comprehension of the subject been cleared away; mistakes explained; and hypotheses re-established as proven facts. The methods employed for the necessary investigations have naturally improved and increased with such extreme rapidity that a severe task presents itself to one who desires to separate the more from the less valuable ones. It was therefore with extreme satisfaction that I greeted the opportunity of bringing out an English edition of this working hand-book on the various, but most essential methods used in the applications of "Immunity." The author of this volume has, by his exhaustive research and extensive practical experience as a teacher, treated his field with such fulness and preciseness of detail that it is of value not only to the laboratory student, but also to the clinician.

Its already favorable reception in Germany will, it is hoped, be extended to it in America, especially by those whose lack of familiarity with the German language has kept this work beyond their reach.

The chapter on vaccines has been slightly revised and elaborated to conform more closely with the most recently advocated methods of Sir A. E. Wright, to whom the editor is indebted for his experience. Otherwise there has been no need to alter the original text, with the exception that here and there some features which may be of special interest to the English reading public, have been inserted.

I wish in the present connection to express my deep thanks to my teacher Dr. Citron for offering me the privilege of this undertaking, and to the publishers, Messrs. Blakiston & Co., without whose hearty coöperation this would have been impossible, my sincere appreciation.

A. L. GARBAT.





# TABLE OF CONTENTS.

## CHAPTER I.

	PAGE
INTRODUCTION . . . . .	I
Definitions of immunity and antibody. The law of specificity. The necessity of control tests.	

## CHAPTER II.

LABORATORY EQUIPMENT . . . . .	7
General technique. (Technique of injection. The methods of obtaining and preserving serum. Bacterial filtration. Dilutions. Measurement of small amounts of bacteria.)	

## CHAPTER III.

ACTIVE IMMUNITY . . . . .	21
Immunization with living and dead virus. (Vaccination against small-pox; antirabic vaccination; antityphoid inoculation.)	

## CHAPTER IV.

ACTIVE IMMUNITY . . . . .	36
Immunization with bacterial extracts. Aggressin experiments.	

## CHAPTER V.

TUBERCULIN DIAGNOSIS . . . . .	45
Koch's method; cutaneous reaction; Moro's ointment reaction; intracutaneous reaction; ophthalmo-reaction; the specificity and prognostic value of the tuberculin reactions. Mallein. Tricophytin.	

## CHAPTER VI.

TUBERCULIN THERAPY . . . . .	60
The technique of the tuberculin therapy; old tuberculin; new tuberculin; bovine tuberculin. Nastin.	

## CHAPTER VII.

TOXIN AND ANTITOXIN . . . . .	74
The serum therapy of diphtheria.	

## CHAPTER VIII.

TOXIN AND ANTITOXIN (continued) . . . . .	85
Definition of toxin. Tetanus toxin. Botulism toxin. Dysentery toxin. Staphylolysin.	

## CHAPTER IX.

THE TOXINS OF THE HIGHER PLANTS AND ANIMALS AND THEIR ANTIBODIES. FERMENTS AND ANTIFERMENTS . . . . .	95
Snake poison. Paroxysmal hemoglobinuria.	



## CHAPTER X.

AGGLUTINATION. . . . .	
Macroscopic test, microscopic test. Group agglutination. Bacterial agglutinins. Hemagglutinins. Transfusion tests.	105

## CHAPTER XI.

PRECIPITINS . . . . .	120
Bacterial precipitation. Proteid precipitation.	

## CHAPTER XII.

BACTERIOLYSINS AND HEMOLYSINS (CYTOLYSINS) . . . . .	131
Technique of bacteriolytic tests. Pfeiffer's phenomenon. Bactericidal test by plate method of Neisser and Wechsberg. Hemolysins. Cytolysins.	

## CHAPTER XIII.

METHOD OF COMPLEMENT FIXATION . . . . .	152
Principles of this method. Antituberculin. Ehrlich's side-chain theory. Serum diagnosis of syphilis and of diseases caused by animal parasites.	

## CHAPTER XIV.

TECHNIQUE OF THE COMPLEMENT FIXATION METHOD . . . . .	171
The original method of Bordet-Gengou. Wassermann-Bruck's modification. The technique of the serum diagnosis of syphilis. Echinococcus disease. Epidemic meningitis, tuberculosis, gonococcus infections, typhoid fever. The differentiation of proteids according to Neisser-Sachs.	

## CHAPTER XV.

PHAGOCYTOSIS. OPSONINS AND BACTERIOTROPINS. . . . .	196
Technique of opsonic index determination and of Wright's vaccine treatment. Neufeld's method of examination for bacteriotropins.	

## CHAPTER XVI.

MALIGNANT TUMORS . . . . .	216
Studies in reference to their immunity; serum reactions; transplantation experiments. Meiostagmine reaction.	

## CHAPTER XVII.

ANAPHYLAXIS. . . . .	221
----------------------	-----

## CHAPTER XVIII.

PASSIVE IMMUNITY. (SERUM THERAPY) . . . . .	231
Bacteriolytic sera. Special serum therapy.	

## CHAPTER XIX.

CHEMOTHERAPY. . . . .	240
Definition. Atoxyl. Salvarsan. Chemotherapy of malignant tumors.	

## LIST OF ILLUSTRATIONS AND CHARTS.

FIG.	PAGE
1. A room in the laboratory of the Royal Institute for Infectious Diseases (Berlin) . . . . .	7
2. Standard for measuring the size of platinum loops (Czaplewski) . . . . .	8
3. Intravenous inoculation (after Uhlenhuth) . . . . .	9
4. Intraperitoneal inoculation (after Uhlenhuth) . . . . .	11
5. Removal of peritoneal exudate in Friedberger's position (original) . . . . .	12
6. Veno-puncture (original) . . . . .	13
7. Wet-cup method for obtaining blood (original) . . . . .	14
8. Test-tube for preservation of serum (original) . . . . .	15
9. Pukal filter . . . . .	16
10. Filtration through pukal filter . . . . .	16
11. Reichel filter . . . . .	17
12. Lilliputian filter . . . . .	17
13. V. Pirquet's tuberculin test (original) . . . . .	51
14. Ophthalmodiagnosticum for tuberculosis (original) . . . . .	52
15-16. Diagram for the complement fixation reaction . . . . .	154
17. Diagram for the complement fixation in syphilis . . . . .	166
18-20. Technique for the determination of the opsonic index according to Wright . . . . .	204
21-22. Technique for the determination of the opsonic index according to Wright . . . . .	204
23-24. Technique for the determination of the opsonic index according to Wright . . . . .	205
25-26. Technique for the determination of the opsonic index according to Wright . . . . .	206-7
27. Phagocytosis of tubercle bacilli . . . . .	208
28-30. Technique for intravenous injection of salvarsan . . . . .	205-2
<b>CHART.</b>	
1. Example of a diagnostic tuberculin reaction . . . . .	48
2. Example of hypersusceptibility by diminution in the tuberculin dose . . . . .	65
3. Marked increase of weight in a tuberculous individual in spite of continued fever . . . . .	67
4. Treatment with S. B. E., almost without reaction. Immunization against B. E. . . . .	71
5. Opsonic curve after a small dose of staphylococcus vaccine . . . . .	200
6. Opsonic curve during treatment with new tuberculin . . . . .	201
7. Increase in the opsonic index for gonococci by Bier's hyperemia . . . . .	202
8. Auto-inoculation with tuberculin after physical examination and massage . . . . .	202





# IMMUNITY.

## CHAPTER I.

INTRODUCTION.—DEFINITIONS OF IMMUNITY AND ANTIBODY.—THE LAW OF SPECIFICITY.—THE NECESSITY OF CONTROL TESTS.

The diagnosis of infectious diseases can be approached in several ways. In addition to the aid obtained from clinical signs such as the course of the temperature, the changes in the various organs, the exanthemata, etc., the finding of the specific etiological agent of the disease, or the specific antibodies developed by the reaction of the organism, are of equal or even greater importance. The course of an infection depends not only upon the nature, the number, and the virulence of the infecting agents, but also upon the behavior of the infected body. *One must consider a disease as the result of the interaction of both of these factors without necessarily being able to attribute the various symptoms to either the one or the other.* Although the general reaction of the organism is varied, it can nevertheless be shown that in spite of even individual differences, the characteristic bacteria and their products bring about a distinct symptom-complex which is usually concomitant with a significant defense on the part of the organism. The means which the body employs in this protection are *cellular* and *humoral* in nature. In fact, there is a group of infectious diseases in which the cellular reaction predominates, and another in which humoral changes are pre-eminent; and between these extremes are various intermediate forms. Thus, the constantly changing picture of tuberculosis always shows the tubercle as its typical product of cellular reaction; similarly leprosy and syphilis have their peculiar cellular changes. More difficult, however, to recognize by the unaided eye or even the microscope, are the finer biological alterations which take place in the body fluids during the course of infectious diseases. Here, special methods are necessary to detect and differentiate the various humoral changes which occur for the main part in the blood serum. As is known at present, the humoral as well as the cellular immunity reactions are not limited to infectious diseases, but also express normal physiological and pathological conditions. With the conception of Ehrlich's side-chain theory the bridge of understanding for the humoral reaction was built, and it at once became evident how the physiological phenomena of nutrition and production of energy are identical in their nature with processes which



under pathological circumstances lead to the formation of anti-infectious bodies. In an analogous and no less ingenious manner, Metchnikoff has shown that the same cell group of mesenchymal origin which the organism stations against bacterial invasion has physiological and physio-pathological functions to fulfill in the whole animal scale. In the lower animals, these cells aid in the metamorphosis of the body structure, thus leading to the disappearance of entire organs. In the female, they aid in the involution of the uterus after labor, while in the aged they destroy the nerve cells in the senile atrophied nerve centers or finally as chromophages turn the hair gray. *The border-line between the physiological and pathological status is biologically not sharply demarcated.* It is one single chain of manifestations which possess numerous transitional phases. As the methods of serum diagnosis can prove reactions much finer even than those accomplished by chemistry, their application has not been limited to the chapter on infectious diseases.

By their means also, proteids, even though manifest in minutest traces, can be differentiated. Similarly, the secret of blood relationship has begun to be unravelled; and there is a possibility even of solving the problems of metabolism.

Closely associated with serum diagnosis is the serum therapy. Even though the general application of the latter is not as widely developed as that of the former, it must be remembered that through this medium diphtheria has been transformed from a fatal to a combatable disease, and incidentally made the name of Behring immortal. To-day, attempts are constantly being made to treat other bacterial and toxic diseases by specific therapy and it is to be hoped that success will soon be met with.

The study of serum therapy and serum diagnosis is undertaken in various ways. It is comparatively simple to learn only the purely technical details. All large laboratories have trained assistants for the performance of certain reactions or groups of reactions with absolute precision. Although as we have said, they do such work as assigned to them with accuracy, they are nevertheless far from a thorough understanding of the subject of serum diagnosis. Unfortunately this blind method of procedure has recently been advocated to an alarming extent. In addition, the practical success which the Wassermann reaction has met with has inculcated the desire in a certain class of physicians, for carrying out this test alone, and thus to become independent of the use of large laboratories. To meet this demand, short courses have been established and the serum diagnosis of syphilis taught with lightning rapidity. That such a state of events is absolutely injurious is clearly evident. It is impossible for one to be a specialist in a certain reaction and at the same time be ignorant of the other phases in the study of

immunity. Unreliable and erroneous results are the inevitable outcomes of such unscientific work.

The plan followed in this book consists in taking up all of the important principles and methods of immunity, even though at present some may attract no direct practical attention. The principle of the now widely important Wassermann reaction had been described years previously by Bordet and Gengou, but merely from a purely theoretical standpoint. Only with the development of the Wassermann test did it attain its 'practical importance.

To start systematically, it is necessary, primarily, to understand certain terms frequently employed. First, the word of Immunity. immunity requires explanation:

After an individual has recovered from an infectious disease, he passes into a state where he is less or even not at all susceptible to the same infection, although no macroscopical, microscopical or chemical change can be shown to have taken place in his system. This condition is one of immunity. And as the body itself by its own struggle with the invading bacteria has brought about this immunity, it is known as "active immunity." Jenner and Pasteur have employed this mode of immunity acquired spontaneously by overcoming an infection in their principle of prophylactic vaccination. The exact nature of this active immunity is only partially understood. It can be shown, however, that the individuals thus actively immunized have within their organism reaction bodies of a specific nature directed against the infecting elements and their poisonous products. These reaction bodies, which circulate mainly in the blood serum, are known as *Antibodies*.

The antibodies are of different classes depending entirely upon their varied forms of activity. While some, such as the agglutinins and precipitins, have the property of grouping their respective invading agents into small clumps or precipitates, without at the same time embracing protective powers, there are other antibodies which act essentially for the defence of the organism. They attain this by neutralizing the poison of the bacteria (antitoxins) or by destroying the bacteria (bacteriolytins), or so altering the bacteria that the latter can be more easily destroyed by the cells (bacteriotropins, opsonins). The last three types of immunity can be designated respectively, as antitoxic, bactericidal, and cellular immunity. Naturally there are many intermediate forms. It is very probable that besides these well-recognized forms of immunity there may be others, still unknown. Cellular immunity must surely have a far greater range of importance than is at present ascribed to it. There is, no doubt, a distinct cell immunity which acts without the aid of any serum substance and is known as "Tissue Immunity" ("histogene" Immunität).



If the serum of an animal which has been immunized, and containing antibodies, is injected into another normal but non-immunized animal, the latter acquires the power of being immune against the specific infective agent. In this case the immunity was not established by direct cell activity on the part of the animal, for the organism remained passive, and had, as it were, immunity thrust upon it. This form of immunity in contradistinction to "active immunity" is designated as "passive immunity."

The forms of immunity thus far mentioned were all "acquired" either by the spontaneous recovery from the infection or the artificial transmission of the curative antibodies. In contrast, however, to this "acquired" immunity there is a "natural" immunity by which is understood that some animal species are not at all susceptible to certain infections. Thus, man has a natural immunity against a group of diseases markedly fatal for some of the lower animals, *e.g.*, chicken-cholera and hog-cholera. That this natural immunity is almost always cellular in character is undeniably true; and the most important form of this natural armament against infection is the powerful leucocyte, capable of engulfing and destroying the invading enemy. In other words, phagocytosis.

Finally one should speak of a "local" and "general" immunity, meaning to express thereby the different resistance and susceptibility that various organs of the same individual display; and also of a "relative" and "absolute" immunity in order to differentiate quantitatively a transitory immunity from one that is of long duration.

Conception of Antibody. Another term very often employed is "antibody." This, as has already been explained, is a name used to designate the specific bodies which the organism produces as a reaction against the infecting agents and their toxic products. Antibodies are also formed when animals are injected with foreign proteids not of bacterial origin, such as the blood from a different animal species, egg albumin, etc. In order that these antibodies may be obtained, the substances employed must enter the system "parenteral," *i.e.*, some way outside of the gastrointestinal tract.

In older literature the terms antibody and protective body were used synonymously. That is decidedly incorrect, inasmuch as not all antibodies possess the power of protection and not every actively immune organism, demonstrable antibodies. Furthermore, antibodies as the bacteriolysins which are generally considered to have protective powers, and correctly so too, can exist in a system in large numbers without necessarily rendering that organism immune.

How complicated various chapters in the study of immunity can be will be clearly evidenced by a few of the author's experiments with the hog-cholera

bacillus. Rabbits rendered actively immune by inoculation with extracts of hog-cholera bacilli possess a serum which when injected into an animal of a different species, as the guinea-pig, will render the latter passively immune. If, however, the serum is injected into another animal of the same class (another rabbit), no protective power is transmitted. In other instances it was shown that the rabbit which was being treated with the purpose of active immunity was in reality never immune, as it always succumbed when injected with living bacteria even though its serum contained bodies which were perfectly able to passively protect guinea-pigs against the same deadly infection.

Just as it is incorrect to consider an antibody and protective body as one and the same thing, it is equally erroneous to deny the existence of protective bodies, because their presence cannot be demonstrated by a certain method of laboratory examination. It must be kept in mind that there are still many unsolved problems in the subject of immunity and that therefore only the positive findings should be the basis for drawing conclusions.

In order to learn the nature of these antibodies attempts have been made to isolate them chemically. Thus far all such trials have been unsuccessful. It is even uncertain whether these so-called antibodies are definite chemical entities. Only the effects of the serum as a whole are known, and the ingredients in it to which these activities are attributed are thought of as antibodies. For didactic purposes antibodies, as antitoxins, agglutinins, etc., will be spoken of in this book when the antitoxic or agglutinating properties, exclusively, are meant.

In spite of the individual differences which are ascribed to the various classes of antibodies, there is one quality possessed by all—their specificity. To explain this by a rather crude example, may be mentioned the fact that typhoid antibodies will give their various reactions of immunity only when these are performed with the typhoid bacillus, and cholera antibodies only when performed with the cholera vibrio. Substances which lack this essential property of specificity cannot be considered antibodies, although they may fulfill all other requirements. There are indeed limitations to this fast rule, but these will be considered subsequently. For the present the following can be taken as a fixed fact; namely, that every true antibody is absolutely specific, and that all substances or bodies which are not specific cannot be real antibodies. *The law of specificity is the fundamental principle of serum diagnosis.* As soon as the specificity of a reaction becomes doubtful, its diagnostic importance suffers greatly. In the following pages, therefore, the question whether or not a reaction is specific will be repeatedly discussed, and it will be the aim in every way possible, especially by the use of control tests and experiments to outline the limits of this specificity. Here, even at so early a stage of the discus-



sion, the absolute necessity of these control tests must be urged; though it may appear superfluous to the beginner, that for apparently simple experiments, controls are performed which consume more time than the actual diagnostic test itself. Probably also the desire will arise, and perhaps be satisfied, to omit these control experiments. Notwithstanding the possibility that for a long time perfectly good results will be obtained, it cannot be too often or too emphatically impressed upon all workers in immunity methods, that the only guard against mistakes and failures in diagnosis is necessarily found in control tests. And especially in doing research work, the latter are indispensable. For, experimental work which involves reasonable possibilities, or has any pretension towards plausibility, warrants no true scientific conclusion without the employment of such tests.

The author has made it a rule, whenever new findings in serum diagnosis are published, always to look for the given control experiments. If these are insufficient, then no matter what the contents are, the value of the research is slight, for all its claims only may, but not necessarily must, be correct.

## CHAPTER II.

### LABORATORY EQUIPMENT.—GENERAL TECHNIQUE.

Although some of the tests of serum diagnosis are comparatively simple and can be performed in one's office or even at the bedside, in most instances a laboratory equipment is essential. This of course does not at all imply the necessity of such elaborate apparatus as one is accustomed to find in our present up-to-date bacteriological or serological

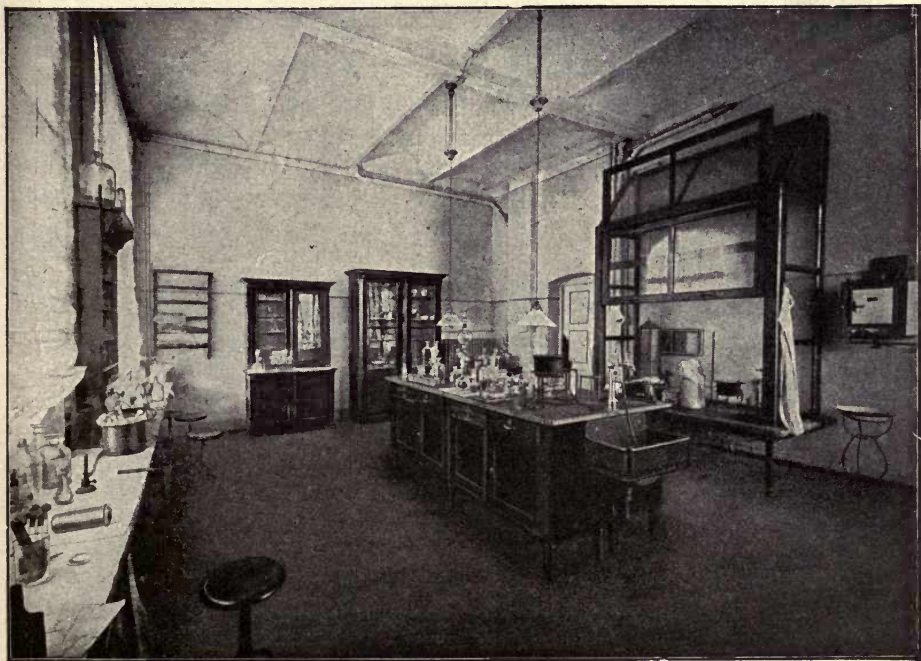


FIG. 1.—A room in the laboratory of the Royal Institute of Berlin for the study of infectious diseases.

laboratories where a great deal of complicated research work is done. For the practical application of serum diagnosis, as employed at the hospital or in private practice, an outfit much less costly is perfectly sufficient. As regards the question of a room, the selection of one with two windows, allowing the entrance of sufficient light, is indispensable. At the same time, however, some arrangement should be made in connection with the windows in order that the direct rays of the sun be prevented from striking one's desk. Strong sunlight may weaken or even destroy



the virulence of cultures, or bring about many changes in sera. Even diffuse daylight should not be considered as entirely inert. A general rule to be remembered is never to expose any biological reagent, be it a bacterial culture or any form of its derivative, a serum, or any other substance to daylight any longer than is absolutely necessary. If this dictum is followed, one will avoid many a difficulty.

To conform with this idea, it is wise to have upon the table a small closet into which the cultures and sera can be placed for the time that they are being used. Such a convenient receptacle can be made out of a large cigar box, painted black inside and out, with its lid replaced by a small black curtain.

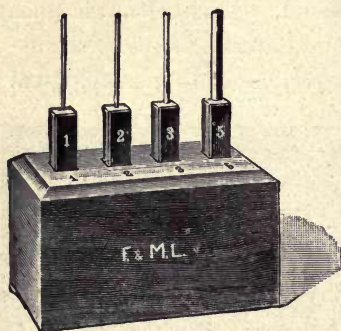


FIG. 2.—Instrument (After Czapelewski) for the standardization of platinum loops.

The table or desk at which one works should be near the window, and covered with filter-paper, upon which should come a glass or asbestos plate. Instead of a wooden table it is certainly more elegant, but costlier to have a top plate of glass. Upon the table there should be a Bunsen burner, a microscope, a lamp for microscopic work at night, a dish filled with sublimate or cresol into which the infected substances, old cultures, used pipettes and graduates are placed.

It is very convenient to have running water and a hood in the same room. Still neither of these is absolutely necessary. As for larger apparatus—must be mentioned, a thermostat, a mechanism for shaking, a dry sterilizer, a good autoclave, a water-bath, an instrument sterilizer, a water or electrical centrifuge, an ice chest, a closet for instruments and glassware, and finally animal cages of the kind that are easily cleaned.

As for instruments and glassware the following are required: scalpels, scissors, forceps, glass-cutter, sterilizable syringes of various sizes, graduates of 10, 25, 100, and 1000 c.cm. capacity each, pipettes of 1 c.cm. with 1/100 divisions and pipettes of 10 c.cm. with 1/10 divisions, a sterilizable pipette retainer, Erlenmeyer flasks, Petri and Kolle's dishes, test-tubes, dark glass flasks, ordinary water glasses, funnels, glass tubing of various sizes, and test-tube racks. Furthermore, a platinum needle and a platinum loop are required. For making a loop of a definite size, and one which can always be referred to, the small instrument devised by Czapelewski is of great help. It consists of four round metal bars 1, 2, 3, and 5 mm. in diameter around which the platinum wire can be twisted in order to make a standard loop (Fig. 2).

For making a loop of a definite size, and one which can always be referred to, the small instrument devised by Czapelewski is of great help. It consists of four round metal bars 1, 2, 3, and 5 mm. in diameter around which the platinum wire can be twisted in order to make a standard loop (Fig. 2).

All instruments and glassware used for serum work should be perfectly clean. It is best to have all the glassware plugged with non-absorbent cotton, and sterilized by dry heat. It is never advisable to clean the glassware with strong acids, alkalies or other strong chemicals. If this has been done, the chemicals must be thoroughly removed by washing, as the slightest trace may interfere with the accuracy of some tests.

All used glassware should at once be placed into a disinfecting solution. For this purpose, lysol, lysoform and cresol solutions are highly recom-

mendable. Sublimate is less efficient because it coagulates albumins and thus may lead to plugging of pipettes which may have contained blood rests. If highly infectious material has been examined, it is best to place the entire disinfectant solution containing the used glassware into the autoclave, sterilize it there, then wash the supply thoroughly with soap, dry and resterilize it by dry heat for 1 to 2 hours at 120° C.

### The Technique of Inoculation.

Both for serum diagnosis and serum therapy, the serum is required from animals which have been artificially immunized against the bacteria or their products of secretion. Almost without exception, this immunization is produced by injecting the animal with the infectious virus. The method of inoculation is either *intravenous*, *intra-peritoneal*, or *subcutaneous*.

The technique of *intravenous injection* varies somewhat with different animals. In *rabbits*, the veins running along the outer margins of the ears are most suitable. The assistant sits upon a chair, holds the hind legs and body of the rabbit tightly fixed between his knees and thus has his hands free to steady the rabbit's ears. Another method consists in placing the rabbit upon the

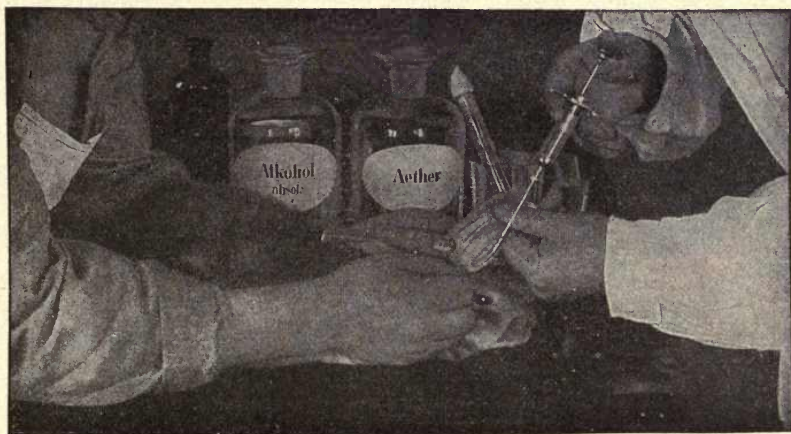


FIG. 3.—Intravenous inoculation. (After Uhlenhuth.)

table and firmly holding him there while the injection is made (Fig. 3). The ear is first struck gently with the fingers and washed with alcohol and xylol. If the hair is very long, it should be clipped. If the vein running along the outer margin of the ear is exceptionally small, it can be made more prominent by compressing it between the thumb and index finger at the root of the ear. No force should be used with the injections; the fluids should be allowed to flow into the blood stream very slowly. Glass



syringes, or such as can be sterilized easily, are preferable. Air bubbles are to be carefully guarded against in order to exclude the danger from air embolism.

If infectious material is used for injection, it is advisable in such instances to place a small piece of cotton moistened in alcohol or a 5 per cent. carbolic acid solution around the point of union between the needle and the barrel of the syringe to prevent the possible escape of any fluid which usually occurs at this point.

After inoculation is completed, the needle should be quickly withdrawn, a small piece of non-absorbent cotton placed upon the needle puncture and compression applied. If non-virulent bacteria or albumin is injected, the bleeding may be almost instantly controlled by firmly squeezing the vessel above the puncture wound with the edge of one's finger nail.

In guinea-pigs intravenous inoculation is more difficult, as here there are no large superficial veins. The Jugular or Iliac vein is therefore chosen, and must be dissected free. It is not necessary to tie off the vessel, but the wound should be firmly compressed by means of clean gauze or cotton. Morgenroth has substituted the simpler method of

Intracardial *intracardial* inoculation. The point of maximum pulsation of Injection. the heart to the left of the sternum is made out by palpation and a thin sharp needle is inserted at the specified area. The spurting of blood indicates that the needle is within the heart. Thereupon the already filled syringe is carefully fitted on to the needle and the contents slowly injected. The syringe is then detached from the needle and blood is again allowed to spurt out in order to be absolutely convinced that the needle was still in the heart. It is next quickly withdrawn. By this method it is possible to inject about  $1\frac{1}{2}$  c.cm. directly into the blood stream.

In dogs, sheep, goats, horses, etc., the intravenous injection is given into the jugular vein directly through the skin which must be thoroughly shaved, cleaned and disinfected. Compression by the finger makes the vein more prominent.

In dogs the popliteal vein is frequently selected. In man the intravenous injection is given into one of the veins on the anterior surface of the elbow joint.

Several general rules are to be observed when giving intravenous inoculations. First of all, only *small* quantities of fluids should be injected; secondly, the temperature of the fluids for injection should not differ from that of the body; thirdly, substances that are strongly hemolytic may produce marked disturbances or even sudden death of the animal; fourthly, if an animal is to be frequently inoculated it is best to puncture the vein for the first inoculations as far peripherally as possible and give each subsequent injection more centrally, for very often thrombi are formed at the site of inoculation.

Intraperitoneal Injection. INTRAPERITONEAL injection is employed most frequently among rabbits and guinea-pigs. The main danger associated with this method is possible injury to the intestines; but by heeding the following advice, this can be prevented. The

animal should be fixed or held head down. In this position, the loops of intestines tend to sink toward the diaphragm. This is further helped along by gentle downward massage over the abdomen thus leaving an area, above the bladder, which is sometimes free from intestines. Another protective

measure consists in using a blunt canula which can be made by breaking off the sharp point of the needle. As it is at times difficult to pierce the skin with this blunt instrument, it is advisable to previously make a minute incision through the cutis and subcutis with a sharp pair of scissors and pass the needle through this small opening. The needle should not be plunged directly into the peritoneal cavity, because at the withdrawal, the injected fluid easily escapes through the punctured opening. First, it is inserted subcutaneously upward, in the long direction of the animal; then the hand is raised and the needle forced horizontally forward through the peritoneum, thus leaving the opening in the peritoneum at a different level than the one through the muscles and fascia, thereby making the escape of fluid more difficult. One readily realizes that he has gone through the peritoneum by a relaxation of the reflex abdominal rigidity (Fig. 4).

For the intraperitoneal injection in guinea-pigs, *Friedberger* has devised a procedure which is very satisfactory and furthermore does away with the necessity of an assistant. It can also be employed in *Pfeiffer's* test for the removal of exudates from the peritoneal cavity. The guinea-pig is allowed to creep into the breast pocket of the laboratory gown until its head and thorax are inside of the pocket. Its hind legs are grasped between the middle and ring fingers of the left hand and flexed on its back, thus giving a free exposure of the lower parts of the abdomen (Fig. 5).



FIG. 4.—Intraperitoneal injection of rabbit.  
(After Uhlenhuth.)



Sub-  
cutaneous  
Injection.

SUBCUTANEOUS inoculation is the simplest of all methods. A fold of skin is elevated between the thumb and index finger of the left hand and the needle plunged into the subcutaneous tissue. In rabbits and guinea-pigs the skin of the back or abdomen is chosen, as the subcutaneous tissue here is not tense. In goats, sheep, and horses the skin of the neck and shoulder region is preferred.

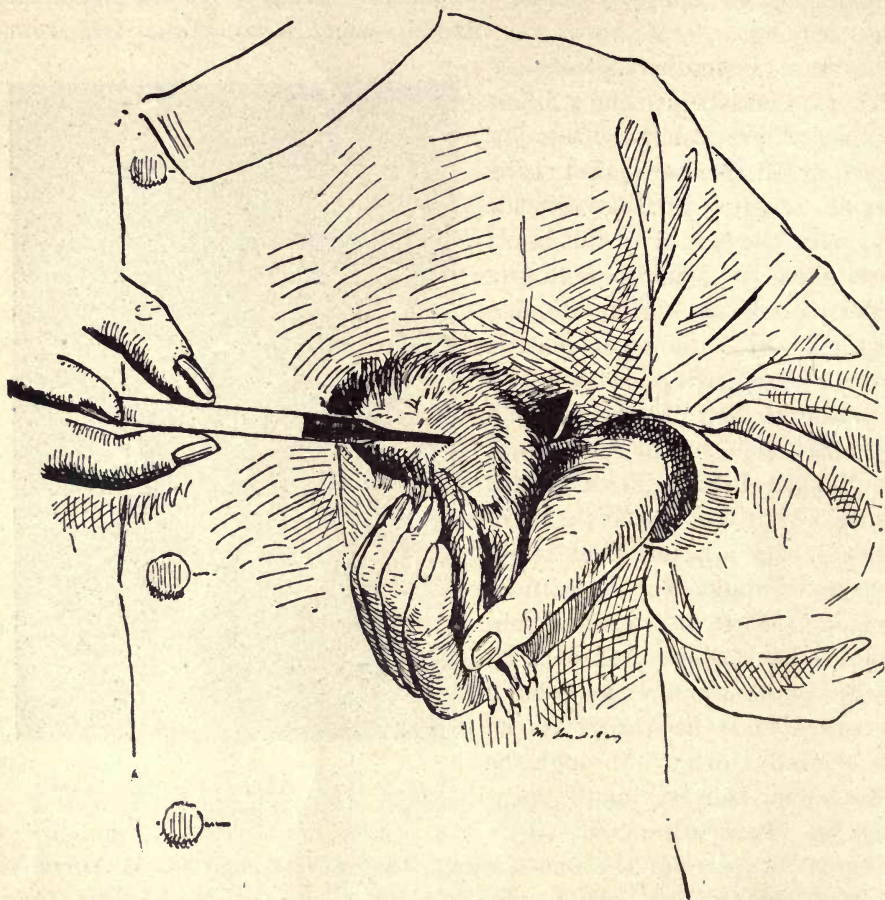


FIG. 5.—Removal of peritoneal exudate. Guinea-pig held in Friedberger's position. (*Original.*)

The skin of the back and abdomen is to be avoided because following the injection edema frequently arises, which may extend to the lower extremities and thus interfere with locomotion.

If abscesses arise after subcutaneous injection, they should be opened, washed out with lysol solution and dressed with iodoform gauze.

### The Methods of Obtaining and Preserving Serum.

Venesection or venous puncture is the method best adapted for obtaining blood from animals. The veins employed for that purpose are those

which have already been mentioned in connection with intravenous injections. A simple, large, hollow needle is all that is required. Suction with a syringe is superfluous. Only in Morgenroth's method of removing blood directly from the heart of guinea-pigs is aspiration necessary. From rabbits enough blood can be collected by making an incision into the vein along its long axis with a sharp knife, or by dividing the vein transversely with the scissors. The blood thus collected is not absolutely sterile.

In man, if only a small quantity of blood is required, it can be obtained from the finger or ear. If, however, a larger amount is necessary, puncture of one of the veins in the bend of the elbow with the Strauss canula is resorted to. It goes without saying that this area must be properly disinfected with soap and water, ether, alcohol or sublimate solution. Wright's method for collecting moderate quantities of blood will be reviewed in the chapter on opsonic studies.

If the vein is prominent, the canula is thrust into the vein directly through the skin. Here the author has found it more convenient to point the canula upward, *i.e.*, in the direction of the blood stream.<sup>1</sup> In cases where the vein does not stand out it can be made to do so either by applying pressure with the finger upon its central part or placing a tight rubber bandage or rubber tube about the arm. This should not, however, be tight enough to obliterate the radial pulse. In very fat individuals, occasionally even these means may not suffice so that the vein must be dissected free and incised. After completion of the venesection the arm is elevated, slight pressure made upon the wound with sterile cotton and a bandage applied. If a small amount of blood is sufficient, and, as in most serological examinations absolute sterility is not essential, venesection can be replaced by the method of wet-cupping. For this procedure a scarifier and Bier cup are required. The technique is as follows (Fig. 7).

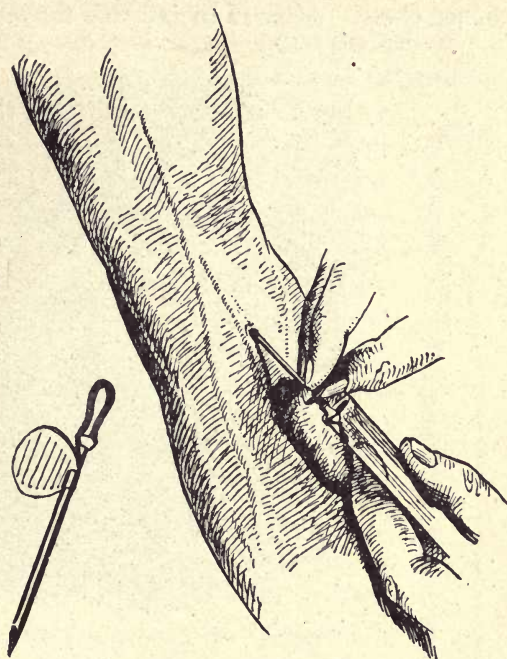


FIG. 6.—Puncture of vein. (*Original.*)

<sup>1</sup> The editor has found that more blood is obtained by thrusting the canula into the vein in the reverse direction.



Some part of the skin of the back is thoroughly disinfected and a well-fitting Bier cup firmly applied. Suction arises and the skin assumes a dark bluish-red appearance. After half a minute the cup is removed, the scarifier applied and the cutting edges set free. The scarifier is then reapplied, but this time at right angles to the previous incisions and the edges again set free. Suction is again made by the Bier cup and the blood is thus forced out from the multiple incisions.

The blood obtained by any of the above methods is collected into a sterile vessel (graduate, flask, test-tube) and allowed to coagulate. The clot is then separated from the sides of the vessel by a sterile glass rod or platinum needle, the vessel plugged with absorbent cotton and placed into the ice chest. After 12 to 24 hours the serum begins to separate out from

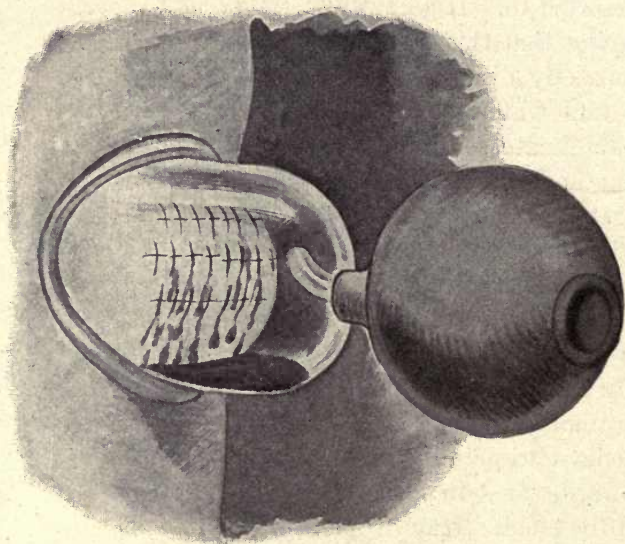


FIG. 7.—Obtaining blood by the wet-cupping method.

the clot. If the serum is required immediately, the blood is allowed to flow directly into centrifuge tubes, the clot separated from the sides and the tubes centrifugalized. With a well-regulated centrifuge serum appears after several minutes.

There are several rules to be kept in mind when using a centrifuge.

- |             |   |
|-------------|---|
| Rules for   | 1. The machine must be well oiled.  |
| the Use of  | 2. The counterbalance must be absolutely of the same weight.  |
| Centrifuge. | 3. The centrifuge should never be suddenly stopped, but allowed to do so of its own accord.   |
|             | 4. In starting it, the motor should be gradually turned on.   |
|             | 5. If the centrifuge is slightly out of order it should not be used, but repaired at once, otherwise it may be ruined forever.        |
|             | 6. One should never centrifugalize with cotton plugs in the test-tubes. If the latter must be sealed, rubber stoppers should be used. |

The color of a serum is greatly variable, depending mainly upon its hemoglobin or fat content. Blood taken at the height of the period of digestion shows a chylous serum. The serum of nursing women contains milk, that of icteric people contains bile. For most serological examinations these elements in the serum are inert, and do not interfere with the reading of the results. In precipitin reactions, however, the serum must be absolutely clear.

If serum is to be kept for a long time, there are several ways that it may be retained without losing its activity. The method chosen depends upon the serum substance which is to be preserved.

As will be pointed out again, substances are either *thermostabile* or *thermolabile*. The preservation of thermostabile substances (agglutinins, amboceptors) is usually very simple. It is sufficient to place the clear serum, which has separated from the clot, into a sterile test-tube plugged with absorbent cotton, and to put it into the ice chest away from the light. To reassure its perfect preservation one may add to it some phenol in such proportion that the carbolic is present to the extent of a  $1/2$  per cent. solution, e.g., to 9 c.cm. of serum add 1 c.cm. of a 5 per cent. phenol solution. The latter should be added drop by drop and agitated, so as to avoid the formation of precipitates. Another method, which the author employs almost exclusively for the preservation of sera containing amboceptors, consists simply in heating the sera at  $56^{\circ}\text{C}$ . for a half hour and then placing them into the ice chest. Inactivation has the advantage of stopping molecular changes produced by ferment actions of fresh serum. Furthermore, heating acts as a sterilizer for isolated air germs which may have found their way into the serum during the process of getting it. In this form, a serum can be kept in the ice box for several weeks without any material change. Occasionally one finds that a serum will undergo contamination in spite of inactivation, so that if a serum is to be preserved for several months, it is advisable to seal it in a test-tube. For this purpose a brown glass tube slightly drawn out at its upper end is employed (Fig. 8). The serum is placed into this sterilized tube and then the latter is sealed in the flame at its narrow part. Bacterial and organ extracts are well kept in this way. The best method of preservation consists in evaporating the serum to dryness in a *vacuum desiccator*. This procedure is rather complicated and can therefore be employed only in institutions.



FIG. 8.—  
Tube used  
for preservation of  
fluids.  
(Original.)

A vacuum desiccator with heatable plates is used. The serum is poured in very thin layers in sterile flat dishes and allowed to dry out in the desiccator at a temperature of  $30^{\circ}\text{C}$ ., later on at  $35^{\circ}\text{C}$ . in a vacuum of 3 cm. mercury. The dried serum



forms a yellowish-red horny mass which is scraped off from the dish and ground up in a mortar into a yellowish powder. The serum powder is then sealed in a brown glass tube.

When this dried serum is to be used, the tip of the ampoule is broken off, and several drops of isotonic salt solution at a temperature of  $30^{\circ}\text{C}$ . are poured in, in just sufficient an amount to moisten the wall of the glass tube. By rolling the tube to and fro, one finds that the serum powder will easily stick to the moistened wall. The granules are allowed to swell up and after they have done so, enough isotonic salt solution is added to make up the original volume.

For the preservation of thermolabile substances, the method of freezing has been suggested. *Morgenroth* has devised for this purpose a simple and handy apparatus named *Frigo* which can be obtained from *Lautenschläger, Berlin*. Although for most tests this method of preservation has been employed with success, *Neisser's* clinic reports that sera preserved in the *Frigo* with the idea of retaining their complement did not give as accurate complement fixation experiments as did similar fresh sera.

*Friedberger* advises the addition of 8 per cent. salt solution for the preservation of the complement. When the serum is to be used it is diluted tenfold with distilled water, so that a 10 per cent. dilution of complement is obtained. By the addition of the salt, the resistance against harmful effects of light, room and body temperature, and chemical substances like phenol is increased, but the thermolability of the complement remains the same. Drying a serum in a desiccator is not to be advocated for the preservation of the complement, as during such procedure a portion of the complement is lost. Once the serum is in its dried form, however, the remaining complement is retained and in addition, has become resistant against high heat.

### Filtration of Bacteria.

It is important in many serological studies to be able to separate bacteria from their fluid media or suspension. This is accomplished either by centrifugalization or filtration. The first method does not completely

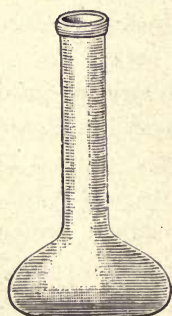


FIG. 9.—Pukal filter.

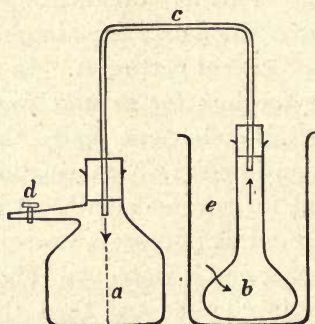


FIG. 10.—Filtration through a Pukal filter.

free the fluid of its bacteria, but if this is desired the method of filtration is essential. In this connection, however, one must bear in mind that many albuminous, or albumen-like substances, few colloids and even some toxins, do not pass the filters and are therefore held back. Bacterial filtration is simplified by preliminary centrifugalization or passing the fluid through

filter-paper. Different porous materials have been used for bacterial filters, of which especially suitable are porcelain, infusorial earth and asbestos. The filtration apparatus consists of the respective filter and the receptacle which receives the filtrate. Filtration takes place by differences in pressure, where either the fluid is forced through by high pressure or sucked through by a vacuum formed in the receiving vessel. The following are some of the filters most commonly in use.

1. CHAMBERLAIN'S CYLINDER FILTER, F, used in the Pasteur Institute at Paris. The filter cylinder is made of infusorial earth and may be attached to any water outlet.

2. PUKAL FILTER, made of burnt kaolin, is used especially for the filtration of large quantities of fluid. The filter *b* is placed into the beaker *e* containing the toxin and bacterial fluid. The filter is then closed by a rubber stopper, perforated by a central opening through which runs a glass tube bent at right angles, and this in turn is connected with either an air or water pump for producing a vacuum inside of the filter. Between the filter and vacuum pump can be interposed a sterile jar *a* (Figs. 9 and 10).

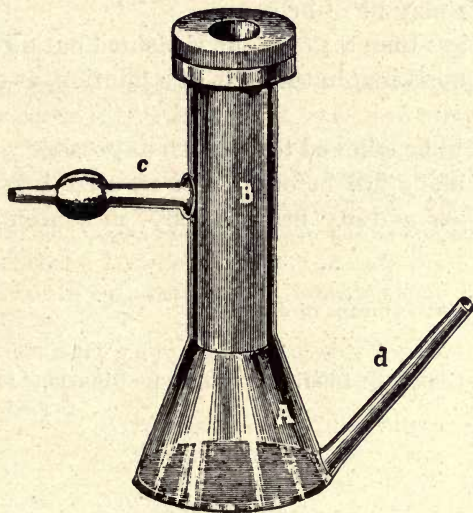


FIG. 11.—Reichel filter.

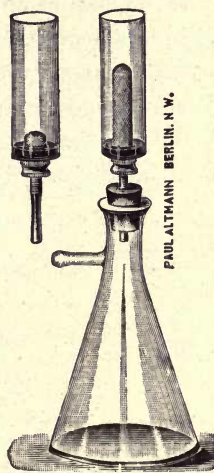


FIG. 12.—Lilliputian filter.

3. THE REICHEL FILTER (Fig. 11) consists of a glass receiver A, having a side neck *c* and at the bottom a tube-like outlet *d*. A porcelain filter B fits into the glass jar and rests upon the margin of the flask by means of a broad collar. The point of junction is made air tight by means of a rubber cap with a central opening, through which the cylinder can be filled. When in use *d* is shut off by a rubber tube with a pinch cock and *c* in which lodges a small piece of cotton is connected with a water pump that is instrumental in bringing about a vacuum. The function of *d* is to allow the removal of samples of the filtrate and finally to obtain the entire filtrate.

4. THE LILLIPUTIAN FILTER, candle-like in shape, and made of infusorial earth, is employed for the filtration of very small quantities. The filter is cemented upon a metal tube which is screwed, so that it is air tight, into a well-fitting glass cylinder open at the top. The tube is passed through a rubber cork which tightly closes an exhaust



flask. The fluid to be filtered is placed into the glass cylinder and sucked through into the flask by means of a vacuum produced here. For the purpose of collecting very small quantities a test-tube may be placed into the exhaust flask (Fig. 12).

### Preparation of Dilutions and Measurement of Small Amounts of Bacteria.

*All serological methods are to be considered on quantitative bases. In serum diagnosis as well as in the therapy, the amount of the serum employed is the deciding factor.* Similarly, the number or amount of bacteria required either for the purposes of immunization or serological reactions is of extreme importance.

*One cubic centimeter is the unit of measure for serum and all fluid material (Bouillon cultures, exudates, etc.). If small quantities are required, it is best to dilute the fluid with 0.85 per cent. saline solution. The exact preparation of dilutions is one of the most essential technical procedures in serum diagnosis.* Some general rules may be of help.

1. Never should amounts less than 0.1 c.cm. be measured out directly. For beginners even 0.1 is best measured in the form of a dilution, as errors are apt to occur very easily.

2. The *decimal system* should be adhered to as much as possible.

3. The dilution should be made just before it is to be used, inasmuch as many substances retain their activity best, or only, in concentrated form.

The following is an example of correct forms of dilutions:

Toxin	Dilution of toxin, 1 : 10	Dilution of toxin, 1 : 100	Dilution of toxin, 1 : 1000
0.1 c.cm. =	1 c.cm.		
0.05 c.cm. =	0.5 c.cm.		
0.01 c.cm. =	0.1 c.cm.	= 1 c.cm.	
0.005 c.cm. =		0.5 c.cm.	
0.001 c.cm. =		0.1 c.cm.	= 1.0 c.cm.
0.0005 c.cm. =			0.5 c.cm.

The stock dilution of 1 : 10 is made by measuring off 1 c.cm. of toxin and adding 9 c.cm. of 0.85 per cent. of saline.

The dilution 1 : 100 can be made by taking 1 c.cm. of toxin and adding 99 c.cm. of saline. It is more practicable, however, to take 1 c.cm. of the 1 : 10 stock dilution and add 9 c.cm. of saline. If the dilution 1 : 10 is not present and only a small amount of the dilution 1 : 100 is desired, the latter is made by taking 0.1 toxin : 10.0 NaCl sol. Similarly 1 : 1000 = 0.1 : 100 = 1 c.cm. of the dilution (1 : 10) : 100 = 1 c.cm. of the dilution (1 : 100) : 10.0.

The following table shows the details of various dilutions:

Dilution 1 : 10	Dilution 1 : 100	Dilution 1 : 1000	Dilution 1 : 10,000
1 c.cm. + 9 c.cm. NaCl sol. = 0.1 c.cm. + 0.9 c.cm. NaCl solution. = 0.2 c.cm. + 1.8 c.cm. NaCl solution. = 0.3 c.cm. + 2.7 c.cm. NaCl solution. = 3 c.cm. + 27 c.cm. NaCl sol. = 10 c.cm. + 90 c.cm. NaCl sol.	1 c.cm. + 99 c.cm. NaCl sol. = 0.1 c.cm. + 9.9 c.cm. NaCl solution. = 0.2 c.cm. + 19.8 c.cm. NaCl solution. = 0.3 c.cm. + 29.7 c.cm. NaCl sol. = 1 c.cm. of dil. 1 : 10 + 9 c.cm. of NaCl sol. = 2 c.cm. of dil. 1 : 10 + 18 c.cm. NaCl sol., etc.	1 c.cm. + 999 NaCl = 0.1 c.cm. + 99.9 c.cm. NaCl = 0.2 c.cm. + 199.8 c.cm. NaCl sol. = 2 c.cm. + 1998 c.cm. NaCl sol. = 1 c.cm. of dilution 1 : 100 + 9 c.cm. NaCl sol. = 0.1 c.cm. dil. 1 : 10 + 9.9 c.cm. NaCl sol. = 0.1 c.cm. of dil. 1 : 100 + 0.9 c.cm. NaCl solution.	0.1 c.cm. + 999.9 c.cm. NaCl sol. = 1 c.cm. of dilution 1 : 1000 + 9 c.cm. of NaCl sol. = 0.1 c.cm. of dilution 1 : 100 + 9.9 c.cm. NaCl sol. = 1 c.cm. dil. 1 : 100 + 99 c.cm. NaCl sol. = 0.1 c.cm. dil. 1 : 10 + 99.9 c.cm. NaCl sol.

In preparing these dilutions, it is best to measure off the small quantities 0.1-1 c.cm. with a pipette; allow this to run into a well-graduated measuring glass and add enough saline to make the required dilutions. For example, if 30 c.cm. of a dilution 1 : 100 is desired, 0.3 c.cm. should be measured off with a pipette and allowed to flow into a 50 or 100 c.cm. graduated cylinder and saline solution added up to 30.0 c.cm.

It should always be one's aim to get along with small quantities of the substance to be diluted. If, for example, 8 to 10 c.cm. of a toxin dilution 1 : 100 are required 0.1 c.cm. of toxin + 9.9 c.cm. of saline should be taken and not 1 c.cm. of toxin and 99 c.cm. of NaCl sol.

Before making any dilution one should always calculate the total amount of substance required; as for example in the following experiment:

1. Animal 0.1 c.cm. Toxin subcutaneously
2. Animal 0.05 c.cm. Toxin subcutaneously
3. Animal 0.01 c.cm. Toxin subcutaneously
4. Animal 0.001 c.cm. Toxin subcutaneously

Here, the total quantity of toxin necessary is found, by adding, to be 0.161 c.cm. This represents the minimum amount. It is always advisable to make an allowance for some loss and at the same time bring up the amount to a round or even number. 0.2 c.cm. of toxin would fulfill all these requirements. This amount is measured off by a pipette, placed into a graduated cylinder and saline added up to 2.0 c.cm., making a dilution of 1 : 10. Then 0.2 c.cm. of this dilution (1 : 10) is taken, placed into another graduate, and again diluted with saline up to 2.0 c.cm. thus making a dilution of 1 : 100. The above problem therefore of injecting the various animals, can be completed as follows:

1. Animal receives 1 c.cm. of dilution 1 : 10
2. Animal receives 0.5 c.cm. of dilution 1 : 10
3. Animal receives 1 c.cm. of dilution 1 : 100
4. Animal receives 0.1 c.cm. of dilution 1 : 100



The unit for measuring the amount of bacteria grown upon a solid medium is represented by a standard sized loop. This platinum loop takes up about 2 mg. of bacterial substance. It is prepared as explained in Fig. 2. If smaller amounts of bacteria are used, dilutions must be made.

For instance,  $\frac{1}{4}$  of a loopful of bacteria is desired; 1 loopful is suspended in 1 c.cm. of saline and 3 c.cm. of saline added. As a result, 1 c.cm. of this emulsion contains  $\frac{1}{4}$  of a loopful of bacteria. If  $\frac{1}{16}$  of a loopful is necessary 1 c.cm. of the above dilution is added to 3 c.cm. of saline, thus making 1 c.cm. of this last mixture contain  $\frac{1}{16}$  of a loopful of bacteria.

## CHAPTER III.

### ACTIVE IMMUNIZATION.

#### IMMUNIZATION WITH LIVING AND DEAD VIRUS.

Active immunization depends upon the principle, that an organism in overcoming a slight infection, either naturally or artificially acquired, develops enough protective bodies to withstand a similar, severer, natural, or acquired infection. Moreover, it serves primarily the purpose of prophylaxis. In laboratories, active immunization of animals is also frequently undertaken with the view of obtaining sera for diagnostic and therapeutic purposes.

In the manufacture of large quantities of serum, the horse is the animal used almost exclusively. Occasionally cows, sheep, donkeys or mules are selected. In small laboratories usually rabbits, guinea-pigs, white mice, rats, and only occasionally goats or sheep are employed.

The process of immunization evokes a marked disturbance in the general health of the animals. For this reason they must be well kept in warm places, and well fed. As far as their power of producing antibodies is concerned, there are individual differences even among the same species of animals; thus if five horses are immunized against diphtheria, some will give much better curative sera than the others. In general, the younger animals are preferable.

Any substance which, when injected into an organism, can stimulate the production or formation of antibodies, has been conveniently termed "antigen." After the injection of such an antigen, special notice should be taken of the animal in reference to temperature, weight, the excitation of diarrhea or the occurrence of abscesses, infiltrates, edema or paralysis.

If an animal dies, a careful postmortem, and if possible, a bacteriological examination should be made. It should be the aim to ascertain if death was induced by the inoculated antigen, by contamination or secondary infection. One should always keep in mind the possibility of some of the animal epidemic diseases.

Epidemic diseases occurring in rabbits are:

Animal Infections. 1. RABBIT SEPSIS.—Presents itself in the form of bronchopneumonia and marked nasal catarrh. It is very infectious. Sick animals should at once be isolated or killed and their cages thoroughly disinfected.



2. COCCIDIOSIS gives changes in the liver due to the settling of the coccidi ova forms. The parasites are present in the pus and are easily recognized microscopically. Following labor, *guinea-pigs* are very susceptible to sepsis.

IN RATS, trypanosomiasis is of frequent existence, but is not pathogenic.

The antigens are injected either *subcutaneously*, *intraperitoneally* or *intravenously*. Only on exceptional occasions is another entrance path chosen.

The Tech- As regards the amount to be injected, one cannot very well  
nique of give general rules. It is important to prevent severe re-  
Active Im- actions, although the question is still a disputed one, whether  
munization. marked reactions tend to produce a better immunity. It is certain, how-  
ever, that inoculations of antigens in such minute doses as to apparently  
give no reaction, can still lead to immunity and the production of  
antibodies.

*Occasionally a single injection suffices for immunization. Repeated inoculations are usually necessary, especially so when a "highly valent" serum is desired, i.e., one containing a great number of antibodies or having high protective properties.*

*When repeated inoculations are undertaken, there are various methods of procedure.*

1a. A small dose of antigen is injected. If a reaction sets in, one waits until this reaction has entirely subsided, then (not before the fifth day) the second injection—a somewhat larger dose—is given. After an interval of 5 to 8 days, a third injection of a still higher dosage is administered, and so on, again.

1b. The intervals are the same, but the amounts of antigen remain the same at each injection.

Both of these methods give excellent results and therefore are most frequently used.

2. For several successive days, a small or medium dose of antigen is injected. Each injection produces only a slight reaction.

This last scheme according to Fornet is especially suitable for obtaining precipitation sera. As is evident, it has the advantage of gaining the immunity rapidly.

3. Inoculations are given at very long intervals (intermissions of four weeks or more). This method produces good sera, but has the disadvantage of requiring too long a time.

The methods of active immunization can also be divided according to the nature of the antigen.

1. Immunization with a living virus,
2. Immunization with a dead virus,
3. Immunization with bacterial extracts,
4. Immunization with bacterial toxins.



## I. Immunization with a Living Virus.

This method of immunization simulates most closely the immunity attained spontaneously in overcoming an infection. This immunity is very strong and lasts for a long period of time, but it is attained with difficulty; frequently the dose of virus injected causes serious symptoms of infection. Various procedures have therefore been advocated so to diminish the toxicity of the immunizing agent that only immunization effects, and no toxic symptoms be obtained. This was attempted either by the reduction of the number of organisms employed, so that very minute doses were inoculated, or by the diminution of the infectious nature of these bacteria (virulence so called).

The first method, however, was not found applicable to all cases. The infectious nature of the different bacteria varies markedly. The same bacterium reacts differently with different animals. While some animals possess a natural immunity against certain bacteria, others exhibit a distinct susceptibility to the same micro-organisms. The conceptions therefore of *pathogenicity* and *virulence* are purely of a relative nature. In talking of the pathogenicity of bacteria, one should always mention the class of animal for which these bacteria are pathogenic.

Bail has used this principle of pathogenicity in classifying bacteria. He Bail's Classi- mentions the following three classes:

fication of a. Saprophytes.

Bacteria. b. Half or partial parasites.

c. Whole or pure parasites.

To the class of *saprophytes* belong all those bacteria which when injected even in larger doses do not produce any characteristic disease; these are also known as *apathogenic*—e.g., *hen cholera bacilli* for human beings.

Classed as *half parasites* are those bacteria, according to Bail, the infectious nature of which depends upon the quantity of bacteria injected. While the injection of a rabbit with 1/1000 of a loopful of a typhoid culture will produce no evidences of disease, one-tenth of a loopful will result in slight increase in temperature, loss of appetite, and eventually a local redness at the site of the injection. One loopful may bring about the death of the animal. The manifestations are dependent entirely upon the number of bacteria injected. The smaller the number, the milder the symptoms, until one reaches the stage below which no disturbances at all are visible.

*Pure parasites* are those which have no sublethal dose. Even the smallest amount, when injected, will produce death. As examples, the tubercle bacillus for guinea-pigs, and bacilli belonging to the group of Hemorrhagic Septicemia for rabbits. Of the last mentioned 1/10,000,000,000 of a loopful of some cultures kills a rabbit within twenty-four hours with the symptoms of a septicemia; in other words, the injection of 1 c.cm. of a dilution of one loopful of culture in ten million liters of water suffices to kill the rabbit. Furthermore, the bacteria increase so greatly in the body of the rabbit that they can be demonstrated in every drop of blood and in all organs and body fluids.

The same organism is a saprophyte for the human being and a half parasite for the guinea-pig if injected subcutaneously and a complete parasite by intraperitoneal injection.



tion. *The conceptions therefore of complete or partial parasite as well as of saprophyte are only relative and are dependent upon the bacteria, the animal species, and the mode of infection.*

It is now clear that immunization with living bacteria can only be undertaken if the latter belong to the class of half-parasites. Pure parasites are excluded from this method. As an example of such procedure can be given the immunization of a guinea-pig by intraperitoneal injections with living typhoid bacilli. Preliminary to this, the virulence of the typhoid culture must be ascertained.

*a. Preliminary test to titrate the virulence of the typhoid culture.*

1. Guinea-pig 1./I. 1909 1/20 loopful of typhoid culture intraperitoneal.
  - 2./I. active.
  - 8./I. alive.
2. Guinea-pig 1./I. 1909 1/10 loopful of typhoid culture intraperitoneal.
  - 2./I. active.
  - 8./I. alive.
3. Guinea-pig 1./I. 1909 1/8 loopful of typhoid culture intraperitoneal.
  - 2./I. slightly sick, does not eat.
  - 3./I. active.
  - 8./I. alive.
4. Guinea-pig 1./I. 1909 1/6 loopful of typhoid culture intraperitoneal.
  - 2./I. sick, does not eat, hair raised.
  - 3./I. still sick.
  - 4./I. more active.
  - 8./I. alive.
5. Guinea-pig 1./I. 1909 1/5 loopful of typhoid culture intraperitoneal.
  - 2./I. sick, does not eat, hair raised.
  - 3./I. very weak, when placed on side remains so.
  - 4./I. †
6. Guinea-pig 1./I. 1909 1/4 loopful of typhoid culture intraperitoneal.
  - 2./I. †

From this experiment it becomes evident that the lethal dose of this particular strain of typhoid culture is 1/4 to 1/5 of a loopful for guinea-pigs by intraperitoneal injection. Immunization therefore must be started with a smaller dose—e.g., 1/10 of a loopful.

*b. Immunization.*

1. Guinea-pig 8./I. 1909 1/10 loopful of typhoid culture intraperitoneal.
  - 16./I. 1/8 loopful of typhoid culture intraperitoneal.
  - 22./I. 1/4 loopful of typhoid culture intraperitoneal.
 Animal remains active and healthy.
  - 30./I. 1 loopful of typhoid culture intraperitoneal.
  - 5./II. 2 loopfuls of typhoid culture intraperitoneal.
 Animal remains active and healthy.
2. Guinea-pig 8./I. 1909 1/10 loopful of typhoid culture intraperitoneal.
  - 16./I. 1/4 loopful of typhoid culture intraperitoneal.
 Animal remains active and healthy.

3. Guinea-pig 8./I. 1909 1/10 loopful of typhoid culture intraperitoneal.  
 16./I. 2 loopfuls of typhoid culture intraperitoneal.  
 17./I. animal is sick and does not eat.  
 18./I. animal is very weak.  
 19./I. †

Control animals always die within twenty-four hours, as in previous experiment, on injection of 1/4 of a loopful.

From experiment with guinea-pig 1, it can be learned, that by gradual increase of the immunizing dose, a state of immunity is reached which can overcome an infection produced by a high multiple of the dosis letalis.

Experiments 2 and 3 prove that even a single preliminary injection suffices to prevent the death of an animal upon subsequent receipt of the lethal dose of the same bacteria; but this single inoculation is not sufficient to protect the organism against a very severe future infection. The attained immunity is therefore only relative, not absolute.

Analogously it is possible to immunize by subcutaneous and intravenous injections. The latter method is usually the one of choice when half parasites are employed, as the highest and quickest grade of immunity is thus reached. It carries with it, however, the greatest danger, and frequently results in death of the animal.

The method of immunization with small doses of living, fully virulent bacteria, has thus far been made use of only in animals. *In man this experience has not been carried into effect. It is feared that the bacteria may increase very rapidly and give rise to severe disturbances.* The method has therefore been altered and *instead of using virulent material for immunization, only a weakly infectious or attenuated virus is employed.*

### Vaccination against Small-pox.

This is the best known example of active prophylactic immunization. To Jenner belongs the credit of having been the first one to apply this principle. Vaccine. Vaccination against small-pox consists in inoculation of an attenuated form of small-pox germs, the diminution in virulence being brought about by passage through the body of a calf, a less susceptible animal than man. The vesicles formed on the vaccinated person contain these attenuated germs. This lymph can be used for the inoculation of other individuals, as the germs do not regain their virulence by repassage through man.

Inasmuch as it is not within the scope of this book to go into the details of the preparation of the lymph or the technique of vaccination, a brief survey of the benefits of vaccination will amply suffice and this may be seen from the table hereunto appended.

The mortality from small-pox per 100,000 population was in the years



	1862-1876	1882-1896
in Prussia and Bavaria .....	51.6	0.7
in Austria .....	75.2	38.6
in Belgium .....	79.5	18.2
in England .....	25.3	2.9
in Sweden .....	26.9	0.5

This method of immunizing against a virulent virus by inoculating with an attenuated form of the same, is known as *Jennerization*. *Pasteur recognized that this method had general application and similarly used attenuated but still living cultures, "vaccins" so called, to immunize against hen cholera, swine plague, and anthrax.* The same principle underlies Pasteur's anti-rabic vaccination.

### Antirabic Vaccination.

In all civilized countries there exist special institutions, either directly under the city control or appointed by the city, where the Pasteur treatment for rabies is conducted. It is the duty of the general practitioner, on getting a suspicious case of rabies, to advise his patient to undergo this special therapy and to send the rabid animal, its head or brain preserved in glycerin, to the institute as soon as possible for the purpose of ascertaining the presence of rabies. Up to very recently the actual cause of hydrophobia was unknown. Negri had described parasites, known as Negri bodies, in the large nerve cells of the cerebral cortex, cerebellum, etc.<sup>1</sup> In man, infection usually occurs as a consequence of the saliva of rabid animals (dog, cat, wolf, skunk) gaining entrance to wounds from bites or scratches. Roux and Nocard found that the saliva of experimentally infected animals is already infectious 2-3 days before the first symptoms appear. Thus, rabies may be transmitted to an individual by an animal apparently healthy at the time. Remlinger has formulated the following very instructive outline referring to the indications for antirabic treatment.

Principle of Rabies Treatment.	as Negri bodies, in the large nerve cells of the cerebral cortex, cerebellum, etc. <sup>1</sup> In man, infection usually occurs as a consequence of the saliva of rabid animals (dog, cat, wolf, skunk) gaining entrance to wounds from bites or scratches. Roux and Nocard found that the saliva of experimentally infected animals is already infectious 2-3 days before the first symptoms appear. Thus, rabies may be transmitted to an individual by an animal apparently healthy at the time. Remlinger has formulated the following very instructive outline referring to the indications for antirabic treatment.		
If the biting animal	1. has died within 10 days after the biting,	}	antirabic treatment is indicated.
	2. has been killed within 10 days after the biting,		
	3. has disappeared within 10 days after the biting,		
	4. is unknown to the individual bitten,		
	(a) becomes ill with rabies,	}	antirabic treatment is indicated.
	(b) dies with suspicious symptoms of rabies or of another disease,		
	5. has remained alive and under observation for 10 days,	}	further observation, treatment if animal dies.
	(c) becomes ill but does not die within 10 days,		
	(d) remains well both during and after this period,	}	no treatment.

<sup>1</sup> Noguchi has now succeeded in artificially cultivating the rabic virus. By inoculating cultures containing these granular, pleomorphic or nucleated bodies, he has reproduced rabies in dogs, rabbits and guinea-pigs.

Pasteur found that rabies can be transmitted to dogs by injecting them subdurally with the brain substance of rabid animals. This ordinary virus substance is known as Street Virus.

The incubation period of rabies is very long. It varies from about three weeks to [possibly] some years. By passing the virus through monkeys, the incubation period is considerably increased. After the successive passage through five or six animals, the virus becomes so weakened that infection is almost impossible. Reversely, increase of the virulence may be affected by passing the virus through a successive number of rabbits, as these are very sensitive to the disease. After passage through a large number of such animals, the incubation period is gradually shortened from about three weeks or a little less to a constant period of six or seven days. Further diminution in the period of incubation was impossible and therefore Pasteur called this "*Virus fixe*." His first experiments in immunization were made by passing the weakened monkey virus through rabbits and then treating dogs with the spinal cords of the latter.

Later on, Pasteur discovered that instead of passing the virus through monkeys, he could *diminish its virulence by drying the spinal cords* derived from rabid animals, for varying periods of time. In this way he could prepare an entire series of graduated strengths. The material used for this drying was not the street virus, but that obtained by successive passage through rabbits or "*virus fixe*" which possessed very constant immunizing and infectious properties. By drying the "*virus fixe*" over caustic potash at a temperature of  $23^{\circ}$  to  $25^{\circ}$  C. for five days, its regular incubation period of 7 days was very much prolonged. Increase in the length of drying caused the entire loss of virulence.

Pasteur immunized dogs as follows: He began with the injection of a virulent spinal cord which had been dried for thirteen days and every following day injected subcutaneously some fresher spinal cord, *i.e.* (dried for a lesser period of time), until finally he used virus dried only for one day. The animals thus treated were immune against the bites of rabid dogs as well as subdural, subcutaneous, and intravenous infection with "*virus fixe*" and street virus. This procedure was strongly recommended by Pasteur, who brilliantly contributed the observation, that if an animal was infected but did not as yet show symptoms, these could be prevented by a similar *modus operandi*, as above mentioned.

In man, the inoculation is carried out on the same principle. The fact that the incubation period of hydrophobia is very long, makes the prophylactic inoculations of greater service. Only rarely is this period less than six weeks, usually considerably longer—up to 584 days, entirely dependent upon the virulence of the virus and the point of infection.



### Technique of Antirabic Vaccination in Man.

The actual vaccine consists of 1 c.cm. (2-3 mm. length) of the substance of the spinal cord of a rabbit which has been killed by inoculation with the fixed virus, rubbed up into a fine emulsion with 5 c.cm. of sterile 0.85 NaCl solution. About 1 to 3 c.cm. of the resulting fluid are injected subcutaneously into the skin of the abdomen. A cord dried for fourteen days is used for the first injection, emulsions of less attenuated virus are used on succeeding occasions until finally a portion of a spinal cord dried for only three or four days is employed. Pasteur's schemes of the actual doses can thus be drawn up.

#### a. For infections at points distant from the central nervous system (*mild infections*).

Day of injection.....	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15
Number of days cord was dried...	14 + 13	12 + 11	10 + 9	8 + 7	6 + 6	5	5	4	3	5	5	4	4	3	3
Amount injected in cubic centimeters.	3.0	3.0	3.0	3.0	2.0	1.0	1.0	1.0	1.0	2.0	2.0	2.0	2.0	2.0	2.0

#### b. For head wounds (*severer infections*).

Day of injection.	1		2		3		4	5	6	7	8	9
Number of days cord was dried.	14 + 13 In A. M.	12 + 11 In P. M.	10 + 9 In A. M.	8 + 7 In P. M.	6+6		5	5	4	3	4	3
Amount injected in cubic centimeters.	3.0 A. M.	3.0 P. M.	3.0 A. M.	3.0 P. M.	2 c.cm. A. M.	2 c.cm. P. M.	2.	2.	2.	1.	2.	1.

Day of injection..... 10 11 12 13 14 15 16 17 18 19 20 21

Number of days cord was dried. 5 5 4 4 3 3 5 4 3 5 4 3

Amount injected..... 2. 2. 2. 2. 2. 2. 2. 2. 2. 2. 2. 2.





In severe injuries the entire treatment is repeated after one month's interval.

There is at present no doubt whatsoever as to the value of these antirabic vaccinations. Compiled from a great number of statistics, the mortality of those infected or exposed to infection but untreated is 15 to 16 per cent., while the death rate of those treated at the Berlin Institute during 1898 to 1901 was 0.55 per cent. Similar figures are given by the other institutions.

The serum of individuals who have taken the Pasteur treatment contains antibodies that can neutralize the toxic effects of the rabies virus. If a virulent strain of the latter is mixed with the serum and injected into an animal, no symptoms will develop. A similar serum is manufactured at the Pasteur Institute by the intravenous injection of sheep with emulsions of the street virus or virus fixe. It is used mainly for immunization of animals by the so-called "simultaneous method," whereby mixtures of a virulent virus and the serum are employed. In this way an immunity is attained much more rapidly than by the classical method. A. Marie and Remlinger have made use of this simultaneous method also in man, with good results. It is of value especially when a rapid immunity is essential as in severe infections (bite of wolf, injuries of the face) or neglected cases. The time elapsed between the injury and the onset of treatment is an important deciding factor as to the final result. The following table of Diatrottoff demonstrates this:

The bitten individual started treatment during the	Number of patients treated	Of these died	Percentage
First week.....	4602	26	0.56
Second week.....	991	16	1.66
Third week.....	313	10	3.19

The immunity attained by vaccination is of comparatively short duration; probably only several years. It is advisable therefore in case of reinfection not to depend upon the previous immunity but go through another treatment.

Attempts have been made to employ this principle of virus attenuation for other infections. Behring and Koch tried immunization against bovine tuberculosis by inoculation with living human tubercle bacilli. These can be bought under the name of Beovovaccine (v. Behring) and Tauruman (Koch).

Tauruman is prepared by the Höchst Farbwerke and is put up in sealed glass tubes which contain 0.02 to 0.04 gm. of living tubercle bacilli suspended in 10 c.cm. of normal saline solution. This Tauruman is previously examined in Ehrlich's Institute and note is taken of its purity, quantity of bacteria, virulence against guinea-pigs and avirulence against rabbits (characteristics of the human type of tubercle bacilli).

To this class of experimental work belong also the attempts of Friedmann to immunize against human tuberculosis by the use of the tubercle bacilli of cold-blooded animals, and those of Wassermann, Ostertag and the author, to inoculate against hog cholera with living cultures of mouse typhoid.

Besides the preceding mode of virus attenuation by passage through animals, there are other methods employed for the diminution of the toxicity of the virus. Growing the bacteria at too high a temperature, or exposing bacterial emulsions to light, disinfectants or moderate heating, accomplishes the same purpose.

*The mixture of bacteria with their specific serum* (i.e., serum obtained from animals that have been inoculated with these bacteria), also diminishes the virulence of the bacteria. Such bacteria are designated by Bordet as "sensitized." In this mixture, the bacteria attach their specific antibodies; after centrifugalization, the added specific serum now devoid of its specific antibodies is removed, and the sensitized bacteria can be used as vaccines. Inoculations of the latter rarely produce any infiltration. The same object can be accomplished by injecting bacteria and at the same time also their specific serum. This is technically simpler and is known as the "*Simultaneous Method*." It has shown itself of great value in Lorenze's prophylactic inoculations against swine erysipelas.

**2. Immunization with Dead Bacteria.**—Immunization with dead bacteria was first undertaken by Toussaint, Salmon and Smith, and Chamberland and Roux.

This method is to be distinctly separated from those already discussed. Bail claims that the immunization with living bacteria as well as by aggressins (to be mentioned later) is an immunization against the infectious disease; while the immunization with dead bacteria is an immunization against the bacterial bodies. While this holds true for some bacteria, it is, to say the least, questionable whether it can be considered a general rule.

Whenever a real immunity is desired—that is, protection against disease, a vaccine either in the form of living or attenuated bacteria should be given the preference. Up to a certain degree the extracts of living bacteria, and the natural and artificial aggressins can be similarly employed. If, however, no real immunity, but just a serum containing a great number of antibodies is wanted, as in serum diagnosis, for agglutination, bacteriolysis, complement fixation, etc., then immunization by dead bacteria is just as, if not more so, efficient.

Recently, the question has been raised whether the antibodies produced by immunization with heated antigens are identical with those obtained with unheated antigens. The experiments of Obermeyer and Pick,



which will be referred to under proteid immunization, seem to prove that they are not alike. For laboratory work it is advisable to use living cultures only in cases of absolute necessity.

In heating bacteria to destroy their virulence and thus be suitable for inoculation, we must be very careful not to raise the temperature to such a degree that not only the toxicity but also the immunization power is destroyed. It is best to employ the minimum amount of heat which will kill the respective bacteria.

For most of these as Typhoid, Paratyphoid, Colon, and Dysentery bacilli, Cholera Vibrios, Meningo-, Staphylo-, Strepto- and Pneumococci, one hour at 60° C. is sufficient.

The bacteria are grown upon agar cultures and the required amount is removed and suspended in sterile physiological salt solution or bouillon. This suspension is then placed into a hot water bath or thermostat regulated at 60° C., for one hour.

If the bacteria employed are highly infectious, one must be sure that all bacteria have been killed. This must especially be noted when giving prophylactic inoculations in man. Several drops of the emulsion are therefore transferred to agar tubes and incubated for a day or two. If a growth appears, the emulsion is to be reheated; if not it can be considered sterile.

The mode of immunization with dead bacteria is the same as has been described for the living ones. In general the dosage to be used may be larger.

Small doses are injected at first, followed later on by increasing quantities at intervals of five to eight days, *e.g.*

### **Intravenous inoculation of a rabbit with dead typhoid bacilli.**

*Result.*—Protection against living virulent bacteria, appearance of agglutinins, bacteriolysins, bacteriotropins and complement binding substances in the serum.

1./I. 1909.	Rabbit No. 1.	1 loopful of a typhoid agar slant culture killed at 60° C. and injected intravenously.
6./I.		4 loopfuls of typhoid culture killed at 60° C. and injected intravenously.
12./I.		1 culture of typhoid killed at 60° C. and injected intravenously.
20./I.		Infection with 1 culture of the living typhoid bacilli injected intravenously. Animal remains alive.
	Rabbit No. 2.	Control.
20./I.		Infection: 1/4 loopful of living typhoid bacteria intravenously.
22./I.		†(death).

The use of killed typhoid bacteria for prophylactic immunization has recently been widely adopted. This has been stimulated to a great degree by the successful experiments of Wright, and Pfeiffer and Kolle.

### Wright's Method of Prophylactic Typhoid Inoculation.

The vaccine originally employed by Wright for these inoculations consisted of highly virulent cultures of *Bacillus Typhosus* grown in broth for twenty-four to forty-eight hours (sometimes even for four weeks), and sterilized by heating at 60° C. The vaccine was then standardized, *i.e.*, the strength of the vaccine was fixed in accordance with another of known strength, the dosage of which had been gauged by inoculations in man.

The early form of standardization consisted in determining the toxicity of the virus. Guinea-pigs weighing 250 to 300 gms. were inoculated subcutaneously with 0.5, 0.75, 1.0 and 1.5 c.cm. of the vaccine respectively. Death to some of the animals would come in twelve hours to three days. The amount required to kill a guinea-pig weighing 100 gms. or rather the proportional fraction of the dose which proved fatal to the one of 250 to 300 gms. was taken as the standard dose for injection in man. Wright subsequently found that better results were obtained, if the vaccine was prepared from twenty-four hour cultures grown upon the surface of agar. The growth is then washed off in physiological

Preparation. saline solution. This emulsion is sterilized by subjecting it to a temperature of 55° to 60° C. for one hour, after which the number of bacteria is computed by Wright's method (see Standardization of Vaccines in chapter on Opsonins). After this the emulsion is further diluted with physiological salt solution containing 0.5 per cent. carbolic or 0.25 per cent. trikresol in such a manner that two or three different concentrations are secured, one containing 500 million killed typhoid bacilli per c.cm., one containing 1000 million per c.cm., and one containing 2000 million per c.cm.

The particular strain of typhoid bacillus employed for the vaccine varies. Some use a strain isolated by Leishman in 1900. This is selected not on account of its degree of virulence but on account of its property of being able to stimulate the formation of a great amount of antibodies. The editor and others employ a mixture of eight or ten different strains with perhaps one or two paratyphoid strains.

The typhoid vaccine or typho-bacterin as it is frequently called is administered subcutaneously, usually in the arm at the insertion of the deltoid muscle. The needle should not enter the muscle or find its way between the layers of the skin. The arm should be cleansed as for any other injection with alcohol and iodine.

*The dosage* almost uniformly employed consists of 500 million bacteria for the first injection and 1000 million for each of the two subsequent injections at intervals of eight to ten days. If, however, such an extended period of time is not available, then two inoculations will suffice, the first dose 1000 million and the second dose



2000 million. Wright used the two larger doses. An objection raised to this method is that the general reaction obtained is more severe. The editor, however, has employed these larger doses for inoculating the nursing and medical staff of the hospital and a great number of laymen, without any ill effect.

Women and children should receive a dose in proportion to their weight; a healthy man weighing 150 pounds being designated as the standard of comparison.

*Local and general reactions* follow the inoculations. Thus local redness and swelling of the skin, lymphangitis and enlargement of the neighboring glands are the usual consequences. The inflammation can at times be severe enough to simulate erysipelas. The general symptoms, on the other hand, may consist of a general feeling of illness, headache, little fever, and occasionally nausea, not infrequently accompanied by vomiting. These signs of indisposition, however, pass off rapidly without leaving any permanent ill effects. Debilitated persons frequently present the most profound reactions. Occasionally latent and chronic diseases of a non-typhoidal character may be made active by inoculation. These exacerbations are not serious and usually by diminishing the quantity and increasing the number of doses, these effects can be avoided. Six to eleven days after the injection, an increase in the number of agglutinating, bacteriolytic and bacteriotropic bodies can be demonstrated in the blood of the inoculated individual. The immune bodies reach their height in two to three months after inoculation, and then fall rapidly. In a series of forty cases Garbat was unable to demonstrate complement fixation bodies with any regularity in the blood after prophylactic injection.

As to the results of antityphoid vaccination, opinion is somewhat divided. According to Wright's statistics infections have been diminished by about one-half, and in single series to one-sixth or even one-twenty-eighth of the former, or control number. The mortality too is much lower. Out of 1758 individuals who had been vaccinated, only 142 or 8 per cent. died; out of 10,980 who had not been, 1800 or 16.6 per cent. met death.

The immunity attained is not absolute, for according to Russel, in 1911, among 80,000 persons vaccinated in the United States Army there were twelve cases of typhoid with one death (due to intestinal hemorrhage), and in 1910 six cases occurred with no fatalities. Had it not been for the prophylactic immunization, there would have occurred at the prevailing rates of incidence about 250 cases. Similar favorable statistics have been collected in England, Germany and France. The period of immunity lasts from two to three years.

### Pfeiffer-Kolle's Experiments.

Pfeiffer and Kolle prepare their vaccine by growing typhoid bacilli on agar cultures and suspending a twenty-four hours' growth in physiological NaCl solution. The normal platinum loop is the unit of standardization. A full grown agar culture is considered as 10 normal loops and as such it is diluted in 4.5 c.cm. of saline. This emulsion is placed in a thermostat at 60° C. for two hours and then tested for its sterility. Sufficient 5 per cent. phenol solution is next added to the suspension to bring the contents up to a 0.5 per cent. carbolic solution, and the final emulsion is again heated at 60° C. for thirty minutes. One c.cm. of the vaccine is thus equivalent to two normal loops of culture. The amounts of vaccine to be injected have not yet been definitely decided upon. The best dosage so far is the following:

For the first injection: 0.3 c.cm. of the vaccine.

For the second injection: 0.8 c.cm. of the vaccine.

For the third injection: 1.0 c.cm. of the vaccine.

The injection is made subcutaneously between the breast and clavicle.

The local and general reactions are the same as those observed with Wright's method. As a result of the injection only increased agglutinins and bacteriolysins have been found in the blood serum. Bacteriotropins have not as yet been examined for.

The effects of these inoculations seem to be very good. Protection is prolonged according to the increase in the number of injections, and if inoculated individuals do become infected, they run a very much milder course of the disease.

The following statistics as given by Kuhn indicate the results:

	<i>Inoculated.</i>	<i>Non-inoculated.</i>
Very slightly ill.....	186 (50.13 per cent.)	331 (36.55 per cent.)
Moderately ill.....	96 (25.88 per cent.)	225 (24.85 per cent.)
Severely ill.....	65 (17.52 per cent.)	234 (25.80 per cent.)
Deaths.....	24 ( 6.47 per cent.)	116 (12.80 per cent.)
	<hr/>	<hr/>
	371 (100 per cent.)	906 (100 per cent.)

The prophylactic immunity according to Kuhn lasts one year. Kolle has undertaken similar experiments against cholera.



## CHAPTER IV.

### ACTIVE IMMUNIZATION.

#### **Immunization with Bacterial Extracts.—Aggressin Experiments.**

The marked infectious nature of the organisms belonging to the class of "pure parasites" makes it very difficult to produce an immunity against them. They possess no sublethal dose in their living state, and if used when dead, will produce no prophylactic immunity. By artificial attenuation of these living virulent bacteria Pasteur succeeded in obtaining vaccines of several of them. The methods that he employed were, however, totally impracticable, for not infrequently, by the use of the vaccine, the disease which it was the object to prevent was instigated. It was therefore a distinct and important triumph when Bail and Weil showed that immunity against these parasites could be attained by using as vaccine antigen, the so-called "*aggressins*"; *i.e.*, exudates from animals that had been infected with the respective bacteria.

Bail's explanation of the aggressin-immunization method is entirely theoretical. He believes that during an infection, the bacteria secrete certain agents which counteract or entirely destroy the infected organism's protective powers, especially phagocytosis. These bodies he called aggressins and they were distinguished by the fact that they were formed by living bacteria, and only in the living body. According to Bail, the pathogenicity of bacteria depends upon their power to produce these aggressins. If this theory be correct, it should be possible to demonstrate aggressins, especially in infections where the protective power of the organism is almost nil, as for example an infection produced by the bacteria belonging to the group of hemorrhagic septicemia. Unfortunately, in actual practice this is not so.

The following experiment gives an idea of the true nature of these aggressins and how they are obtained.

At first, an infecting agent—the bacillus of swine pest, may be chosen. This micro-organism belongs to the same class as chicken cholera and fowl plague, and is distantly related to the human pest. For rabbits, this bacillus is a pure parasite, for guinea-pigs, by subcutaneous inoculation, a half parasite.

#### **The Obtaining of Aggressins.**

One drop of a twenty-four-hour broth culture of this swine pest bacillus, in 5 c.cm. bouillon, is injected intrapleurally in a rabbit in the following manner.

A small incision is made in one of the intercostal spaces on the side of the chest, and through this wound a long canula is introduced into the pleural cavity. The animal as a rule rapidly succumbs to the infection. On autopsy, the pleural cavity is found to contain an exudate of a reddish-brown color (hemorrhagic) on the side where the inoculation was given, and of yellow serous fluid on the other side. The bloody exudate, about 15 c.cm., is removed with a sterile pipette, placed in a sterile centrifuge tube to which is added 1.5 c.cm. of 5 per cent. carbolic acid drop by drop (making the entire solution a 1-2 per cent. carbolic acid dilution), agitated continually in order to prevent precipitation, and followed by centrifugalization at a high speed for many hours until it becomes very clear. The upper clear part which is now free of bacteria, or very nearly so, is pipetted off and heated for three hours at 44° C. Its sterility is then tested and if no growth appears after forty-eight hours, it is considered sterile.

### First Fundamental Aggressin Test.

*(Its Power of Increasing Severity of Infections.)*

No.	Animal	Date	Amount of infective material	Aggressins	Result
1	Guinea-pig.	6/IV'05.	1/100 loopful of swine pest subcutaneously.	.....	Remains alive.
2	Guinea-pig.	6/IV'05.	1/100 loopful of swine pest subcutaneously.	+ 1.5 c.cm. of aggressins subcutaneously.	† on third day.
3	Guinea-pig.	6/IV'05.	.....	+ 1.5 c.cm. of aggressins subcutaneously.	Remains alive.
4	Guinea-pig.	6/IV'05.	1/100 loopful of swine pest subcutaneously.	+ 3 c.cm. subcutaneously.	† on second day.
5	Guinea-pig.	6/IV'05.	.....	3 c.cm. subcutaneously.	Remains alive.
6	Guinea-pig.	6/IV'05.	1/1000 loopful subcutaneously.	+ 2 c.cm. subcutaneously.	On 7/IV very ill; on 8/IV very ill; 9/IV very ill; 10/IV very thin; 11/IV begins to pick up slowly and remains alive. Marked infiltration around point of injection.



It can be deduced from this experiment that 1/100 of a loopful of swine pest culture, which represents 1/10 of a fatal dose for a guinea-pig by subcutaneous injection, can be converted into an acutely fatal dose by injecting the aggressin simultaneously or a half hour before the swine pest culture.

The aggressin itself is only *slightly toxic*, and the quantity injected is well borne by the guinea-pig. Its power of increasing the virulence of the infective material varies directly with its quantity, *i.e.*, the greater the dose of aggressin, the more rapidly is death occasioned. If, however, only small doses of the culture are given, and in addition to this, the aggressin is injected, the animal does not die, but becomes exceedingly ill, thus indicating the effect of aggressins. In this connection it might be well to add that the aggressin may be given twenty-four hours previous to the time of infection.

On microscopical examination of the aggressin exudate, only very few cells, but a great number of bacteria are present. The bacteria here have increased during the short time after the infection to a far greater extent than they would have done in an artificial medium. The body, continually in combat against their increasing toxicity, finds itself powerless when its limited fighting capacity, decreasing in proportion to the rise in strength of the hostile micro-organisms, is expended; and ultimately succumbs to the infection. During the struggle between the protective forces of the organism and the invading bacteria, many of the latter are destroyed and these disintegrated bacteria are found within the exudate. From this fact Wassermann and Citron formed the conclusion that the aggressins are not as Bail claimed, secretory products of live bacteria produced during the conflict between the bacteria and the body organism, but rather the products of broken down bacteria. Therefore, Bail's supposition that aggressins are only obtained in the living body is erroneous and can be shown to be so by the fact that aggressins may be reproduced whenever the essential requirements can be had, and these are:

1. Large numbers of bacteria.
2. Non-poisonous agents which can disintegrate these bacteria.

Aggressins thus obtained are known according to Wassermann and Citron, as "artificial" in contrast to Bail's "natural" ones.

### Wassermann and Citron Method of Obtaining Artificial Aggressins.

Cultures are grown in mass on Kolle's flask-plates. A Kolle's agar plate is equivalent to twelve agar slants. For the inoculation of these flasks a long platinum loop is needed which transfers some of the culture to the plate. The transferred material is then spread over the entire surface of the flask by a large triangular platinum loop. The latter is made by inserting into a holder both ends of a not too thin platinum wire, about 20

cm. in length which is then shaped into a triangular form. While still red hot, this triangular loop should be introduced in the flask and allowed to cool there. Before the culture is spread, it is advisable to bend the entire loop to a slight angle by pressing it against the upper wall of the flask, thereby preventing the hot end of the loop holder from coming in contact with the agar surface. It is best also to test the platinum loop upon the surface of the agar in order to ascertain whether it is still too hot.

After twenty-four hours of incubation there is usually a pronounced growth upon the plates. This culture is then washed off either by serum or distilled water ("serous" or "aqueous aggressin"). The former may be obtained fresh from a rabbit. Usually 10 to 12 c.cm. of fluid per flask is required; 3 or 4 c.cm. are first poured upon the culture growth and the mass scraped gently but quickly with the triangular loop. Then the remainder of the fluid 7 to 8 c.cm. is poured in to release the still adherent bacteria. The turbid milky emulsion is collected either in a small dark glass Erlenmeyer flask or a brown bottle. This is then placed into a proper apparatus and shaken for one to two days at room temperature. Enough 5 per cent. carbolic acid is added to make a 1/2 per cent. phenol solution, and the emulsion is centrifugalized and sterilized in the same manner as has been described for the natural aggressins.

The tendency of aggressins toward increasing virulence ("infektions beförderung") is the same whether these aggressins are artificial or natural.

From the following experiment it can be seen that the bacteria contain some substance which is easily soluble in the body fluids and in distilled water, and which increases the infectious nature of the respective bacteria when injected simultaneously with them. In small doses, this substance is not poisonous, in large doses it may be, but is not necessarily so. There is no definite relation between the poisonous *qualities of the aggressin and its power to increase the virulence of an infection*. This disproves the assumption of some authors that the action of the aggressins is dependent upon the toxicity of the endotoxins.



## Experiment.

No.	Animal	Date	Amount of infective material from agar culture	Serous aggressin	Watery aggressin	Results
1	Guinea-pig.	29/V '05.	1/200 loop of swine pest subcutaneously.	.....	.....	Remains alive.
2	Guinea-pig.	29/V '05.	1/200 loop of swine pest subcutaneously.	+ 2.5 c.cm. subcutaneously.	.....	† after twenty-four hours
3	Guinea-pig.	29/V '05.	.....	+ 2.5 c.cm. subcutaneously.	.....	Remains alive.
4	Guinea-pig.	2/VI '05.	1/200 loop of swine pest subcutaneously.	.....	2 c.cm. subcutaneously.	† after three days.
5	Guinea-pig.	2/VI '05.	1/200 loop of swine pest subcutaneously.	.....	3 c.cm. subcutaneously.	† after three days.
6	Guinea-pig.	2/VI '05.	.....	.....	3 c.cm. subcutaneously.	Remains alive.
7	Guinea-pig.	2/VI '05.	.....	.....	4.5 c.cm. subcutaneously.	† in twenty-four hours.
8	Guinea-pig.	2/VI '05.	1/200 loop of swine pest.	.....	.....	Remains alive.

## Second Fundamental Aggressin Test.

(*Its Property of Active Immunization.*)

Bail and his pupils believe that when bacteria invade a normal organism, it is the aggressin power of these bacteria which determines whether or not, by their multiplication, disease will set in. If infection does take place, it continues until the "aggressive" nature of the bacteria is curbed. As there are some bacteria which on injection do not produce any disease, Bail attributes this phenomenon of immunity to the missing "aggressive" action of the respective bacteria. It is not merely the presence of bacteria which is the criterion for the existence of disease; as long as they are void of their "aggressive" property, they have actually become saprophytes.

Accordingly, Bail believes that the bactericidal immunity is no true immunity because it can be obtained by injection of dead micro-organisms or by live bacteria in such minute doses that no specific symptoms are produced, *i.e.*, no aggressins are produced within the body. "If the immunity lacks the "anti-aggressive" component, which alone governs the existence of disease, one gains only an *apparent immunity* against the exciting factor of the disease, but not against the *disease itself*."

Bail places the utmost stress upon the difference *between an immunity directed against the exciting agent* of the disease (bactericidal immunity) and that against the disease itself (anti-aggressive immunity).

Immunization against the disease is only possible if the *aggressin* reaches the body of the animal to be immunized. This is possible either by employing Pasteur's method of vaccine inoculation, *i.e.*, the injection of bacteria, the "aggressive" nature of which has been weakened but not destroyed, or by direct inoculations of aggressins. The latter is by far the simpler and more reliable mode of procedure, being productive of a true immunity.

Nowhere does this problem appear of such extreme importance as where immunity against a pure parasite is contemplated, as in the case of swine pest and chicken cholera. While it is exceedingly difficult, in fact almost impossible to immunize against these bacteria either with dead or living germs or vaccines, this task is readily accomplished by the injection of non-poisonous aggressins, inasmuch as they are well tolerated. In addition, these bacteria are of help in definitely deciding whether or not an aggressin immunity is at all possible.

Weil, a co-worker of Bail's, has carried out these experiments for chicken cholera, while the author has done the same for swine pest.

### *Example of Active Immunization with Natural Aggressins.*

#### *a. Slow Immunization.*

##### Rabbit I.

- |          |      |  |
|----------|------|--|
| 6./IV.   | 1905 | 1st injection: 1.0 c.cm. natural swine pest aggressin intraperitoneally. |
| 17./IV.  |      | 2d injection: 1.0 c.cm. natural swine pest aggressin intraperitoneally.  |
| 25./IV.  |      | 3d injection: 1.0 c.cm. natural swine pest aggressin intraperitoneally.  |
| 1./V.    |      | 4th injection: 2.0 c.cm. natural swine pest aggressin subcutaneously.    |
| 12./V.   |      | 5th injection: 2.0 c.cm. natural swine pest aggressin subcutaneously.    |
| 16./VI.  |      | 1st infection; with 1/100 loopful of swine pest culture intravenously.   |
| 17./VI.  |      | Perfectly well.  |
| 8./VII.  |      | 2d infection; with 1 loopful of swine pest culture intravenously.        |
| 15./VII. |      | Perfectly well.  |
| 22./IX.  |      | 3d infection; with 1 loopful of swine pest culture intravenously.        |
| 3./X.    |      | Perfectly well.  |

##### Rabbit I.

##### Controls.

##### Rabbit II.

- |         |      |  |         |      |  |
|---------|------|--|---------|------|--|
| 16./VI. | 1905 | 1/100,000 loopful of swine pest culture intravenously. | 8./VII. | 1905 | 1/10,000 loopful of swine pest culture subcutaneously. |
| 17./VI. |      | † found dead.  | 9./VII. |      | † found dead.  |



*b. Rapid Immunization.*

## Rabbit II.

- 8./IV. 1905 Injection of 4 c.cm. of natural swine pest aggrassin subcutaneously.  
 10./IV. Animal shows slight ill effects.  
 13./IV. Perfectly active.  
 26./IV. 1st infection: 1/10 loopful of swine pest culture subcutaneously.  
 16./VI. 2d infection: 1 loopful of swine pest culture intravenously.  
 Animal remained active.

*Controls.*

## Rabbit III.

- 26./IV. 1905 1/10,000 loopful of swine pest culture subcutaneously.  
 27./IV. †.

These experiments prove conclusively that by the method described above it is possible to attain a high grade of immunity. In this connection, however, it is very important to adhere to what Bail pointed out, namely, *that a long period should elapse between the last inoculation with the aggrassin and the first infection; the reason for that being, that during the period of immunization, and following it for a long time, there is a condition of hypersusceptibility to infection.*

*Example of Active Immunization with Artificial Aggressins.*

## Rabbit 1.

- 3./VI. 1905 1st injection: 4 c.cm. of watery extract of swine pest bacilli subcutaneously.  
 14./VI. 2d injection: 2 c.cm. of watery extract of swine pest bacilli subcutaneously.  
 25./VI. Removal of some blood.  
 4./VII. 3d injection: 3 c.cm. of watery extract of swine pest bacilli subcutaneously.  
 21./VII. Infection: 1/10 loopful of swine pest culture subcutaneously.  
 3./X. Animal alive and healthy.

## Rabbit 2.

- 19./VI. 1905 1st injection: 2.5 c.cm. of serous extract of swine pest bacilli subcutaneously.  
 9./VII. 2d injection: 2.0 c.cm. of serous extract of swine pest bacilli subcutaneously.  
 12./VII. 3d injection: 4.0 c.cm. of serous extract of swine pest bacilli subcutaneously.  
 24./VII. 1st infection: 1/10 loopful of swine pest culture subcutaneously.  
 22./IX. 2d infection: 1 loopful of swine pest culture intravenously.

Animal remains perfectly well.

## Rabbit 3.

- 16./VI. 1905 Injection: 2.5 c.cm. of watery extract of swine pest bacilli subcutaneously.

- 4./VII. Infection: 1/100 loopful of swine pest culture subcutaneously.  
15./VII. Small local infiltrate. Very active.  
3./X. Animal alive and perfectly well.

Control animals inoculated on the days of infection died within twenty-four hours after inoculations of 1/100,000 loopful of culture intravenously, 1/10,000 loopful subcutaneously.

It is evident from the above, that an immunity against pure parasites can be obtained just as well by *one or several injections of extracts of living bacteria*, as by injections of natural aggressins. Since the production of aggressins by a struggle between the bacteria and distilled water can be excluded, it can be taken without further explanation that in the development of those substances which have a tendency to increase the virulence of bacteria, or which can be used to produce an immunity, the bacteria play a passive rôle, in that they are only extracted by the dissolving agent. The difference between the anti-bacterial and anti-aggressin immunity is therefore not a qualitative one, as in both instances they are the substances that are set free from the bacteria which stimulate the formation of antibodies. When *living virulent bacteria are injected for the purposes of immunization*, they increase so rapidly that a proper dosage is impossible and the animals frequently die before enough antibodies are liberated. In addition, antibodies are also generated against the capsule of the bacteria (bacteriolysins).

The only difference between immunization with *morphologically well preserved* but dead bacteria and that with aggressins is that within the latter the bacterial substances which tend to bring about the immunity have not been altered by previous heating, but exist in their natural easily absorbable form. Moreover, by using the extracts one does away with certain toxic substances which are found within the bacterial capsules, and which are rather toxic to subcutaneous tissue, producing necrosis and marasmus.

### The Third Fundamental Aggressin Experiment.

Here, it is demonstrated that the serum of animals immunized by aggressins either artificial or natural, contain antibodies which (1) can neutralize that property of aggressins whereby they increase the virulence of bacteria; (2) produce a passive immunity against infection with living bacteria.

As for the biological structure of these antibodies, or anti-aggressins as they may be called, it may be said that they belong to the class of amboceptors, shown by the complement fixation methods.

The practical employment of aggressins as a method of immunization offers distinct advantages, namely:

1. Absence of any possible dangerous effects.
2. Absence of or only very slight local and general reactions.



3. The high degree and long duration of the immunity gained by prophylactic inoculations.

4. The possibility of immunization against pure parasites.

5. The facility with which the inoculation material is preserved.

The disadvantages, however, may be summarized as follows:

1. The manufacture of the inoculation material is rather complex and with some pathogenic bacteria (pest), not without danger.

2. The increased susceptibility during the interval between the inoculation and the onset of immunity.

The last point applies not only to aggressins, but equally to other methods of active immunization. In times of an epidemic, aggressin immunization should never be undertaken.

When one bears in mind the great advantages derived from the employment of this form of immunization, its extensive use should be expected; especially so as animal experimental work with the most important of infectious bacteria: typhoid, cholera (Bail), colon (Salus), dysentery (Kikuchi), staphylococcus (Hoke), has proven it to be quite successful. It is therefore no false prophecy, to say that this method will be employed more and more frequently in the future; particularly for pest, results obtained in animal experimentation by Hueppe and Kikuchi have more than sanctioned its employment in man.

Other methods of immunization based upon the Aggressin principles have been advocated, but none have attained any practical significance.

Mention however must, in passing, be made of the work of Brieger's Briegee and his co-workers Mayer and Bassenge. Briegee's Bacterial had made extracts of typhoid and cholera bacilli, in the main Extracts. identical with artificial aggressins. As far as his sterilization was concerned, he obtained that by filtering the extract through the Pukal filter. One should remember that by this procedure many important substances are lost, but in spite of this, his results of inoculation in man have been most encouraging, and there is a possibility that his method may take the place of Wright's or Pfeiffer and Kolle's, as the reactions are very much milder.

Entirely different from the extracts of living bacteria are those made from previously killed ones. Neisser and Shiga among others have immunized against half parasites in this manner. This is not surprising since the dead bacterial bodies can be similarly used for this purpose. As a general rule, wherever dead bacterial bodies cannot be used for immunization, their extracts will also be found inefficient. The oldest bacterial extracts in use are the tuberculins.

## CHAPTER V.

### TUBERCULIN DIAGNOSIS.

As a member of the class of bacterial extracts, tuberculin merits especial consideration, because it is used not only for immunization, but also for diagnostic purposes. Tuberculin diagnosis can be employed in several ways.

1. As Koch's subcutaneous method.
2. As the cutaneous reaction (v. Pirquet) and ointment reaction (Moro and Doganoff).
3. As intracutaneous reaction.
4. As ophthalmic reaction (Calmette).

#### *Koch's Subcutaneous Method.*

In the chapter on aggressins it was shown that when a normal animal was inoculated with a certain definite quantity of bacterial extract, it could readily withstand any effects of such inoculation. If, however, a similar quantity was injected into an animal previously infected with the same bacterium, dangerous symptoms would be in evidence and if the dose were large enough, death would be likely to follow.

With these facts for reference, the following experiments will be easily understood. A number of tuberculous guinea-pigs, and a number of normal ones as control, are injected with varying doses of tuberculin. After twenty-four hours some of the tuberculous animals are dead, others very ill, while the normal guinea-pigs remain perfectly active. Just as in the aggressin experiment, we have here a bacterial product in itself possessing only slight toxic qualities but which has so increased the virulence of the infection already existing, that an ailment which is usually of a slowly progressive nature becomes transformed into an acute one, terminating in the death of the animal.

The close analogy between the experiments with aggressin as the injected substance, and that of the tuberculin, will become more clear when the nature of the latter is perfectly understood.

Four to six weeks old pure cultures of tubercle bacilli grown in 5 per cent. of glycerin bouillon are filtered, and the filtrate then evaporated down to 1/10 of its original volume.

The resultant fluid, known as tuberculin, is dark brown and syrupy in nature, and keeps indefinitely.



It consists, therefore, of a 50 per cent. glycerin extract of the soluble products of metabolism of the tubercle bacillus.

A part of the glycerin has, however, been used up for the nutrition of the bacteria and thus it is highly probable that after four to six weeks the bouillon contains less than 5 per cent. glycerin and the evaporated solution less than 50 per cent. The specific substances contained within the tuberculin have not been definitely established. As probable elements, however, may be recorded products of secretion of the living bacteria, of degeneration of the dead bacilli and finally the glycerin soluble substances extracted from the bacterial bodies during the heating. No doubt, all these substances and many others about which we lack information, are directly concerned in the activity of the tuberculin.

Another of the many unsolved questions which here present themselves may be mentioned: whether any substances exist in the filtrate which are thermolabile, and therefore destroyed or modified by the heating? According to Bail's researches, the aggrassin of the tubercle bacillus differs from all other aggrassins in that it is not thermolabile and can moreover withstand high grades of temperature. In spite of this, though, attempts to eliminate the heating during the manufacturing of the tuberculin should merit consideration.

If merely the term "Tuberculin" is used, one always has in mind the above filtrate tuberculin, also known as Old Tuberculin.

The experiment with the tuberculous guinea-pigs has its analogy in the use of tuberculin in the case of man. Here, however, in order to avoid dangerous symptoms far smaller doses of tuberculin are selected.

If therefore of two individuals one is tuberculous and the other not, and both are injected with the same amount of old tuberculin 0.001 c.cm., the healthy individual remains perfectly normal while the tuberculous person shows a typical symptom complex which can be described under,

1. General reaction.
2. Focal reaction.
3. Local reaction.

The *General Reaction* consists of, *fever, headache, malaise, nausea, insomnia, cough irritation, palpitation*, etc. The most constant symptom is *increased temperature*; the other manifestations may only be very mild or even entirely absent.

The *Focal Reaction* exhibits evidences of a fresh inflammatory process in the suspicious or old tuberculous foci. In cases of lupus, laryngeal, and iris tuberculosis, this inflammatory reaction can be distinctly seen. In pulmonary tuberculosis the previously vague physical signs may now become definite; rales may appear, dulness may be increased, and eventually pains in the chest may arise.

The *Local Reaction* is noticed at the point of inoculation. In spite of the sterile needle and thorough disinfection, the skin around the site of the injection becomes red, swollen and painful. That this is not due to dirt infection is proven by its absence in non-tuberculous individuals.



[The *local reaction* has been recently advocated by Hamburger as a very delicate diagnostic method. He carries out the test as follows: 1/10 c.cm. of a 1:10,000 dilution of tuberculin is injected just beneath the skin of the forearm or back. The needle should be of fine caliber and the syringe should never have been used for more concentrated solutions of tuberculin. If the reaction is positive a subcutaneous infiltration appears within twenty-four hours. Furthermore, there is a reddening at the site where the point of the needle rested ("Depot reaction" of Hamburger).

If there is no reaction within twenty-four hours, 1/10 c.cm. of a dilution 1:100 should be injected.

This subcutaneous local reaction may also be carried out in the form of an intracutaneous test (see later)].

Of the three types of reaction the general and focal symptoms are the most constant. Both are so characteristic for the existence of tuberculosis, that their appearance justifies the diagnosis. In practice, however, it is the *general reaction, or almost exclusively the manifestation of fever*, which is taken as the guiding symptom in Koch's subcutaneous method.

The focal reaction in all non-visible tubercular lesions is determined by subjective methods, while increase in temperature is alone an objective finding.

In carrying out the subcutaneous tuberculin test, one must remember several practical points which are of help for the correct interpretation of the results. These may be summed up thus:

Inasmuch as the rise of temperature is of diagnostic importance, *no patient with any fever should be subjected to the inoculation*. For several days previous, the patient's temperature should be taken every three hours and only if the temperature does not exceed 37° C. per axilla should the tuberculin diagnosis be undertaken.

*The quantity of tuberculin to be injected is also of the utmost consequence.*

Too high doses should be avoided, as the specificity of this reaction, like all other biological reactions, is limited quantitatively. While small doses of tuberculin will give a rise of temperature only in tuberculous individuals, larger doses may give the same rise even in healthy people. In addition, too large doses as a rule produce a general reaction which might be very severe and injurious.

*The dosage advised by Robert Koch for the diagnostic tuberculin reaction is as follows:*

1. 0.0001 c.cm. T. (for very weak individuals and children).
2. 0.001 c.cm. T.
3. 0.005 c.cm. T.
4. 0.01 c.cm. T.
5. 0.01 c.cm. T.



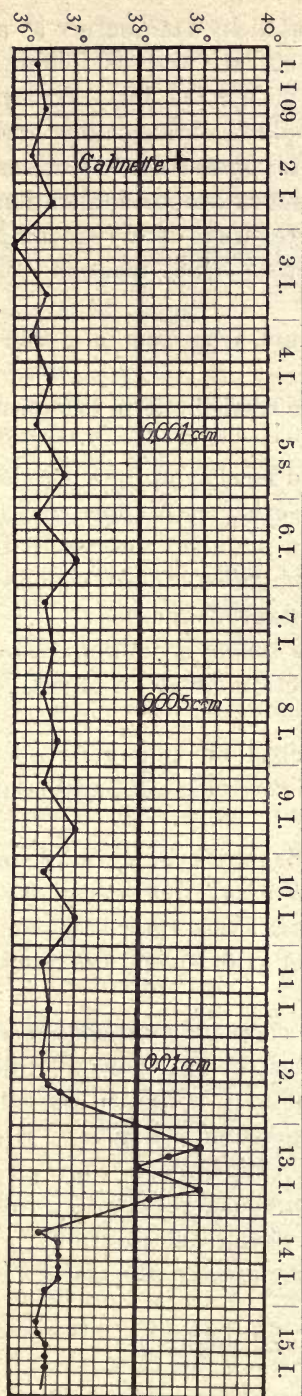


CHART 1.—Example of a diagnostic tuberculin reaction.

The dose chosen at the first injection is 0.001 c.cm. T. Very weak individuals, *i.e.*, those in an advanced stage of tuberculosis or those who have experienced a recent hemoptysis, as well as children, should receive an initial dose of only 0.0001 c.cm. T. Bandelier and Röpke, who have a wide experience in this field, advise 0.0002 c.cm. T. as the primary dose.

Few patients show a distinctly positive fever reaction even with this small dose; by a positive reaction is meant an increase in the temperature so that the latter is at least  $0.5^{\circ}$  C. higher than the highest point before the injection. If the temperature has not increased, the reaction is negative, and after an interval of two to three days of normal temperature the second inoculation of 0.005 c.cm. T. is given. If as happens occasionally after the first inoculation there is a doubtful reaction, *i.e.*, there is an increase of  $0.2^{\circ}$  to  $0.3^{\circ}$  C. then the dosage at the second injection should not be increased to 0.005 c.cm., but the same amount 0.001 c.cm. T. is to be repeated. In a tuberculous individual this repeated injection of 0.001 c.cm. frequently results in a distinctly positive reaction, while in a non-tuberculous patient instead of the former doubtful, a distinct negative reaction is obtained.

The general rules given for the first inoculation also apply to the second with 0.005 c.cm. In a doubtful reaction with this dose, one does not directly proceed to the 0.01 c.cm. dosage, but the 0.005 c.cm. dose is repeated and only after a negative reaction with the repeated 0.005 c.cm. dose is the 0.01 c.cm. injected (see accompanying Chart 1). This represents the maximum amount of tuberculin to be used for diagnostic purposes. Koch advises repetition of this dose if no reaction is obtained. The majority of authorities, however, abstain therefrom. In fact some investigators claim that a reaction obtained after inoculation of 0.01 c.cm. cannot be considered specific, be



cause there are non-tubercular individuals who respond to this quantity of tuberculin.

Most tuberculous persons react after a dose of 0.001 c.cm. to 0.005 c.cm. T.; those, however, who are very far advanced or who suffer from severe cachexia, remain unresponsive to even much greater doses; in addition, patients whose serum contains antituberculin, do not react because the inoculated tuberculin is quickly neutralized.

According to Loewenstein the tuberculin reaction does not depend so much upon the quantity of the tuberculin, as upon the frequency that it is injected. He, therefore, advises that the same amount, about 0.0002 c.cm. be inoculated four times during the course of twelve to sixteen days. In by far the greater majority of tuberculous patients a typical reaction appears after the third or fourth injection. The author has no personal experience with this method, but the reports of other authorities do not exhibit as favorable results as those claimed by Loewenstein.

The inoculation is always to be given subcutaneously, and the back or breast is the best site for it. The dilution is made immediately before the injection, with physiological salt solution or 0.5 per cent. carbolic solution.

In interpreting the result of the reaction one must exclude rises of temperature due to extraneous influences such as Angina, Influenza, etc. Furthermore there are individuals, especially hysterical ones, in whom any injection as such is apt to produce a rise of temperature. To guard against such a possibility an injection of physiological salt solution should be made and thus quiet any suspicion of error.

The diagnostic use of tuberculin is indicated when one is dealing with adults who present *clinical symptoms, or clinically suspicious symptoms of tuberculosis*, but who run no temperature and tubercle bacilli cannot be found.

Tuberculin is contra-indicated in patients with high fever, and during or shortly after hemoptysis or hematuria. In epilepsy, marked cardiac or renal affection, arteriosclerosis, diabetes, and similar conditions, inoculation should be undertaken only under the strictest indications and with great care.

A positive general reaction means that the individual is infected with tuberculosis, but does not throw any light upon the site, the extent, or the prognosis of the infection. The focal reaction allows the diagnosis of the position of the lesion.

### The Cutaneous Reaction.

The cutaneous reaction was first introduced by v. Pirquet, who noticed that by scarification of the skin and application of tuberculin, tuberculous



children would develop a distinct papule at this point, while in non-tuberculous conditions such a reaction would be absent.

### *The Technique of the Cutaneous Reaction.*

"The patient's forearm on the inner side is cleansed with ether; two drops of the pure undiluted old tuberculin are placed upon the skin about 10 cm. apart, and then the skin is scarified first between the two drops, for the purposes of a control, and next within each of these drops.—[A boring scarifier, devised for this, works very easily.]<sup>1</sup> Finally a piece of cotton is placed upon each of these drops and allowed to remain there for ten minutes after which the cotton is removed. A dressing is not necessary."

### *Interpretations of the Reaction.*

Scarification of itself produces the so-called "*traumatic reaction*," i.e., a small wheel with a rose-colored margin appears around each of the three points of scarification. This reaction passes away after several hours and only a small scab remains surrounded by a red rim.

This "*traumatic reaction*" is to be sharply differentiated from the "*specific reaction*." The latter is noticed only upon the upper and lower points where the tuberculin has been applied and consists of a red, indurated papule which rapidly extends in size and elevation, measuring 10 to 30 mm. in diameter. (Fig. 1, Plate I.) The papule may be round or have irregular margins. Scrofulous children show small, irregularly raised follicular infiltrations around the specific reaction. This is known as the "*scrofulous reaction*." It may appear as early as within three hours, but usually occurs within twenty-four hours. It arrives at its maximum within forty-eight hours; occasionally it is delayed and may not develop fully until the third or fourth day and then it begins to fade. Frequently a small pigmented spot remains. General and focal reactions are practically absent.

### **Moro-Doganoff's Ointment Reaction.**

Moro and Doganoff found that a 50 per cent. ointment of tuberculin in lanolin rubbed into the skin without scarification, would give a reaction which consisted of small nodular or papular efflorescences after the nature of Lichen Scrophulosorum. In accordance with the number and size of these nodules as well as the time of their appearance, three grades of reaction are described.

In employing the ointment it should be heated to 25° C. and a quantity about the size of a pea is thoroughly rubbed into the skin of the

<sup>1</sup>The flat end (2 mm. wide) of sterile wooden toothpicks can serve the same purpose.

abdomen or the region of the mammilla, for almost a minute. The diagnostic value of the reaction is variously interpreted.

An almost analogous reaction, described independently of Moro, by Lignières and Berger is to be found in thoroughly rubbing in concentrated old tuberculin into the shaved skin of tuberculous cattle.

### The Intracutaneous Reaction.

Tuberculin even in very weak dilutions when injected directly into the skin of tuberculous individuals produces marked inflammatory infiltrates. This was first observed by Mendel and Mantoux and has been carefully studied in cattle and guinea-pigs by Roemer.

In guinea-pigs the test performed is as follows: the hair of the abdomen is removed by calcium hydrosulfid, 1/10 c.cm. of a 20 per cent. tuberculin solution (0.02 c.cm. tuberculin) is injected with a very fine needle directly into the skin. The result is noted after 48 hours in order to allow the traumatic effects to wear off. Roemer and Joseph differentiate three grades of reaction:

(a) Very susceptible animals (+++) show after 18 to 24 hours a pinkish wheal about the size of a half dollar with a very deep red center. In 48 hours the dark center which represents a blood extravasation attains a greenish hue. After four days a superficial necrosis sets in which leads to sloughing and final scarification.

(b) In less susceptible animals (++) there is no central dark area of the wheal and the latter appears after 48 hours. These animals show a very slight necrosis later on.

(c) The mildest form can be differentiated from the traumatic reaction only in that it does not disappear after 48 hours but remains several days.

In the human being the intracutaneous method has been advised by Hamburger. The results are as yet insufficient to form definite conclusions as to the clinical value of the test.

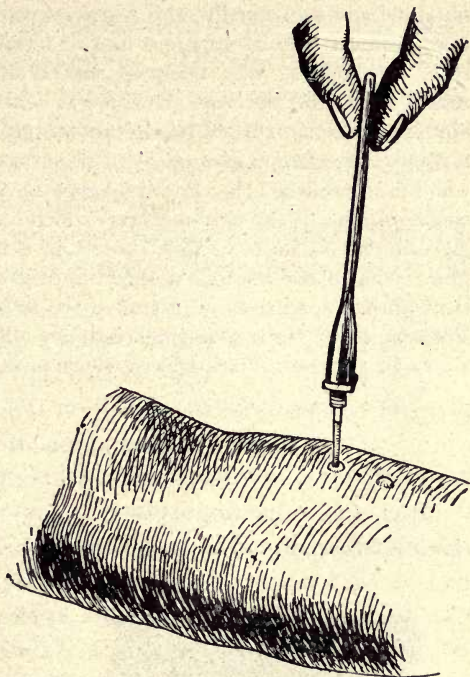


FIG. 13.—Inoculation with tuberculin for the Piquet reaction.



## The Ophthalmic Reaction.

At the discussion which followed v. Pirquet's presentation of his cutaneous reaction, Wolff-Eisner remarked, "that by instilling some 10 per cent. tuberculin into the conjunctival sac, a local conjunctivitis was obtained and occasionally, also a general reaction. The marked severity of the reaction, however, and its apparent lack of specificity, made its diagnostic value improbable." Calmette, who believed that Wolff-Eisner's failure in obtaining accurate results lay in the fact that glycerin was contained in the old tuberculin employed by him, obtained by alcohol precipitation a glycerin-free dry product, which he used in a 1 per cent. solution equivalent to 10 per cent. old tuberculin. It was he, therefore, who first established the clinical diagnostic value of the reaction. But his hypothesis was erroneous, as the mild reactions which he obtained were not due to the absence of glycerin, but because the Lille tuberculin is much weaker than the German preparation. The author was able to show that the old tuberculin could very well be used for the Ophthalmic reaction if, instead of the 10 per cent., a 1 per cent. dilution was made. Thus employed, the reaction is exceedingly mild and specific. Eppenstein later advised a 2 and 4 per cent. dilution in cases where the 1 per cent. solution gave no reaction.

## Technique of Reaction.

It is of extreme importance to have freshly prepared sterile dilutions of the old tuberculin (Höchst Farbwerke). *All the ready-for-use preparations on the market should be discarded. This applies also to the "Tuberculin Test" Calmette's sold by Poulenc Frères.*



FIG. 14.—Ophthalmodiagnosticum for tuberculosis. (After Citron.)

The mishaps and low grade of specificity often ascribed in literature to the ophthalmic reaction can in a great majority of cases be explained by the employment of preparations other than the 1 to 2 per cent. fresh dilutions of the old tuberculin advocated by the author, and in still another number of cases to its employment in conditions where it was distinctly contraindicated. The preparation of fresh tuberculin dilutions is very much simplified by the "*Ophthalmodiagnosticum* for Tuberculosis," of the firm P. Altmann, Berlin N. W. 6 (Fig. 14).

This outfit consists of twelve sealed glass tubes each containing 0.1 c.cm. old tuberculin, a cylinder for the dilution graduated in percentages, and a pipette measuring 0.1 c.cm. fitted with a rubber bulb. One of the sealed ampoules is shaken so that the tuberculin is collected into its broader part and then broken at the designated point near the narrow end. The tuberculin is drawn up to the mark into the pipette and then trans-



ferred into the cylinder. Boiled water or sterile saline is added to the 1, 2 or 4 per cent. dilution mark. The pipette is washed clean in the solution by successive aspiration and expulsion in order to free it completely of the remaining concentrated tuberculin, and can now be employed as the eye dropper.

The solution should be used only on the day it is prepared. The tuberculin in the sealed tube can be kept indefinitely. The pipette and graduate are sterilized by dry heat, boiling or by thorough washing in boiling water.

One drop of the tuberculin dilution is deposited in the inner angle of the eye, and care should be taken that the drop is not immediately expelled, but evenly distributed in the conjunctival sac.

In tuberculous individuals the reaction appears in twelve to twenty-four hours, and according to its intensity can be divided into three grades.

*First Grade.*—Reddening of the caruncle and inner side of the lower lid (+) (see Fig. 2, Plate I).

*Second Grade.*—Same as above but additional involvement of the conjunctiva of the eyeball (++).

*Third Grade.*—Conjunctivitis purulenta, phlyctenulæ and other such severe manifestations (+++).

The reactions of the first and second degree occur most frequently. The manifestations associated with the former of these are so mild that the patient himself does not usually notice them. If the proper dilution is used and the contraindications of this test are observed, a reaction of the third degree is obtained only in exceptional cases. Fever never occurs. The other eye serves as a control. It is advisable therefore before undertaking the reaction, to note carefully any differences that may exist in the conjunctivæ on both sides. It must be remembered that

- |                                      |   |
|--------------------------------------|---|
| Selection<br>of Correct<br>Dilution. | 1. The greater the dilution, the more specific is the reaction.   |
|                                      | 2. The test should not be repeated upon the same eye, even if there was no reaction at all at the first instillation. |

The following procedure should be adopted. A drop of the 2 per cent. tuberculin dilution is placed in the left eye. If a positive reaction takes place, it is of great probability that the patient is suffering from an active tuberculous process and thus the diagnosis is established. If, however, that proves doubtful, and further corroboration is required, the patient should receive, after the first reaction has entirely subsided, one drop of a 1 per cent. tuberculin dilution in the right eye.

If a negative reaction is obtained at the instillation of the 2 per cent. dilution, one drop of the 4 per cent. dilution is placed in the right eye. A negative reaction with the 4 per cent. mixture speaks almost conclusively for the absence of tuberculosis except in far advanced cachectic conditions. A positive result does not, on the other hand, indicate the presence



of tuberculosis, as there are many normal individuals who react to a 4 per cent. tuberculin concentration.

Instead of the tuberculin solution, Wolff-Eisner recommends a 2 per cent. old tuberculin lanolin ointment. The lower lid of the eye is pulled downward and a pea-sized mass of the ointment is gently placed into the conjunctival sac by means of a sterile glass rod. The lid is held fixed for about a minute.

*The ophthalmic reaction* is indicated in all suspicious cases of tuberculosis where the presence of bacilli cannot be demonstrated and where the subcutaneous reaction either on account of the presence of temperature or other reasons cannot be undertaken.

This test is much milder and more agreeable to the patient than the subcutaneous one, and in ambulatory work more significant, inasmuch as it does away with any necessity for considering as a guide the temperature taken by the untrained and usually unreliable patient.

*The ophthalmic reaction is contraindicated in all diseases of the eye, tuberculous or otherwise.* If one eye only is affected, the reaction should not be undertaken upon the healthy eye. Similarly, patients who have had some eye disease, even though many years ago, those who by reason of their occupation are readily exposed to eye diseases, or who live in districts where trachoma is prevalent, should be excluded from the test. The reason being that in those individuals the conjunctival mucous membrane becomes a locus minoris resistentiæ and therefore easily inflamed.

Repeated instillations of tuberculin into the same eye may set up very severe disturbances. Scrofulous children often show reactions of the third degree, inasmuch as they possess the constitutional tendency which makes them easily susceptible to conjunctivitis or phlyctenulæ. In patients with a positive ophthalmic reaction that has subsided, a recurrence of the conjunctival inflammation is frequently observed when they begin to receive subcutaneous inoculations of tuberculin for therapeutic or diagnostic purposes.

### The Specificity of the Tuberculin Reaction.

The one real essential for the practical application of all biological reactions is the specificity of the reaction. There is, however, as will be repeatedly pointed out further on, no single absolutely specific reaction. In fact, it would be more exact to consider these biological reactions only relatively specific; the latter depending upon the quantity of the required antigen and the reacting organism. In this connection it may also be said, that it is never possible to draw an exact line between the specific and non-specific biological reactions. There always will be a doubtful zone. As a general rule, however, the smaller the quantity of antigen that is required and the stronger the resulting reaction, the more probable is the biological specificity.

In tuberculosis this problem is rendered still more complex by the pathological anatomical findings, whereby it is shown that an extraordinary high percentage of individuals have undergone tubercular infection at some time during life. The clinical consideration of tuberculosis, however, does not deal with the diagnosis of these harmless, practically healed tuberculous foci; what the clinician desires to know is whether or not a group of symptoms manifested by a patient is of a tuberculous nature or not. In other words, it is not the latent, inactive, but the active form of tuberculosis that is to be diagnosed. It is that one must view the merit of the various tuberculin tests from this standpoint.

*The reaction of least specificity in adults is the v. Pirquet's cutaneous reaction. In children it is far more specific.*

V. Pirquet has made the following very interesting observation..

Out of 747 children in Escherich's clinic in Vienna upon whom the reaction was tried, there were:

Clinically tuberculous	130, out of which 113 (87 %)	showed a positive reaction;
Clinically non-tuberculous	512, out of which 104 (20 %)	showed a positive reaction;
Doubtful	115, out of which 56 (48.6%)	showed a positive reaction.

Almost all of the tuberculous children who did not react were cachectic.

As for the positive reaction in non-tuberculous cases, the age of the child in large part explains the great differences found.

Whereas healthy infants up to the sixth month almost never give a positive reaction, healthy children of

- 1 to 2 years react in 2 per cent. of cases.
- 2 to 4 years react in 13 per cent. of cases.
- 4 to 6 years react in 17 per cent. of cases.
- 6 to 10 years react in 35 per cent. of cases.
- 10 to 14 years react in 55 per cent. of cases.

In adults one meets with a positive v. Pirquet's reaction in more than 70 per cent. of all cases. V. Pirquet explains this by the presence of latent tuberculosis.

*It therefore becomes self-evident, that the cutaneous reaction in adults is void of any diagnostic value.* A negative reaction only, can be fully relied on, and that, if no cachexia exists.

Ellerman and Erlandsen have attempted to improve upon the diagnostic value of the cutaneous reaction by a quantitative titration of the tuberculin hypersusceptibility. They aimed to get the weakest dilution which still gave a distinct cutaneous reaction. Their results showed wide variations. Of those who reacted to a 1-5 per cent. tuberculin dilution the great majority were clinically tuberculous; at the same time there were many in whom not the faintest clinical suspicion of tuberculosis could be entertained.



In young children on the other hand, v. Pirquet's method should be the one of choice. In addition to its being entirely harmless, and easily applied, it possesses a high diagnostic value.

As for *Koch's subcutaneous reaction*, it is specific, inasmuch as it is a rare exception to get a negative reaction in an active tuberculous process. This occurs only in cases either with very severe cachexia or those with freely circulating antituberculin in the blood. If the latter two possibilities are excluded, the absence of a positive reaction speaks decidedly in favor of the absence of tuberculosis.

The interpretation of a positive reaction as to the existence of clinically active tuberculosis cannot be so definitely answered. From the recent work of most authorities, however, it seems to be taken for granted that a positive reaction does mean an active tuberculosis; still, this statement requires a great deal of consideration and limitation.

In this connection the statistics of Franz are of interest. Out of 400 apparently healthy soldiers in one of the Austrian regiments who in 1901—their first year of service, received an inoculation of 0.003 c.cm. of tuberculin, a positive result was found in 61 per cent. of the cases. In the following year (1902) 100 of the soldiers were re-inoculated and all of those who reacted positively the first time, did so a second time, in some instances even though the second dosage was smaller. Moreover, fourteen others who responded negatively the previous year showed positive results this time, making a total of 76 per cent. Out of 323 men inoculated for the first time in 1902, 68 per cent. reacted positively. It must be mentioned, however, that the majority of the members of this regiment came from a very tuberculous district. The same author also examined a Hungarian regiment in a tuberculous-free district, and under similar circumstances found a positive reaction in 38 per cent. of cases. Although these figures may be exceptionally high, they are without doubt conclusive as to the fact that Koch's reaction cannot be considered specific for "active" tuberculosis. Franz in addition gives important statistics concerning the health of the inoculated soldiers whom he examined for years following the inoculation. The appended charts taken from the most recent publication of Franz (Wien. Klin. Woch., 1909, No. 28) tabulate what has been said above.

Regiment	Year of injection	No. of soldiers inoculated	Positive reaction	During the period of three years, those that terminated their service through death, invalidity or long leave of absence showed.		
			Negative reaction	Tuberculosis	Disease suspicious of tuberculosis	Other diseases
Bosn. Inf. Reg. No. 1	1901	400	{ +245 (61%) -155 (39%)	17 ( 8 deaths) 5 ( 4 deaths)	22 25	10 7 (1)
Bosn. Inf. Reg. No. 1	1902	323	{ +222 (68.7%) -101 (31.3%)	13 ( 6 deaths) 4	28 (1) 13	7 (2) 5
Inf. Reg. No. 60....	1902	279	{ +108 (38.7%) -171 (61.3%)	4 3 ( 2 deaths)	4 5	8 12
Total. ....		1002	{ +575 -427	34 (14 deaths) 12 ( 6 deaths)	54 (1) 43	25 (2) 24 (1)

Regiment	Time of observation	No. ill with manifest tuberculosis	Those who reacted in 1901 and 1902 to 0.003 c.cm. tuberculin	
			Positive	Negative
Bosn. Inf. Reg. No. 1. I Ser. (400 men).	From 10. x. '04 until end of 1908.	10	6	4
Same. II Ser. (323 men); Inf. Reg. No. 60 (279 men).	From Oct., 1908, until end of 1908.	6	5	1
		2	1	1
		18	12	6

Specificity of Ophthalmic Reaction. In regard to the *specificity of the ophthalmic reaction*, the conditions are more favorable than in both of the preceding tuberculin tests. The following short chart is explanatory. Positive reactions were obtained in

	Audéoud	Petit	Citron (1st series)	Eppenstein	Schenk and Seiffert
Tuberculosis.....	94.6%	94.3%	80.7%	72.3%	78.6%
Suspicious cases.....	81.0%	61.6%	80.0%	40.0%	30.0%
Normal cases.....	8.3%	18.4%	2.2%	9.0%	5.8%

Calmette's preparation.

1% old tuberculin.



It is evident from the above figures *that by the use of the 1 per cent. tuberculin a grade of specificity is reached which can be considered quite high, as the non-tuberculous react only in a very small percentage of cases, while existing tuberculosis is detected in 80 per cent. of the subjects.* Clinical examinations of the positive reacting patients show that the latter belong to the group of active tuberculosis. Absolute reliance, however, in the determination as to whether the positive reaction given is due to an active or latent tuberculosis, cannot even be placed on the ophthalmo reaction.

According to several authors, it is claimed that typhoid fever, rheumatism, and syphilis (in the stage of eruption) are very prone to give a positive ophthalmo reaction, without the presence of a simultaneously existing tuberculosis.

In conclusion, therefore, the author finds it difficult to make any general statement as to the preference of one or the other test for diagnostic purposes.

In children the application of the Pirquet reaction, and in adults the ophthalmo reaction, are given preference to Koch's subcutaneous reaction; provided, no contraindications exist against the first, and that treatment with tuberculin is not to be undertaken. In the latter instance, the recurrent ophthalmo reaction when the tuberculin therapy is instituted, authorizes the use of Koch's subcutaneous diagnostic method.

### **Mallein, Trichophytin.**

Similar to old tuberculin, the Mallein (Hermann and Kelning) has been obtained from cultures of Glanders bacilli and the Trichophytin (Plato) has been isolated from the Trichophyton fungi. Mallein has already attained its practical application for the diagnosis of glanders in veterinary medicine. Like tuberculin it is harmless in normal organisms, but brings about temperature and a local reaction at the site of the injection when inoculated into glanders stricken animals. Various general symptoms may also appear. Its employment in a manner analogous to the ophthalmo reaction is also possible.

### **The Prognostic Value of the Local Tuberculin Reactions.**

The fact that the tuberculin reaction in cachectic tuberculous individuals is usually negative has led different observers to regard the degree of the local reaction in a given case as a guide to the prognosis. If, as is considered, the strength of a reaction depends upon the dose of tuberculin, the degree of susceptibility of the organism and its reactive power, the following theoretical possibilities may be considered.

(a) *With mild clinical manifestations*, a reaction obtained only after large doses of tuberculin would speak for a favorable prognosis because the hypersusceptibility of the individual toward tuberculin is still mild.

A severe reaction after a very small dose is less favorable, denoting a very much increased hypersusceptibility.

(b) *With very marked clinical manifestations*, a reaction obtained only after large doses of tuberculin would speak for an unfavorable prognosis because the reactive power of the organism is weak. A severe reaction after a very small dose is more favorable, denoting a stronger reactive power of the individual.

*Wolff-Eisner and Teichmann* formulated the following three hypotheses in regards to the conjunctival test.

1. A positive conjunctival reaction is indicative of an active tuberculosis; associated with manifest clinical symptoms a strongly positive test would speak for a more favorable prognosis than if the reaction were weak or negative. The same interpretation can be applied to the cutaneous reaction.

2. A negative reaction (conjunctival or cutaneous) with manifest tuberculosis would point toward an unfavorable prognosis. The outlook is also unfavorable if the ophthalmic reaction is positive but the cutaneous test negative.

3. With the cutaneous test, the local reaction sometimes continues longer than four days. This is known as a "prolonged or continued reaction" and is found among normal subjects, among individuals in whom the lesions are undergoing a healing process and those in whom the tuberculosis has existed for over ten years. The prognosis therefore is favorable.



## CHAPTER VI.

### THE TUBERCULIN THERAPY.

Right at the beginning it must be made clear, that tuberculin is not to be considered as a curative agent against tuberculosis, but rather in the light of a *bacterial extract for active immunization*. In the previous chapter it has been shown that while there are some infectious diseases where immunization can be accomplished by the use of bacterial extracts and dead bacteria, there are others where immunization is possible only when living vaccines or aggressins of living bacteria are employed. In both of these instances, however, healthy individuals are being treated to be protected from future infection. An exception is presented by rabies. In this disease, the vaccination against the active symptoms is instituted after the infection has already taken place, but the redeeming feature about its treatment is the existence of the very long incubation period. Therapeutic use of tuberculin, however, is a form of active immunization which belongs to neither of the above classes. The principle involved here is entirely different, and the question arises if it is at all possible to obtain an active immunity by the injection of an antigen in a condition where infection has already taken place, and produced pathological changes. [In other words, where spontaneous immunization has failed.]

An answer to this question is to be found in Koch's fundamental experiments which have been the basis as well as starting point of the entire tuberculin study.

If a normal guinea-pig is inoculated with tubercle bacilli, the point of inoculation very soon closes. After ten to fourteen days there appears at this site a small hard nodule which finally ulcerates. This shows no tendency to heal and remains so until the death of the animal. If, however, an already tuberculous guinea-pig is similarly inoculated, while the point of inoculation also closes, no indurated nodule appears. Instead, a necrotic process of the skin sets in after the second day, which finally terminates in the casting off of the slough and the formation of a flat ulceration that heals rapidly. It does not matter whether living or dead tubercle bacilli are used for the second infection.

In explanation of the above phenomenon it must be said that although the first injection had a fatal effect upon the animal, it must have stimulated certain immune reactions within the organism which became manifest after the second inoculation. That a condition similar to this, or even more favorable exists in man, is proven by the fact that while the large

majority of people become infected with tuberculosis at some time during their life, only a small number show symptoms referable to the disease and the rest undergo spontaneous cure.

Koch further showed that the injection of tuberculous guinea-pigs with large doses of tubercle bacilli produced rapid death, while frequently repeated small doses evinced favorable effects upon the site of injection and the general condition of the animals. In this way he proved the beneficial influence which successive inoculations exert upon the primary infection.

In the employment, however, of dead tubercle bacilli in man for therapeutic purpose a serious difficulty presented itself. It was found that the inoculated dead bacilli were not absorbed, but remained for a long time at the seat of the inoculation instigating suppurative processes. On intravenous application, formation of tubercular nodules was noticed.

Koch realized that these harmful effects were due to the non-absorbable parts of the tubercle bacilli; in the main the bacterial capsules. He therefore attempted to extract the immunizing substances, and in this way brought about the old tuberculin.

It may be questioned, whether this old tuberculin is identical with tuberculous antigen; whether it is a feasible preparation for purposes of immunity; whether it contains all the important elements of the tubercle bacillus; if not, which are lacking? The specificity of immunity reactions has already been dealt with sufficiently to make it clear that immunizing a healthy individual with old tuberculin will bring about an immunity only against the substances contained within this preparation. That that does not meet the requirement is proven by the fact that an animal immunized against tuberculin will not be protected against a later infection with living tubercle bacilli. It cannot therefore be expected that immunization of a tuberculous individual with old tuberculin will protect him against living tubercle bacilli. The expectation, however, that his immunity will be raised against old tuberculin only, is fully justified.

Furthermore, we have seen in the aggressin experiments, that inoculation of animals with the aggressin antigen was sufficient to increase the immunity so that a subsequent infection was not attended by any harmful effects. In this case the injected living bacteria are not destroyed, but their ill effects upon the immunized organism have been paralyzed. In other words, the parasites have been transformed to saprophytes. That a similar state of affairs exists in the use of antitoxic sera will readily be seen. The antitoxic diphtheria serum, for example, neutralizes the toxin and thus cures the disease. The bacteria themselves, however, remain intact and also infectious for untreated individuals. Only later on are they absorbed by the phagocytes. When therefore in an individual who has passed through a course of tuberculin treatment there are found fully virulent tubercle bacilli in the sputum, it is no proof, if that is to be the only corroborative evidence, that the tuberculin treatment had been inefficient. In fact, there are strong possibilities that the tubercle bacilli have become transformed into saprophytic bacteria. It is, however, a noteworthy and important fact, that immunization with tuberculin proves no protection against later infection with living tubercle bacilli, while in the case of aggressins and toxins this is possible.



Although tuberculin cannot be considered as the aggressin or toxin of the tubercle bacilli, it simulates these substances with sufficient closeness to warrant its use in tuberculosis. It brings about an immunity against some of the poisonous products of the tubercle bacillus, leaving the others to be taken care of by the natural fighting powers of the individual.

The knowledge that this old tuberculin represents only a partial aggressin, or toxin, and by that is meant that it does not contain all the necessary elements for the establishment of a true immunity, has led to the production of a large group of preparations which are supposed to supply the missing properties of the old tuberculin.

The more important of these preparations were originated by Robert Koch. Those which are of frequent use are:

- a. Old tuberculin (T. Tuberculin)—preparation described on page 45.
- b. Original old tuberculin (T. O. A. Tuberculin Original Alt.)

The latter consists of the original filtrate of the tubercle bouillon culture and varies from the old tuberculin in that it is not heated and reduced to 1/10 its volume. The omission of heating is certainly not without effect, inasmuch as high heat modifies in some way the soluble bacterial substances. This preparation has not been used therapeutically by Koch himself. Spengler and especially Denys, who have made wide use of it under the name of "Le bouillon filtré," have been its main supporters.

c. Vacuum tuberculin (V. T.) is the original tuberculin which has been reduced in vacuum to 1/10 its volume.

d. The aqueous tuberculin of Maragliano (Tuberculina Aquosa) is closely allied to the above tuberculins. It contains all the water soluble extracts of the living tubercle bacilli obtained by extraction of the living bacteria in distilled water, followed by filtration. As is evident, it is prepared on the same principle as Brieger's bacterial extracts and Wassermann-Citron's artificial aggressins.

The above mentioned tuberculin preparations are all very much alike in that they contain the soluble bacterial elements. Their action therefore corresponds more or less to that of old tuberculin.

Another set of preparations have as their basis the insoluble elements of the bacteria and cannot as such, in either living or dead form, be absorbed. Since, however, the absorption of bacteria is a prerequisite to their proper action, it was necessary to so alter the body substances of these bacteria that they could be taken up. Koch found that this was best accomplished by thoroughly pulverizing the bacilli in large mortars. And by this means the first preparation which he obtained was

e. New tuberculin T. R. (Koch) (Tuberculin Rückstand or Residual Tuberculin).

Cultures of young tubercle bacilli are thoroughly dried in vacuum and finely ground in mortars. The pulverized bacilli are agitated in distilled water and the

turbid fluid is centrifugalized. The sediment thus obtained composes the T. R. or the tubercle bacilli residue.

T. R. therefore contains the aqueous insoluble components of the tubercle bacillus, while the soluble ones are retained in the opalescent supernatant fluid which Koch calls TO (Tuberculin Original).

T. R. is readily assimilated by patients. If carefully administered it produces very little infiltration and only slight temperature and general reaction. Its price is comparatively high (1 c.cm. costs 8.50 marks).

The first preparation which contained both the soluble and insoluble elements of the living bacilli was the

f. New Tuberculin—Bacilli emulsion (B. E.) which consists of T. R. + T. O.

The living tubercle bacilli are first pulverized in a mortar and then suspended in salt solution. Centrifugalization is not necessary, but sedimentation is required; 50 per cent. glycerin is added for preservation purposes. Next to T. the new tuberculin B. E. has been most carefully studied.

B. E. also lacks in being an ideal antigen inasmuch as immunity attained by its injections is not at all proof against subsequent infection.

Closely resembling the B. E. is

g. The Tuberculin of Béranek.

Béranek produced two tuberculin preparations of which one is in the main identical with TOA, while the other is an extract of tubercle bacilli with 1 per cent. of phosphoric acid. These tuberculins are mixed and applied together. Sahli reports good results with this mixture.

Although none of the described tuberculin preparations can be considered a true antigen of the tubercle bacillus, they undoubtedly have a favorable effect upon tuberculous individuals. To a certain extent the benefits are derived by the mechanism of partial immunization. This in itself does not, however, explain the entire phenomenon of their successful action.

On examination of the tuberculous organs of animals treated with tuberculin, there will be found within the healthy tissue surrounding the tuberculous foci, a fresh inflammatory reaction. This consists of a sero-fibrinous exudate and a zone of leucocytes intruding to a certain extent upon the tubercular lesion. *Tuberculin acts only upon tuberculous tissue which is still alive and not upon dead, cheesy or necrotic structures.*

If enough tuberculin is given so that death of a tuberculous guinea-pig occurs, the changes found are striking. On dissection, about the point of inoculation Koch reports a marked congestion of the blood vessels giving a red and often an almost dark violet appearance. This discoloration extends for a greater or less distance from the



site in question. The neighboring lymph glands are similarly reddened. Besides the tuberculous changes present within the liver and spleen, these organs show on their surface many blackish-red spots varying in size from that of a pin-point to a hemp seed, and resembling very closely the ecchymosis found in some infectious diseases. On microscopical examination are found no blood extravasations, but very widely distended capillaries directly surrounding the tuberculous foci. The capillaries are so densely plugged with red blood cells that it seems almost impossible for the circulation to have continued in these places. In exceptional cases only, are the blood vessels ruptured and the escaped blood found within the tuberculous foci. The lungs present similar changes, but not as regularly or of such characteristic appearance. The small intestine is often deeply and evenly congested. In all this symptom-complex, the never failing and almost pathognomonic feature is the hemorrhagic-like spots on the liver surface.

Koch considered that the tuberculin brought about the death of the tuberculous tissue. He interpreted the disappearance of the reaction after repeated inoculations with tuberculin, as evidence that the entire tuberculous structure had been destroyed; in other words that healing had set in.

Accordingly, in the first tuberculin era, the erroneous tendency arose to consider those tuberculous patients as cured who after gradually diminishing reactions to tuberculin had become entirely refractory to it. Actually, these individuals had merely become immunized against old tuberculin, and if another preparation such as new tuberculin had been injected, a reaction would have recurred.

Basing their conclusions on experimental work, Wassermann, Bruck and also the author have shown that besides the factor of partial immunization, it is the *focal action* of the tuberculin which is the beneficial agent in its therapy.

The inflammatory hyperemia produced leads to a destruction of the tuberculous tissue, while at the same time the inflammatory process recedes. In addition there is a formation of connective tissue which encapsules the focus and with it also is associated the local stimulation of antibodies.

### The Technique of Tuberculin-therapy.

Three distinct periods can be noted in the history of this therapy. The first began in the memorable year, 1890, when Robert Koch made known his discovery of tuberculin. At this time, the aim of tuberculin treatment was to cause very marked reactions and to continue with the injections until no further reaction was obtained. In lupus, glandular or bone tuberculosis 10 mg. was the initial dose. In tuberculosis of the lungs 1 mg. was the beginning. If the patient reacted to this amount, he received *daily* inoculations of this dose until no reaction appeared. Then 2 mg. T. were given and the same procedure repeated. Quite frequently, depending upon the strength of the individual concerned, 10 mg. was given as the primary inoculation in phthisis, and then rapidly increased. While Koch himself very soon recognized that this rather severe treatment was suitable only for incipient or moderately advanced cases, very sick and far advanced phthisis patients were similarly treated by many physicians.

Following such procedure, decidedly unfavorable results were obtained in the latter class of patients and consequently a marked waning in the enthusiasm which first greeted the tuberculin therapy was the inevitable outcome. Thus the once highly praised remedy was entirely rejected.

During the second period only very few former followers of Koch continued their studies. They, however, made it their business to investigate the causes which accounted for the failure of tuberculin therapy. Their researches led to new principles in the treatment, and to more exact knowledge of its indications as well as contraindications.

The success obtained by the untiring efforts of these investigators brought about after many years a revival of the interest in this therapy. It was again taken up (third tuberculin era) and there is no doubt that when properly handled, tuberculin in well selected cases is of decided benefit. Nevertheless, even at the present day, a conclusive opinion as to many of its details cannot be formed. Attempts are being made to set the individual treatment upon a biological instead of upon the more or less schematic basis thus far employed.

### Old Tuberculin (*Tuberculin Koch T.*).

While it was the aim in the early era of tuberculin treatment to produce very strong general reactions, it is the general consensus of opinion at present that it is best so to arrange the tuberculin therapy as to avoid a general reaction and especially the fever. Goetsch was the man who first called attention to this.

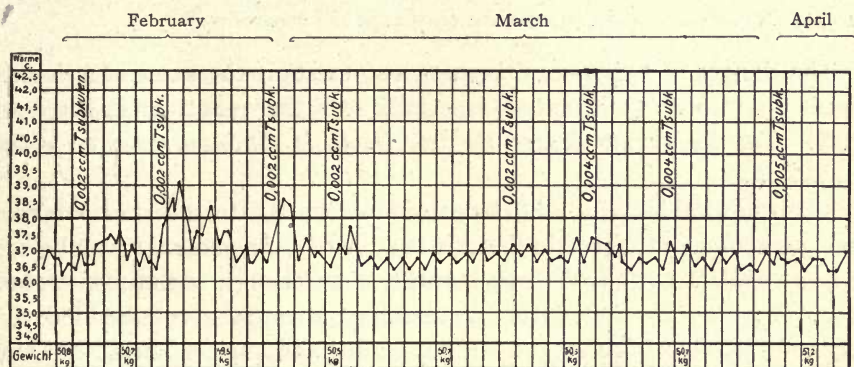


CHART 2.—Example of hyper-susceptibility brought about by going back to a smaller dose.

With such object in view, one must begin with small doses. Some men start with  $1/100$  mg., others with  $1/10$  mg. T. If no reaction is incited, the dose is increased in five to seven days to  $5/100$  mg. and then to  $1/10$ ,  $2/10$ ,  $4/10$ ,  $8/10$ , 1, 2, 4, 6, 8, 10, 20, 40, 80, 100, 150, 200, 300, 400, 600, 800, and 1000 mg. This last amount represents the maximum dose. If the patient still gives a focal reaction with such a dose, it is best to repeat it at intervals of two to four weeks, until finally no reaction is apparent. Occasionally one will advance with the doses at a more rapid rate, but in general, inoculations should not be repeated more than twice a week.



If at any time a distinct or even doubtful reaction occurs, it is absolutely necessary to await the complete subsidence of the latter, and then the same dose is to be repeated. In such a case, an interval of at least eight to ten days should elapse. The dosage should under no condition be diminished, as thereby instead of immunity, hyper-susceptibility is the result. Therefore, patients who at a short time previously had a diagnostic tuberculin test performed should receive the highest dose employed in this test as the initial prescription for their tuberculin therapy. The following chart illustrates the condition of hyper-susceptibility occasioned by a diminution in dosage. (Chart 2.)

Patient had a localized one-sided apex tuberculosis. At a diagnostic tuberculin injection, he reacted only when 0.01 c.cm. T. was employed. After the interval of a month the patient was advised tuberculin treatment. Contrary to the rule just cited, he received as a first injection not 0.01 tuberculin but 0.002 c.cm. T. With this small dose he already had an increase of temperature, although coming rather late, and not quite typical. After this reaction had disappeared, without any other manifestations, the same dose of 0.002 c.cm. tuberculin was repeated and as evident from the chart, a very marked response was inaugurated. This was accompanied by a chill, vomiting, headache, general pains and weakness. In addition there was a slight relapse after the aforementioned symptoms had disappeared. In order to immunize this patient against his hyper-susceptibility, it was advisable to repeat the dose of 0.002 c.cm. T.; the reaction reappeared, but in a very much milder form. It was only after the fifth inoculation of the same dose that no reaction was in evidence. Thus was the hyper-susceptibility overcome and the patient treated in the general way.

The danger of hyper-sensitiveness also exists if the same reactionless dose is too frequently repeated; especially so if the quantities injected are small. *The higher the dosage, the less liable is the occurrence of hyper-susceptibility.*

This question is above all to be considered when after a certain interval, a second course of tuberculin therapy is advised. In general it can be carried out after a period of three months, even though sometimes certain difficulties may be met with. Petruschky strongly recommended this treatment in successive stages. (Etappenbehandlung.) The author is of the opinion that it is best to retain the patient as long as possible at his acquired immunity (tuberculin) by stretching the course of treatment over a long period of time. He therefore repeats an inoculation of the maximum dose, every three or four weeks and when hyper-susceptibility arises, he changes the preparation and begins with a small dose again.

As for the technical details of the treatment, several practical suggestions may be made.

1. The inoculation should, if possible, be given in the morning hours, for a restless night usually follows an injection in the evening.
2. It is best so to arrange the dilutions that the patient receives a fraction of 1 c.cm. at each injection.

3. The site of injection should be alternated between the back and the breast.

4. The temperature should be taken every two or three hours and a chart kept.

5. Disturbances in the general condition of the patient without the presence of fever are to be considered in the light of general reactions just as much as fever without other disturbances.

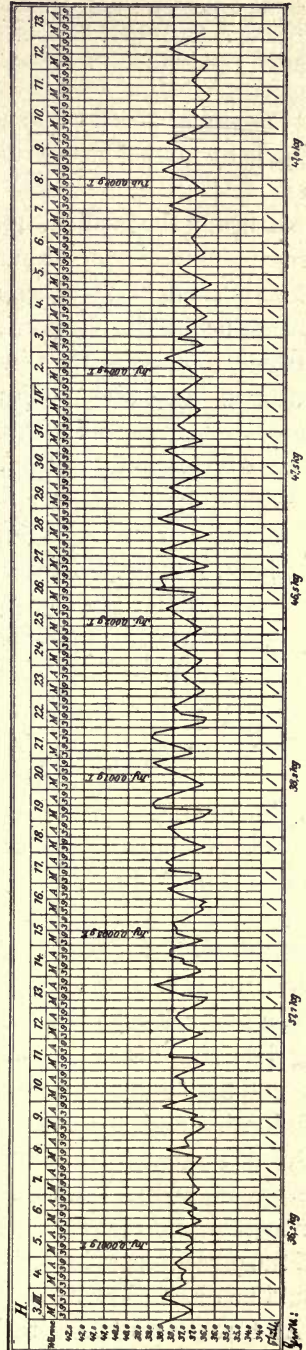
6. The patient's weight should be taken regularly every week, and the dose should be increased provided no loss in weight has taken place.

7. In cases where the pulse increases in rate or becomes poorer in quality, the treatment should be undertaken very carefully and the pulse constantly kept as guide. Slowness of pulse can, as a rule, be considered a *signum bonum*.

Especially favorable for the tuberculin treatment are the individuals with a beginning, localized pulmonary tuberculosis, or cases of lupus, and renal tuberculosis, as reported by Lenhartz. The presence of fever leads some to consider such application as contraindicated. This is indeed incorrect, as frequently it is observed that a chronic fever entirely disappears during a course of treatment, and very often remains away. Even if the fever continues, a good result in the general condition of the patient is nevertheless obtained. (Chart 3.)

Patient H.—Nineteen years old with distinct tuberculous habitus, on admission to the medical service presented a marked infiltration and catarrh of two-thirds of the right lung with a cavity in the upper lobe; infiltration of the left lobe and a great number of tubercle bacilli in the sputum; marked weakness and continuous fever. In five weeks the patient had gained 11 kg. in weight—8 kg. in one week.

Simultaneously his general condition improved very much; the night sweats disappeared, and the cough diminished, but the number





of bacilli still remained the same, and the physical signs of the lungs unaltered. Subsequently the patient received treatment with B. E.; the temperature finally subsided, the cough and sputum likewise, and the bacilli became few and at times entirely absent for several days in succession. In fact, the general condition became excellent. Objectively, there was no demonstration of catarrhal affection.

It might be noted that such a remarkable increase in weight in so short a time is by no means the rule, although good effects are observed in many cases.

Naturally the medical treatment should not be limited to the tuberculin therapy. Even in the immunization of healthy animals, attention is paid to their housing and feeding; so much more imperative is this consideration when applied to sick human individuals.

Bearing in mind that in any treatment, success is only achieved by creating a favorable medium, the same good influences should not be neglected in tuberculin therapy. Rest and forced feeding are curative factors which one cannot omit, and the best places for obtaining these, at the beginning at least, are hospitals, sanatoriums or convalescent homes. When in such a way, the general status of the patient is improved, ambulant therapy may be instituted.

As for the contraindications to tuberculin treatment, it is very difficult to set general rules. Here the opinions of various authorities differ greatly. While for example, Möller and others consider hemoptysis as a distinct contraindication, Aufrecht and Krämer claim that under tuberculin therapy hemoptysis is decidedly improved. It is easy to understand this difference in attitude, if the changes in the focal reaction are considered. There is no doubt that hemoptysis may be excited by increased supply of blood and the inflammatory process associated with the inoculation of tuberculin. The more severe the focal reaction, the greater is this possibility. On the other hand, the new formation of connective tissue and the absorption of the tuberculous tissue will diminish the frequency of hemorrhage. With a general tendency toward hemoptysis, it is therefore best to wait a long time after the cessation of the latter, and then begin with small doses. The patient should be under careful observation and by constant physical examination any possible focal reaction should be controlled. If, in spite of this, hemoptysis does set in, one should not at once be discouraged. An interval of about fourteen days is to be allowed, and then the treatment again undertaken. Frequently, the hemoptysis will cease. If not, or if the patient loses in weight and becomes weaker, the tuberculin therapy should be discontinued.

As further contraindications, Möller mentions marked general weakness, fever, heart affections, epilepsy, and hysteria. In full agreement with Bandelier and Røpke, the author does not consider any of the above as

cause for the non-employment of tuberculin. Only where absolute cachexia, without any possibility for improvement exists, is this therapy to be omitted. In all other conditions, an attempt is by all means justified. Experience, of course, plays an important rôle in the selection of suitable cases. For a beginner, it is advisable to gain practice by the treatment of uncomplicated cases before undertaking those of greater difficulty.

## 2. New Tuberculin-Bacilli-Emulsion (*B. E.*) and New Tuberculin *T. R.*

Treatment with new tuberculin follows along the very same lines set down for old tuberculin.

New tuberculin *T. R.* is the mildest of all preparations. It is very suitable for the beginning treatment of susceptible patients. When the individual does not react to large doses, it is well to start in with *B. E.* *B. E.* can also be employed without producing any reaction, although it is somewhat more difficult.

The dosage scheme advised by Bandelier and Røpke is as follows:

1/1000, 2/1000, 3/1000, 7/1000, 10/1000 mg.,  
 15/1000, 2/100, 3/100, 5/100, 7/100, 10/100 mg.,  
 At intervals of 1 to 2 days;  
 15/100, 2/10, 3/10, 5/10, 7/10, 10/10 mg.,  
 At intervals of 2 to 3 days;  
 12/10, 15/10, 2, 2 1/2, 3, mg.,  
 At intervals of 3 to 4 days;  
 4, 5, 6, 7, 8, 9, 10 mg.,  
 at 4 to 6 to 10 days intervals.

In susceptible patients, it is best to increase the dosage only by one-half mg. even when large doses are administered. Ten mg. *B. E.* represents the maximum dose.

The author himself follows a different scheme from that of Bandelier and Røpke. The injections are given less frequently, only about once a week, but the dose is always increased twofold, fivefold and even tenfold without any excessive reactions.

Fever is obtained much less often with new tuberculin than with *T.* The reaction usually is in the form of lassitude, nausea, weakness, insomnia, etc.

The treatment with new tuberculin is particularly favorable in cases where a low continuous fever is present. It also is more potent in destroying the bacilli of the sputum. The author therefore prefers the *B. E.* to all other tuberculin preparations. Bandelier and Røpke have also obtained gratifying results with the *B. E. therapy* as is evident from the following statistics of 205 patients treated at the sanatorium at Kottbus.



Stage		I	II	III
Cured (A) .....	23 = 11.22%	10 = 37.04%	13 = 10.48%	0 = 0 %
Completely able bodied (BI) .....	98 = 47.80%	12 = 44.44%	77 = 62.09%	9 = 16.63%
Satisfactory result (A+BI) .....	121 = 59.02%	22 = 81.48%	90 = 72.57%	9 = 16.63%
Able bodied in terms of law (BII)...	70 = 34.15%	5 = 18.52%	30 = 24.19%	35 = 64.81%
Total improvement (A+BI+BII) ..	191 = 93.17%	27 = 100 %	120 = 96.76%	44 = 81.44%
Negative result (C) .....	14 = 6.83%	.....	4 = 3.24%	10 = 18.56%
Bacilli in sputum on admission ....	114 = 55.61%	4 = 14.81%	63 = 50.81%	47 = 87.04%
Absence of bacilli or sputum at discharge.	59 = 51.78%	4 = 100 %	49 = 77.78%	16 = 34.34%

The objection has been frequently advanced that B. E. is absorbed with difficulty and tends to produce infiltrations. This may be readily overcome by preliminary use of T. R. or by the employment of sensitized B. E., as has been advised by Meyer and Ruppel.

By sensitized B. E. is understood a bacilli emulsion which has been mixed with the tuberculous serum of a horse or ox containing anti-tuberculin. This mixture brings about a union between certain of the antibodies and substances contained within the bacteria. The tuberculous serum is then removed by centrifugalizing and washing the mixture with physiological salt solution.

The sensitized B. E. (S. B. E.) is milder than B. E. and in its character is more like T. R. The infiltration is much less, or entirely absent, due, as a general rule, to the fact that sensitization of bacteria tends to neutralize those substances which produce infiltrations. This last has been demonstrated by the author in the case of mouse-typhoid, and swine pest bacilli, where marked infiltrations following their inoculation have been avoided by sensitization. The following chart illustrates preliminary treatment with S. B. E. followed by B. E. (Chart 4.)

The patient was a female who at the time of admission presented double-sided apical and suspicious intestinal tuberculosis. Tubercle bacilli were present in the sputum. The patient was discharged from the clinic as relatively cured, *i.e.*, all manifestations of illness had disappeared with the exception of slight dulness over one of the apices which, however, could have been attributed to cicatrization. In addition, there was normal vesicular breathing, no temperature, no catarrh, and a good general condition. Undoubtedly it is difficult to say whether this case is cured. Only years of observation can prove this. A temporary latency of symptoms must always be considered. Suffice it to say, that the patient was a great deal improved and able to return to her work.

On close observation of this chart it will be noticed that practically no reactions occurred in spite of the rather rapid increase in the dosage. Slight temperatures were manifest only occasionally (0.0001 c.cm. *S. B. E.*). Even though the same dose was not repeated on account of the long intervals between the individual injections, no increased reaction appeared after subsequent inoculation. When 0.1 c.cm. of *S. B. E.* produced no reaction, the susceptibility to *B. E.* was tested. Doses of 0.0001 *B. E.* to 0.5 *B. E.* were administered at short intervals, without any symptoms. Only after 1.0 c.cm. of *B. E.* was there a slight increase of temperature with rather marked general manifestations—(headache, pains in the extremities, weakness, etc.), which subsided within twenty-four hours. On repetition of the same dose, no reaction occurred.

Six days later, when for the third time 1 c.cm. of *B. E.* was given there set in a much more marked general disturbance, as evidence of hyper-susceptibility. (See Chart 2.)

### 3. Bovine-tuberculin.

Koch's differentiation between bovine and human tuberculosis led to the attempt at immunization of cattle with human tubercle bacilli. (Bovo-vaccine of Behring, and Tauruman of Koch).

Spengler tried to reverse this use and employ the milder, infectious bovine bacilli for the tuberculin therapy in man. He used these

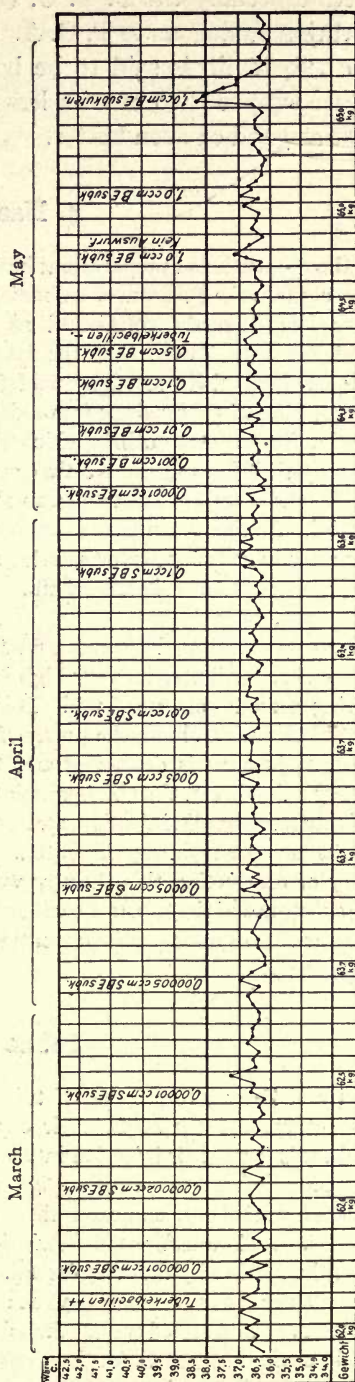


CHART 4.—E. H. 41 years. Intestinal tuberculosis and a double-sided pulmonary tuberculosis. Treatment with *S. B. E.*; almost without any reaction. Immunization with *B. E.* Relative cure. Disappearance of cough, expectoration, bacilli, catarrhal symptoms of lung and intestinal symptoms. Increase in weight.



bacteria to make up preparations similar to the *old and new tuberculin*. He favored especially the P. T. O. (Perlsucht Original Tuberculin) *i.e.*, the preparation analogous to T. O. A.

Bovine tuberculin is said to be borne better than the human. The reactions are supposed to be of a less severe nature, and the therapeutic results just as good or even better.

#### 4. Nastin.

All the above-mentioned preparations have as their aim the production of an agent which is to contain the substances embodied within the tubercle bacilli, and which are more or less correctly considered as representing their poisonous elements. Deycke and Reschad showed that the fat-like material encapsulating the bacteria, to which is ascribed their strongly acid fast character, also plays an important rôle in the question of tuberculosis immunity. These men prepared a wax-like substance, nastin, from a streptothrix which they found as a saprophyte in a case of leprosy—streptothrix leproides. Nastin closely resembles the fat-like substance of the tubercle bacilli and with it one can immunize healthy guinea-pigs against living virulent tubercle bacilli. In the treatment of tuberculosis, however, it has no beneficial effect. On the day after inoculation, fever sets in, sputum increases in great quantities and contains large amounts of tubercle bacilli. In leprosy, slight improvement has been noticed by its use.

Metalnikoff has confirmed the above findings and further shown that the bee moth, *Galeria Molinella*, attributes its very high immunity against tuberculosis to a strong wax dissolving ferment possessed by it. It is probable, too, that inoculations of nastin produce antibodies which have the power of dissolving fat. In this way the capsule of the tubercle bacillus is destroyed and the antigen is liberated to be absorbed. While healthy animals can thus be immunized, tuberculous individuals would be indirectly receiving a tuberculin injection, and its amount would depend upon the quantity of tubercle substance suddenly liberated. It seems to the author that the more rational way of conducting this therapy would be, to first obtain a high immunity against the substance of the tubercle bacillus by injection with B. E., and then to follow this by treatment with nastin. Such treatment may prove an interesting new step in tuberculin therapy.

#### Tebesapin.

Noguchi and Zeuner found that if tubercle bacilli are exposed to the action of soaps of unsaturated fatty acids, their capsule is penetrated and the bacteria are destroyed; in this form their injection into animals will produce no infection or only a mild slowly progressive one. Guinea-pigs which remain healthy after receiving such saponified dead organisms, can after three months withstand infection by virulent tubercle bacilli. At Zeuner's recommendation a preparation known as Tebesapin has been put on the market by Schering-Berlin. It is made up in the following manner: Tubercle bacilli are shaken for four days at a temperature of 37° C. with an emulsion of sodium oleate 1:60 in distilled water; the mixture is then heated for one hour at 70-72° C. and again shaken at 37° C. for three days; then centrifugalized and filtered. It is preserved by the addition of 0.4 per cent. trikresol. Various dilutions of the preparation can be obtained.

Preliminary treatment of goats and guinea-pigs with Tebesapin seems to offer a certain protection against future infection. Evidence of its beneficial action in man is still insufficient. Its harmlessness, however, has been demonstrated, so that one is justified in its use.

Pursuing the same plan, Deycke and Much have attempted to obtain products for immunization by shaking tubercle bacilli with lecithin, neurin, cholin and lactic acid. Their findings have been disputed, although the author has reached conclusions similar to those of Deycke and Much. Further corroborative evidence is necessary.



## CHAPTER VII.

### TOXIN AND ANTITOXIN.

So far, the preceding chapters have dealt with immunization by the bacterial bodies and substances extracted from them. Further attention must, however, be paid to the products of secretion of bacteria, namely the toxins. Only few classes of bacteria have true soluble toxins such as are possessed by tetanus and diphtheria bacilli. The symptom-complex incited by the toxin-producing bacteria differs decidedly from that of the sepsis class.

A comparison between anthrax and tetanus certainly exhibits striking differences. Although both are wound infections caused by characteristic bacteria, smears of the pus from wounds, in the case of anthrax, display on examination numerous bacilli, while in the case of tetanus, the bacillus is very sparsely found. Even carefully prepared anaerobic cultures, or inoculations in mice of the pus itself, do not always demonstrate the tetanus bacillus. In the blood, lymph glands and viscera of anthrax cases, excessively large numbers of microbes can be found, while even in the most fatal cases of tetanus, there is nowhere any evidence of bacteria or their spores. Where so many living foreign organisms are found invading the individual, no hypotheses are necessary for explanation of the associated marked disturbances as in anthrax; it is, however, more complex to understand the severity of the symptoms in conditions like tetanus, where such exceedingly scant bacteriological findings exist. Here the micro-organisms play only a secondary rôle, the entire symptom-complex being produced by a poison extruded from the bacteria. In diphtheria, conditions are similar to those in tetanus, although in the former the bacilli can be readily demonstrated both microscopically and by culture. Even though, however, the localization of the bacteria in diphtheria is confined to organs not absolutely essential for life—diseased tonsils—these themselves do not explain the alarming situation observed in this disease; the real cause of the illness is to be found in the toxin which is secreted by the bacteria, and distributed by the blood stream throughout the entire system.

That a toxin really exists, and is not hypothetical, Roux and Yersin, as well as Kitasato, have proven by demonstration of the poisonous agents in the bouillon cultures of both diphtheria and tetanus. As most cultures

show only slight tendencies to toxin formation, a virulent toxin may necessitate a special strain of the bacterium.

The length of time required by cultures for the production of moderate amounts of toxins is by no means constant. With diphtheria this varies from several days to 2 to 3 weeks. As a general rule, if toxin is not liberated within the first four weeks it will most probably not appear after that time. It is isolated by filtering the bouillon culture first through filter paper to remove the pellicle, and then through a bacterial filter to get rid of the bacteria. A layer of toluol 1 to 2 cm. is added for the purposes of sterilization and it is advisable to agitate the toxin and toluol thoroughly every day to prevent contamination.

It does not fall within the scope of this book to take up the various methods proposed for obtaining and preserving the various toxins. It is the object merely to review the details associated with their mode of action and standardization.

The first and most important member of this group is the diphtheria toxin.

The diphtheria toxin is first tested by subcutaneous injections into guinea-pigs 250 gms. in weight. The action of the toxin is entirely dependent upon the dosage; the more toxin injected the more rapidly does death occur. This, however, is not to be taken in mathematically correct proportions—i.e., twice the dose does not produce the same action in one-half the time. *A certain period of time must always elapse before death can take place, the minimum being about one day.* This interim is known as the period of incubation and it is the existence of this that goes to make one of the essential characteristics of a true toxin. A toxin requires a definite period of time for its action to become manifest; and even the largest dose of toxin *cannot diminish the length of this period below a certain minimum.* On the other hand, the length of the incubation time can be increased by the injection of a smaller dose, so that ultimately a dose small enough is obtained which is not instrumental in producing death (Dosis sublethalis).

If a guinea-pig is inoculated with a quantity of toxin sufficient to kill it in three to four days, nothing abnormal is evident the first day; various manifestations of illness, however, follow soon after.

Edema appears at the site of inoculation. The animal stops eating, sits in a corner, and reacts poorly to sound. Gradually it becomes weaker, so that when placed upon its back it does not resume its normal position; the temperature which at first rose somewhat, falls abruptly and then death takes place.

At autopsy, a gelatinous and strongly hemorrhagic edema is found which starts at the site of the injection. On opening the abdominal cavity one finds but very little peritoneal exudate, strongly injected vessels of the mesentery, and especially characteristic, markedly reddened adrenal glands. In the thorax are found bloody pericardial and pleural exudates, and consolidated areas in the lungs.

After the injection of smaller doses, edema likewise arises and becomes larger in extent the slower the case progresses. Besides this, the animal loses in weight. With sublethal doses, edema or infiltration is confined to the site of injection, and finally, with the minutest doses, no edema occurs, but the hair falls out at the place of injection.



Guinea-pigs surviving a dose of toxin may after two to four weeks begin to show paresis first of the hind, then of the fore extremities, and finally even of the muscles of the back and respiration. The most severe types of such conditions, however, may fully subside. They may be considered as analogous to the post-diphtheritic paralysis taking place in man, which is usually of a benign nature.

Besides guinea-pigs other animals suitable for diphtheria experimental work are rabbits (especially by intravenous injection) and pigeons (by intramuscular injection).

The susceptibility of animals towards diphtheria toxin varies greatly, as is seen from the following scale of Behring, the least susceptible animals being mentioned first: mouse, rat, dog, guinea-pig, rabbit, sheep, cow, horse, goat.

*The strength of the diphtheria toxin is estimated as follows:*

Estimation of Strength of Diphtheria Toxin. Guinea-pigs of equal size (250 gms.) receive subcutaneous injections of decreasing amounts of toxin. With a strong toxin, centi- and milligrams or even smaller quantities are of sufficient potency to produce death. Doses such as these are not injected unless diluted in normal salt solution.

For exact results one must not depend upon the findings from the injection of a single animal with each dilution; several should be inoculated with the same dose and the effects, which should be the same in all cases, noted. It is impossible to state beforehand how many dilutions may be necessary. If the various actions dependent upon the successive gradations of dosage are successfully represented, the experiment may be taken as conclusive; that is to say, the smallest doses must leave the animal entirely unaffected, the moderate produce slight local and general symptoms, and the larger ones cause death of the animals. If it should so happen that they all die, a new set of experiments employing a lower scale of dosage should be undertaken.

Thus it is seen that the *action of diphtheria toxin is subject to the quantity of the toxin injected*. If several different diphtheria toxins are tested at the same time, it is at once evident what far reaching differences may arise. While 0.001 c.cm. of one diphtheria toxin kills a guinea-pig in twenty-four hours, a different diphtheria toxin will do the same with a dose ten times as great, e.g., 0.01 c.cm. The second toxin thus contains only one-tenth as many of the active substances. *In order to obtain a uniform method for estimating the strength of a diphtheria toxin and thus get comparative values, a standard unit has been adopted. And this consists of the smallest amount of toxin that will kill a healthy guinea-pig weighing about 250 gms. in four to five days.* This is known as the *minimum lethal dose or dosis letalis minima*. In addition to this "direct toxic value," it is frequently important, especially for the standardization of curative sera, to estimate the "indirect toxic value" by which is meant the amount of antitoxin which a toxin can bind or neutralize.

If an animal, e.g., a goat is injected with a sublethal dose of diphtheria toxin and after the lapse of a certain period of time it is reinjected with a lethal dose, the animal remains alive. In fact it may receive numerous fatal doses, and still survive. This experiment is the simplest in active immunization against a toxin. An examination of the blood serum of the immunized animal will disclose very readily what has taken place. If this serum is mixed with a fatal dose of toxin and the mixture inoculated into a normal guinea-pig, the latter remains alive and perfectly active.

The serum of the immunized animal therefore contains a protective agent which is directed against the toxin and destroys its activity; hence the name antitoxin. But *the antitoxin is specific, i.e.,* diphtheria antitoxin neutralizes only diphtheria toxin and not tetanus. The recognition of these facts and those heretofore mentioned, and the recommendation of the therapeutic use of diphtheria serum belong entirely to v. Behring; righteously may he be called the father of serum therapy.

Although theoretically the serum of any animal immunized with diphtheria toxin can serve as a curative serum for diphtheria, practical experience has taught that it is best to employ horses for this purpose. For laboratory experiments goats should be the animals of choice. It is advisable to use the above animals for the reason that larger quantities of serum are obtained and furthermore because *it has been found impossible to immunize guinea-pigs with previously unchanged diphtheria toxin even if the initial dosage is the smallest subdivision of the minimal lethal dose.* Behring and Kitashima showed that after repeated injections of very minute doses they were able to kill guinea-pigs even with 1/400 of the dosis letalis minima. This is but another example of an effect just opposite to that of immunity and known as *hypersusceptibility* or *hypersensitiveness*, which has already been described in the chapter on tuberculin therapy. If, however, it is desired to immunize guinea-pigs, a *modified form of the diphtheria toxin* must be employed for the first injections. Several modifications are feasible. Behring and Kitasato added iodine trichlorid to the toxin while Roux and Martin chose Lugol's solution; C. Fränkel heated it to 60° C., and Behring advocated the so-called "simultaneous method" (of special aid in tetanus toxin), where mixtures of toxin and antitoxin are injected and gradually the quotient of the latter is diminished until finally it is entirely omitted. If the animals have borne the first inoculations of the modified toxin without any ill effects, one may then use the unmodified toxin.

In contrast to small animals, horses can be immunized with unmodified diphtheria toxin right from the start. Nevertheless great care must also here be exercised. Certain it is, that less risk is run in the employment, with even the larger animals, of a modified toxin. For the production of a good diphtheria serum, healthy horses about five to six years old are used and gradually increasing amounts of diphtheria toxin are injected subcutaneously or even intravenously; thus agreeing with Ehrlich's findings to the effect that the antitoxin content of a serum can be raised by successively increasing the amount of toxin injected. As far as the efficiency of the immune serum is concerned, it is entirely dependent on the animal. Horses vary greatly in their individual predisposition toward the production of an effective serum; some animals



even completely fail to do so, not that the latter are not actively immunized, for they are, but because they contain very little antitoxin within their serum.

It is impossible to recommend a distinct scheme for the immunization of a horse. The intervals between the injections and the size of the dose are varied according to the reaction of the animal toward previous inoculations. *A good rule to follow is, that a fresh injection should be given only if the reaction from the preceding one has entirely subsided. The reactions are both local and general.* The local reaction comes in the form of edema, infiltration, and sterile abscesses; the general, loss in weight and appetite and increase in temperature.

The following chart of Salomonsen and Madsen, of the Copenhagen Serum Institute, serves as an example how a diphtheria serum is produced. A gravid mare 665 kg. in weight was selected and injections were given as follows.

Day	Dose of toxin	Remarks	Day	Dose of toxin	Remarks
1	1 c.cm.		135	.....	Serum removed contained 150 immunity units.
6	1 c.cm.				
12	3 c.cm.		154	.....	Birth of colt.
15	5 c.cm.		177	.....	Serum removed contained 45 immunity units.
23	10 c.cm.				
27	20 c.cm.		184	100 c.cm.	
36	25 c.cm.		188	200 c.cm.	
41	50 c.cm.		195	400 c.cm.	
45	75 c.cm.		205	700 c.cm.	
50	100 c.cm.		213	800 c.cm.	
57	150 c.cm.		223	600 c.cm.	
72	250 c.cm.		232	600 c.cm.	
81	450 c.cm.		242	1000 c.cm.	
92	600 c.cm.		252	.....	Serum removed contained 120 immunity units.
104	900 c.cm.				
119	1000 c.cm.				

The time selected for venesection is important. Antitoxins like any other antibodies do not arise immediately after an injection, but only after a certain *incubation period*. The amount of antitoxin at first gradually increases, then begins to sink, and after that remains constant for a certain period until it finally disappears. If at a time when the serum contains a certain amount of antitoxin a new inoculation is undertaken, the so-called "negative phase" sets in, *i.e.*, the amount of antitoxin within the serum sinks. It is followed by a compensatory rise, "positive phase." By becoming acquainted with the wave-like fluctuations in the antitoxin content of the serum, and renewing the injection at the time of highest content, one can produce a serum with very strong antitoxic qualities. This

was done by Salomonsen and Madsen who by experimentation found that the maximum height of the antitoxic curve was reached on the tenth day after each inoculation. For this reason it is wise to choose this day for the removal of the serum. As regards other sera, *e.g.*, tetanus, different periods have been empirically found to be most serviceable. As the antitoxic curve does not remain at a high point for a long time, the injections should be repeated from time to time. For highly immunized horses, monthly injections usually suffice.

After the serum has been obtained, the important problem which arises is how to keep it sterile. This is accomplished by aseptic precautions at the time of obtaining the serum and eventually by the addition of preservatives such as 1/2 per cent. carbolic acid or 0.4 per cent. tricresol.

This procedure finished, the next step is to estimate the amount of the antitoxin content in the serum.

According to v. Behring and Boer, the value of the serum should be ascertained in respect to its:

- |                     |   |                       |
|---------------------|---|-----------------------|
| 1. Protective power | } | against infection.    |
| 2. Curative power   |   |                       |
| 3. Protective power | } | against intoxication. |
| 4. Curative power   |   |                       |

v. Behring found that these four properties run parallel with each other so that for practical purposes, it suffices to establish only one of these qualities. For diphtheria serum it has proved most serviceable

Standardiza- to estimate the strength of the immunity against intoxica-  
tion of tion, since one is dealing with a purely antitoxic serum.  
Diphtheria

Serum. Behring's original mode of standardization consisted in gradually adding doses of serum to the minimal lethal dose of toxin and injecting the mixtures into guinea-pigs, thus determining the smallest amount of serum capable of preventing death of the animal. It was soon found, however, that this method gave too inconstant results because the individual minimal lethal dose was too variable.

Ehrlich, therefore, modified the process by using ten times the minimum lethal dose. This amount of toxin, mixed with decreasing amounts of serum and made up to 4 c.cm. with physiological salt solution was injected subcutaneously into a guinea-pig. The smallest amount of serum which saved it from being killed on the fourth to fifth day was thus estimated.

The method of standardization used at the present time owes its origin to Ehrlich.

In order to attain uniformity in the comparative value of all sera, Behring and Ehrlich recommended the adoption of two empirical values; "the normal toxin," and the "normal curative serum."

The normal diphtheria toxin is one which contains enough toxin in 1 c.cm. to kill 25,000 gms. of guinea-pigs or 100 guinea-pigs each weighing 250 gms.



A normal curative serum is one of which 0.1 c.cm. suffices to neutralize 1 c.cm. of Behring's normal poison, *i.e.*, is able to overcome the effect of 100 fatal doses. 1 c.cm. of this normal curative serum represents one immunity or antitoxin unit.

The present antitoxin unit was fixed by Ehrlich. He adopted that amount of antitoxin as his standard, which when mixed with 100 times the lethal dose of a then existing toxin, and injected into an animal, was sufficient to so neutralize the toxin that not the slightest evidence of either a local symptom or general illness was present. Ehrlich chose the antitoxin rather than the toxin as the constant of standardization, because the toxin would deteriorate after some time, while the antitoxin could be preserved in a stable, unchangeable form.

In spite of this fact, the new method of titration was still unsatisfactory, inasmuch as the toxin could undergo other biological changes not yet taken into account.—To understand these, the acquaintance of several new terms is essential, and they are, *dosis certe efficax*, *limes+* or *limes death*, *limes 0* or *limes zero*.

While the *dosis letalis minima* represents the smallest dose of toxin which may be fatal in four to five days, the *dosis certe efficax* (dose of certain efficiency) stands for the smallest dose which will surely kill any pig of 250 gms. within this period of time.

By *limes+* (*limes death*) is meant the smallest amount of toxin which after being mixed with an antitoxin unit, will still cause the death of a guinea-pig within four to five days. By *limes 0* (*limes zero*) is understood the dose of toxin which is just neutralized by one antitoxin unit (I. E. = antitoxin unit or "Immunitäts Einheit"), so that no toxin is free and the animal remains perfectly well. *Limes +* therefore implies an excess of poisonous toxin; *L O*, perfect neutralization.

Theoretically speaking, the difference between *L+* and *L O* should represent the minimum lethal dose (*d. l. m.*). This, however, is almost never so, as is shown in the following illustration.

The *d. l. m.* of a certain poison was estimated as 0.0039 c.cm.

*L+* was found to be 0.48 c.cm. = 123 lethal doses.

*L O* was found to be 0.42 c.cm. = 108 lethal doses.

---

Difference                      0.06 c.cm. = 15 lethal doses.

In order to explain this phenomenon Ehrlich considered that there were two other substances contained within the diphtheria bouillon in addition to the diphtheria toxin; namely, diphtheria toxon and diphtheria toxoid. The toxon is a poison which in contrast to the toxin has only a slight affinity for the antitoxin. It is this body which is probably the cause of the paralysis occurring weeks after the infection.

In a mixture like L O, the antitoxin has fully neutralized both the toxon as well as the toxin. If, however, more diphtheria poison is added to the I. E., as is done in the L +, the antitoxin, on account of its greater attraction for the toxin, will combine with the latter and leave the toxon free to subsequently carry out its own functions. The more crude poison is added, the more toxon remains unbound, until a point is reached when no more toxin can be taken up and consequently some is left unneutralized. If the amount of active toxin reaches the *dosis letalis minima*, it is sufficient to kill the animal and thus the limes + is attained.

When, instead of freshly prepared toxins Ehrlich employed older bouillon cultures, the poisonous qualities distinctly sank to about one-half, but the surprising fact was that the L + had not been altered and even though it had lost one-half of its toxic power, it had still retained its initial activity for neutralizing antitoxin.

Ehrlich's explanation was that the diphtheria poison consists of two Toxoid. molecular groups; one the carrier of the toxic qualities, and therefore known as the "toxophore" group, the other uniting with the antitoxin and having the capability of neutralizing it, known as the "haptophore group." The toxophore group is very labile, while the haptophore group, strongly in contrast to it, is characterized by its stability. The toxophore element destroyed, the diphtheria poison loses its toxic qualities, but retains its power to bind antitoxin. A non-poisonous diphtheria toxin possessing such power is designated by Ehrlich as "Diphtheria Toxoid."

The mode of standardization of serum advocated at the present day is applicable exclusively to the L + dose. It is effected by injecting guinea-pigs subcutaneously with mixtures of various doses of diphtheria toxin on hand, plus an anti-toxin unit, and noting the smallest amount of toxin which kills the animal in four to five days. This L + as the constant factor is now mixed with different amounts of the serum to be tested and that quantity determined which just prevents the death of the animal. If for example 1/100 c.cm. is necessary, this serum is considered one hundred times as strong as the standard antitoxin unit, or in other words it contains 100 immunity units.

This method of Ehrlich has been adopted not only in Germany, but almost in all other countries in Europe and also in America. In France the principle varies somewhat, as here the serum is tested both for its protective and curative action. The *protective power* of a serum is considered 50,000 if 0.01 c.cm. of a serum saves a guinea-pig weighing 500 gms. from the fatal consequences following a dose of toxin sufficient to kill an animal of the same weight in thirty to forty hours. The standard therefore takes into consideration the relation between the amount of serum and the weight of the animal. The serum is injected into the guinea-pig twelve hours before the toxin and the animal should not lose in weight during the following six days. The *curative power* is estimated by injecting a guinea-pig with a dose of toxin (sufficient to kill a



control animal in thirty to forty hours) and six hours afterward the serum is injected. The animals remaining alive on the sixth day are considered as cured.

The French method of standardization is built upon the belief of Roux that no parallelism necessarily exists between the protective and curative values of a serum. Kraus and Schwarz have recently published accounts of experiments which corroborate Roux's views. They claim that a very highly valent diphtheria serum has a lower curative value than one less so; that the curative power of a serum does not depend upon the increase or decrease of the antitoxin content during the immunization of an animal, and that Ehrlich's process of standardization, taking into consideration only the protective power, requires additional modification. Berghaus working in Ehrlich's Institute answered the above exceptions in so satisfactory a manner that up to the present day Ehrlich's views are still upheld by the majority of workers in this field.

On the basis of former experiments by Ehrlich and Marx, Roemer has recently suggested the intracutaneous method for estimating diphtheria antitoxin. Its principle depends upon the finding that with intracutaneous injection of mixtures of toxin and antitoxin, even the smallest amount of toxin if not fully neutralized will produce edema.

While in some countries the government institutes have complete control over the production of diphtheria serum, in Germany it is manufactured by private concerns, but under government supervision.

The serum must be absolutely clear, free of bacteria and toxins, especially tetanus toxin, and must not contain more than  $1/2$  per cent. of phenol. It should contain at least the number of antitoxin units designated by the factory.

In the United States the standard antitoxin is distributed by the Public Health and Marine Hospital Service Laboratories. Since 1902 the production and sale of diphtheria antitoxin has been regulated by law.

At frequent intervals, antitoxin is bought in the open market and examined at the hygienic laboratories of the United States Public Health and Marine Hospital Service. Antitoxic serum containing less than a hundred units to each cubic centimeter is precluded from sale.

### The Serum Therapy of Diphtheria.

In man the antitoxic diphtheria serum is used with success for both curative and prophylactic purposes.

For therapeutic application it is of the greatest importance to employ the serum in sufficient quantities and as soon as possible. The value of early intervention can be seen from the following chart of Kossel:

*Large doses of antitoxin should be administered right from the start.* The old practice, still employed by few, of using small doses is to be condemned, for the aim in the treatment is to neutralize as soon as possible all the free and partly bound toxin.

Day of illness	Treated	Cured	Died	Percentage of cures
1	7	7	0	100
2	71	69	2	97
3	30	26	4	87
4	39	30	9	77
5	25	15	10	60
6	17	9	8	47
7-14	41	21	20	51
Indefinite	3	2	1	.....
	233	179	54	77

According to the researches of Doenitz, more recently confirmed and extended by Fritz Meyer, it was established that large amounts of antitoxin can even neutralize toxin already attached to the tissue cells. Men with practical experience like Heubner give 4000 units as the initial dose. In the United States doses as high as 10,000 to 100,000 I. E. have been administered with good results. The view of large dosage is being gradually taken up also in Germany. At any rate it is by far better to give too much than too little. If the first injection does not suffice it should be repeated the next day. The only possible drawback associated with the use of excessive amounts is the possibility of *serum sickness*, to be mentioned later. Netter has found that the administration of 1 gm. of calcium chloride on three successive days prevents serum sickness.

The serum has as a rule been injected subcutaneously. This method is very practical and as far as anaphylaxis is concerned, is the least dangerous. The disadvantage, however, is that it is very slowly absorbed. Madsen and Hendersen-Smith have shown that but a trace of antitoxin can be found in the blood of the patient four and three-fourth hours after the injection, and only after two to three days can larger amounts be demonstrated. In view of this, Morgenroth recommends the *gluteal intramuscular injection*, for here a much more rapid absorption follows. In cases of dangerous illness intravenous injection may be undertaken. For this purpose Meyer advises a serum free of carbolic acid, although this is not absolutely necessary.

The importance of the method of injection is clearly shown by the comparative experiments of Berghaus. In order to save a guinea-pig injected with a definite amount of toxin and followed in 1 hour by antitoxin, it was necessary to employ:

0.08 I. E. by intracardial injection (I. E.=antitoxin unit).

7.0 I. E. by intraperitoneal injection.

40.00 I. E. by subcutaneous injection.

Thus the curative power was increased 500 fold by placing the antitoxin directly into the circulation.



The treatment of diphtheria must by no means be limited to serum therapy. A symptom of grave prognosis is the lowered blood pressure which must be counteracted by infusions of  $1/2$  liter of physiological salt solution containing five to six drops of adrenalin.

The question whether the use of concentrated antitoxin is therapeutically more efficient than the non-concentrated is still a matter for discussion. Numerous authors claim that sera of medium strengths (about 400 I. E.) are most efficient. The highly concentrated sera are much more expensive.

For *prophylactic purposes* 500 to 1000 units injected subcutaneously usually suffice. Protection thus attained lasts about three weeks.

## CHAPTER VIII.

### TOXIN AND ANTITOXIN (*continued*).

#### DEFINITION OF TOXIN, TETANUS TOXIN, BOTULISM TOXIN, DYSENTERY TOXIN, STAPHYLOLYSIN.

The diphtheria toxin and its antitoxin just discussed in detail is of great practical and theoretical importance, and can serve as a type of all true toxins and antitoxins. *Bacterial toxins can be defined as poisons given off by the bacteria, the symptoms resulting from their action appearing after a certain incubation period. The invaded organism reacts by the production of specific antitoxins which neutralize the toxins in amounts, following the law of multiple proportions.*

Further analysis of this definition indicates that a substance can be considered a toxin only when it has a poisonous action, or in the words of Ehrlich when it possesses a toxophore group.

This toxicity does not always manifest itself by necrosis or death as in diphtheria. More frequently the toxin has a somewhat selective action affecting a certain group of organs. Thus a toxin acting upon the central nervous system or blood is designated respectively as a neurotoxin or a hemotoxin. To differentiate a true toxin from other poisonous products obtained from bacteria, it is important to note that all true toxins are elements of *secretion* of the living bacteria, and can be separated from them by filtration. According to this definition, poisons contained within the bacterial bodies themselves, which may be liberated by various mechanical, physical, or chemical means, cannot be considered as belonging to the class of true toxins. These poisons are characterized by peculiar properties and are known as *endotoxins*. In addition it may be remarked that inasmuch as a true toxin requires a period of incubation in order to manifest its action, those toxins which act spontaneously are to be excluded from the former group. R. Krause nevertheless considered some of the poisons isolated from the cholera and cholera-like spirilla (*El Tor Vibrio*) as true toxins even though they lack an incubation period.

The real essential property of a toxin is doubtlessly that one can immunize against it, and be able to demonstrate the presence of antitoxins within the serum of the immunized animal. Ehrlich claims furthermore that the amount of antitoxin produced follows the law of multiple proportions. By this is meant that the relationship between a definite dose of toxin and the amount of antitoxin just sufficient to neutralize it, is constant; so that if ten volumes of toxin hold in bounds ten volumes of antitoxin,



100 volumes of toxin neutralize 100 volumes of antitoxin. This relation is best exemplified by the diphtheria toxin and antitoxin. With the other toxins, conditions are more complicated so that many objections have been raised against the above rule of multiple proportions. (Bordet, Arrhenius, Madsen, etc.)

The true toxins causing infections in man are the

1. Diphtheria toxin.
2. Tetanus toxin.
3. Botulism toxin.
4. Dysentery toxin.
5. Staphylolysin and similar bacterial hemotoxins.

### Tetanus Toxin.

The tetanus toxin is found within filtrates of bouillon cultures of the tetanus bacillus. While partial erobiosis does not entirely eliminate toxin formation, anerobic conditions are by far more favorable for it. The tetanus toxin is of two kinds; the tetanospasmin, and tetanolysin; the former a neurotoxin, the latter a hemotoxin. The tetanospasmin is the more important of the two for the reason that it is the agent which produces convulsions. If susceptible animals such as mice or guinea-pigs are injected subcutaneously or intramuscularly with tetanus toxin, after a certain interval—the incubation period—they will begin to show symptoms due to tetanospasmin. They become hypersensitive to reflex stimulation; clonic convulsions and toxic rigidity of the muscles set in. In animals the spasms appear first in the group of muscles nearest the point of injection, while in man they almost regularly start in the muscles of the lower jaw. By intravenous and intraperitoneal injections, the tetanic spasm appears simultaneously in all muscles of the body; on intracerebral inoculation, Roux and Borrel describe the occurrence of epileptiform seizures, polyuria and certain motor disturbances—the entire set of symptoms being known as cerebral tetanus. Rabbits receiving very small amounts of tetanus toxin intravenously die after gradual emaciation and marked cachexia. This type of infection is designated by Doenitz as tetanus sine tetano. If taken per os, tetanus toxin manifests no poisonous effects. Tetanospasmin is a distinct nerve poison especially affecting the central nervous system.

Experiments by Wassermann and Takaki have demonstrated a close affinity existing between the tetanus toxin and certain organs. These organs differ in different species of animals. Thus in man, horse, and guinea-pig only the central nervous system, while in rabbits in addition to this, also the liver and spleen take up the tetanus poison. If an emulsion of brain tissue and a fatal dose of tetanus toxin are mixed and the mixture injected into mice, the latter remain unaffected. According to Doenitz

only the gray matter and not the white substance of the brain possesses this absorption power. If the brain emulsion is boiled, it loses this affinity for the toxin.

Concerning the way by which the toxin reaches the central nervous system, opinions vary. Most writers, especially Meyer and Ransom, consider that the journey is made along the nerve paths. Zupnik on the other hand believes that it is distributed through the blood stream and is taken up not only by the nervous system, but also to a great extent by the muscles.

That tetanus toxin is very labile is well known. According to Kitasato, five minutes at 65° C. or twenty minutes at 60° C. is sufficient to weaken the toxicity to a great extent, in fact almost to destroy it. Light has a similar effect upon it. Careful as its preservation may be, the soluble tetanus toxin soon becomes attenuated. Hence the best way of keeping it in stock is in a dry form. For estimating the strength of the toxin white mice are employed and are injected subcutaneously with fresh soluble toxin, the lethal dose being the amount which kills the animals in four to five days. Animals more susceptible than mice are horses, they being twelve times as sensitive and guinea-pigs six times as much. Hens possess greater power of resistance, being 30,000 times less susceptible to the toxin than mice.

*Tetanolysin* acts upon the red blood cells and disintegrates them. The erythrocytes of goats, sheep and horses, are best suited for experiments to demonstrate this action. Ehrlich showed that the tetanolysin and the tetanospasmin are really two distinctly different toxins and not one toxin with a twofold function. When tetanus poison is mixed with red blood cells the tetanolysin is absorbed and the tetanospasmin remains free. Even the antitoxins of these two are different.

As far as the standardization of the tetanus serum is concerned, it follows along the same lines as the diphtheria serum, *i.e.*, the L + dose of toxin being the one employed.

"In America the method of standardization was regulated by a law passed in July, 1908, based upon the work of Rosenau and Anderson at the United States Hygienic Laboratories at Washington. Their unit of antitoxin is ten times the smallest amount of serum necessary to save the life of a guinea-pig for ninety-six hours, against the official unit of standard toxin. This toxin unit consists of 100 minimal lethal doses of a precipitated toxin preserved at the hygienic laboratory of the Public Health and Marine Hospital Service. At the hygienic laboratory at Washington a standard toxin and antitoxin are preserved under special conditions, and standard toxin and antitoxin, arbitrary in their first establishment, are kept constant by being measured against each other from time to time. For details of this standardization the original article in the United States Hygienic Laboratory Bulletin 43, 1903, should be consulted."



Serum  
Therapy of  
Tetanus. Also in regard to the efficiency of serum therapy in tetanus opinions differ. There is, however, no doubt that a certain amount of reliance can be placed upon this treatment. Failures in successful application are ascribed to the different paths by which the toxin and antitoxin travel. The former is carried by the nerve fibers, the latter by the blood stream. Thus the serum instead of being given subcutaneously, as is the general rule, is administered by intraneural, intracerebral, and subdural injections. 100 to 200 units should be injected subcutaneously at the site of the infection or its vicinity and in addition the nerve fibers supplying the infected region should be exposed and inoculated with moderate doses of antitoxin at various points along their centripetal course.

The prophylactic use of tetanus serum has met with better results. Behring advises the administration of ten to twenty antitoxin units subcutaneously. Calmette sprinkles upon the open navel at birth a powder made of dried serum as a prophylactic against tetanus neonatorum. Bockenheimer advises an ointment containing the antitoxin as a dressing for suspicious wounds.

Botulism  
Toxin. *The Botulism toxin* is the poison produced by the bacillus botulinus. This is the exciting agent of a type of meat and sausage poisoning described by van Ermenghem in 1896 as Botulism. The bacillus botulinus is a very actively motile anerobic bacterium which grows at room temperature and presents marked gas and toxin formation. A medium in which the toxin is readily produced consists, according to Ermenghem, of an alkaline bouillon made in the form of an infusion from ham with the addition of 1 per cent. of glucose, 1 per cent. of peptone and 1 per cent. of sodium chloride.

The toxin can be demonstrated after 3 weeks of growth, and is then obtained by bacterial filtration. The cultures have a sour odor like butyric acid. The toxin deteriorates easily when exposed to air and light. It is therefore preserved in brown, sealed vials, and kept on ice; or in a dried form in vacuum. Heating the toxin for three hours at 58° C. or one-half hour at 80° C. destroys its toxicity.

Acting unrestrained, the botulism toxin is one of the severest of poisons. It affects susceptible animals even in minutest doses. In contradistinction to other toxins it is fatal even when taken per os.

The characteristic symptoms produced by botulism intoxication consist of hypersecretion of mucus from the mouth and nose, paralysis of eye muscles, urine retention, obstipation, dysphagia, aphagia, and aphoria. No fever, nor any sensory disturbances are in evidence. Death takes place because of bulbar paralysis accompanied by respiratory and cardiac failure.

The poison is absorbed or arrested in the central nervous system. 0.1

c.cm. of an emulsion of central nervous tissue neutralizes three times the fatal dose for mice. Lecithin, cholesterin, as well as fatty substances like butter and oil, act in a similar manner.

Monkeys, rabbits, guinea-pigs, mice and cats are susceptible to the toxin.

Cats usually exhibit the most characteristic clinical picture. Localized and almost pathognomonic paralyzes occur in the form of prolapse of tongue, marked mydriasis, aphonia, aphagia, etc.

In mice, paralysis of the hind extremities sets in after quite a small dose; and death follows in a few hours.

In rabbits and guinea-pigs, moderate doses (0.0003-0.001 c.cm.) occasion no manifestations during the first two to three days, but subsequently, the above-mentioned paralyzes arise and in several hours the animals expire. With larger doses (0.1 to 0.5 c.cm.) the incubation period lasts only a couple of hours and then dyspneic attacks usually succeeded by motor paralysis and death are the consequences.

The strength of the botulism toxin is ascertained by injecting guinea-pigs subcutaneously and observing the time when loss in weight, flabbiness of abdominal muscles and death occur.

The following chart by Madsen exhibits the above principle. (+ means "death.")

Dose in c.cm.	Result	Dose in c.cm.	Result
0.0015	+ on 1st. day	0.0009	Weakness in 3 weeks.
0.0015	+ after 1 1/2 days	0.0009	Loss in weight.
0.0015	+ after 2 days	0.0009	Loss in weight.
0.0013	+ after 2 days	0.0007	Loss in weight in 2 weeks.
0.0013	+ after 5 days	0.0007	Loss in weight in 1 week.
0.0013	+ after 6 days	0.0007	Loss in weight in 1 week.
0.001	+ after 4 days	0.0005	Loss in weight in 1 week.
0.001	+ after 5 days	0.0003	Practically no symptoms; only several days of weakness.
0.001	+ after 5 1/2 days		

Kempner immunized goats against botulism toxin and proved the presence of antitoxins within their sera. Immunization of rabbits and guinea-pigs is only feasible if primary inoculations are made with a toxin previously attenuated by heat for one-half hour at 60° C.

Recently Wassermann has immunized horses against this toxin. *In view of the high mortality and lack of any other specific medication, the use of this serum is strongly advised.* In animal experimentation it shows itself of undeniable value. As for its effects in man, it has not been employed frequently enough to judge.

The botulism toxin and antitoxin unite only very slowly. Otto and Sachs have shown that the inoculation of rabbits with a three hours old mixture of toxin and antitoxin occasioned greater toxic effects when administered intravenously than when given subcutaneously. Only in mixtures twenty-four hours old was this difference overcome.



*Dysentery toxin* was first demonstrated by Conradi. Subsequently from experiments by Rosenthal, Todd, Kraus and Doerr, etc., it became evident that this was a true toxin and not an endotoxin as was originally considered. Only the Kruse-Shiga type of bacillus forms a toxin; for the Flexner type, no definite toxin has as yet been isolated. Recent investigators, however, especially Kraus and Doerr, are inclined to consider the human dysentery of the Kruse-Shiga origin in the light of an intoxication or toxemia similar to diphtheria. The lesions in the large intestine where the bacteria accumulate can be compared to the diseased diphtheria tonsils, while the other manifestations, as the cerebral symptoms, cardiac disturbances, nervous sequelæ, eye affections, etc., can be taken as expressions of the toxemia.

Like the other described toxins, the dysentery toxin can be obtained by filtration of bouillon cultures. The meat infusion must be quite alkaline. The optimum alkalinity, according to Doerr, is obtained by adding 0.3 per cent. soda to litmus neutral bouillon. The precipitate thus formed which increases on sterilization should not be removed by filtration. Doerr also advises finely powdered chalk (20 gms. pro liter) to be added to the weakly alkaline bouillon before the last sterilization. The toxin is formed very gradually; the maximum is derived after two to three weeks. The gray white pellicle upon the surface of the culture can be taken as an indicator for the amount of toxin present.

According to Kraus a good dysentery toxin can also be made by emulsifying the bacteria (grown upon agar) in physiological salt solution and filtering through Reichel filters.

The toxicity of individual strains of dysentery bacilli varies greatly.

The strength of the toxin is diminished by heating for one to two hours at 60° C. Higher temperatures destroy it: 80° C., in three minutes and 90° to 100° C. in one minute.

Acids destroy the toxin probably by the formation of a non-poisonous compound. The addition of a strong alkali restores the toxicity.

Its preservation can be accomplished in a fluid state under the cover of toluol.

The action of dysentery toxin can best be studied by its effect upon rabbits after intravenous inoculation. Large doses kill the animals in very short time, six to seven hours. The ordinary lethal dose produces characteristic symptoms consisting of paresis, diarrhea, which may be bloody, paralysis of the bladder, hypothermia, etc. Death takes place in three to four weeks.

Given subcutaneously, or intraperitoneally, the toxin has only a very mild action. The incubation period is especially prolonged. Given per os, no effect is in evidence.

Besides rabbits the other susceptible animals (to large doses) are monkeys, cats and dogs; chickens, pigeons and guinea-pigs are, in the opinion of Kraus and Doerr, not at all affected by the toxin.

The intestinal changes found at post mortem examination of the animals very closely simulate the pathological alterations occurring in man. A hemorrhagic necrotic enteritis is present which in rabbits is regularly localized in the appendix and cecum, while in dogs the entire intestinal tract and especially the duodenum is attacked, and in monkeys the lower part of the intestine is involved.

The associated nervous manifestations are, according to experiments of Dopter, referred to changes in the spinal cord itself. These are of a nature similar to acute anterior poliomyelitis. Occasionally a polio-encephalitis is added.

*An antitoxic dysentery serum* is obtained by immunization of horses and goats. Various methods have been employed to obtain it. Of the older authors, Shiga and Kruse immunized animals with dysentery bacteria and thus produced a serum which possessed besides its bacteriolytic and agglutinating properties also a weak antitoxic action. Rosenthal, Todd, Kraus and Doerr employed the toxin itself for immunization purposes.

In standardization of the serum the properties to be determined are three. [Kraus and Doerr employ rabbits in this work.]

1. *Its power of neutralizing toxin in vitro.*—Toxin and antitoxin are mixed in various proportions; the mixtures allowed to stand fifteen minutes at room temperature and then injected intravenously.

2. *Its power of neutralizing toxin in vivo.*—The toxin is injected into the right vein and the antitoxin at the same time into the left vein.

3. *Its curative power.*—The antitoxin is injected at various intervals after the toxin.

These three therapeutic factors do not appear simultaneously. The power of neutralization in vitro is first in evidence. Only very much later does the serum develop its curative strength and ability to neutralize in vivo.

In animal experimentation, the antitoxic serum exhibits its neutralizing and curative properties only if injected intravenously.

Dysentery serum has been employed with fairly good results. Only infections caused by the Shiga-Kruse bacilli can, however, be benefited. The serum should be given subcutaneously and as early in the stage of the disease as possible. The dose advised by the authors varies greatly, on account of the difference in strength of the numerous sera and the severity of the infection. In cases of moderate illness, it is as a rule sufficient to give one to two injections of 20 c.cm. of a strong antitoxic serum which can neutralize toxin both in vivo and in vitro. Vaillard and Dopter have injected as many as 80 to 100 c.cm. in the severer cases.

The good effect of the serum manifests itself by an improvement in both the general and local symptoms. If high fever exists, the temperature sinks. If collapse temperature is present, it usually rises. The subjective complaints, especially the sleeplessness, improve. The blood



in the stools disappears; the movements of the bowels become less frequent and the severe pains are absent. Finally, the consistency of the stools changes and at the end becomes normal.

Prophylactic use of the serum has met very favorable confirmation in the work of Kruse, Vaillard and Dopfer, and Rosculet. Rosculet's statistics are especially interesting. In 1905 during a dysentery epidemic in Roumania, Rosculet injected eighteen apparently healthy individuals living at the homes where dysentery cases existed, with 5 c.cm. of the serum. Eighteen similar patients were removed from the dysentery surroundings, but received no serum. The results were that of the first group no fresh cases of infection arose, while of the control group fourteen became infected.

It is rather premature to determine definitely the value of the dysentery serum therapy; enough has been seen, however, to advocate its use whenever possible.

*Staphylolysin, or Staphylohemotoxin.*—According to the experiments of M. Neisser and Wechsberg the pyogenes staphylococci produce a typical hemolysin which is identical for both the aureus and albus cultures. By immunization with this hemotoxin, an antihemotoxin (antilysin) is obtained. Neisser and Wechsberg further discovered that human serum and serum of certain animal species normally contained antistaphylolysin; less, however, in amount than immune sera. Working on the principle that in staphylococcus diseases, a hemotoxin is formed which incites the development of antihemotoxin for the protection of the animal, Bruck, Michaelis and Schulze attempted to employ the presence of antistaphylolysin in the serum as evidence of the existence of staphylococcus infections.

As staphylolysin, a twelve to thirteen day old bouillon culture of freshly isolated staphylococcus pyogenes serves very well. This can be preserved by adding 5 c.cm. of the following mixture to 100 c.cm. of the bouillon filtrate: 10 carbolic, 20 glycerin, 70 aqua. The hemotoxin content is approximated according to the following scheme:

Amount of filtrate	Fresh rabbit blood	Physiological NaCl solution	Result of hemolysis after 2 hours in incubator at 37° C. and 24 hours in ice box
0.2 c.cm.	1 drop.	up to 2 c.cm.	complete.
0.1 c.cm.	1 drop.	up to 2 c.cm.	complete.
0.05 c.cm.	1 drop.	up to 2 c.cm.	complete.
0.025 c.cm.	1 drop.	up to 2 c.cm.	complete.
0.01 c.cm.	1 drop.	up to 2 c.cm.	incomplete.
0.005 c.cm.	1 drop.	up to 2 c.cm.	layer of red blood cells.

Thus 0.025 is the smallest dose which can completely hemolyze the given quantity of red blood cells.

The amount of antilysin is estimated by adding varying amounts of serum to the constant minimal hemolytic dose of the staphylolysin and determining what amounts of serum contain enough antilysin to prevent hemolytic action of the staphylolysin. It is best to allow the staphylolysin and serum to remain mixed for some time before adding the rabbit blood, so as to give the antitoxin a chance to neutralize the toxin.

As every normal serum contains a certain amount of antilysin, it is necessary in order to obtain the pathological variations, to use a normal serum as a control. Such a serum, 0.1 c.cm. of which just suffices to neutralize twice the minimal hemolytic dose, was dried in vacuum and used by Bruck, Michaelis and Schulze, as standard serum.

Estimation of antilysin content of the standard serum:

Twice the minimum hemolytic toxic dose.	0.05	0.05	0.05	0.05	0.05
Standard normal serum.....	0.2	0.1	0.05	0.025	0.01
Result after 24 hours.....	No hemolysis.	No hemolysis.	Slight hemolysis.	Moderate hemolysis.	Complete hemolysis.

These mixtures were allowed to stand for one hour at 37° C. and then 1 drop of rabbit's blood was added, allowed to remain for two hours at 37° C. and twenty-four hours in the ice box.

The standard serum was always freshly prepared in the form of a 10 per cent. solution in distilled water (0.1 : 1).

In the above manner the antilysin content of the serum from patients with distinct or suspicious staphylococcus infections was estimated. The completely neutralizing dose of the standard serum (0.1 above) was taken as 1 and the neutralizing dose of the serum for examination compared with this; if 0.05 c.cm. of a serum x neutralized the same amount of toxin as 0.1 of standard serum, the antilysin value of the serum x was 2.

From the comparative studies of Bruck, Michaelis and Schulze it was concluded that most of the normal sera had values ranging from 1 down; occasionally results as high as 5 were obtained. *Out of twenty-five cases of staphylococcus infections nineteen gave values varying from 10 to 100. Figures as high as these can, according to these authorities, become of valuable aid in diagnosis.*

Although these findings were corroborated by Arndt and others, the method cannot as yet be classed among those of clinical diagnostic importance. Similar study of other infections has not been undertaken.



In addition to the toxins reviewed in these chapters, recent work has proven that toxins may, under certain conditions, be derived from bacteria other than those mentioned, *e.g.*, cholera, typhoid bacteria and meningococci. Problems such as these are still, however, open to scientific discussion; consequently no exact statements can be made here.

## CHAPTER IX.

### THE TOXINS OF THE HIGHER PLANTS AND ANIMALS AND THEIR ANTIBODIES. FERMENTS AND ANTIFERMENTS.

The toxins thus far studied were all secretory products of bacteria. This power of forming toxins is not, however, limited to bacteria alone, as there is a class of higher plants and animals that produce characteristic poisons against which immunization can be undertaken and an antitoxic serum obtained. Aside from snake poison, the members of this group bear little practical medical interest. The detailed study of these plant toxins (Phytotoxin) and those of animal origin (Zootoxin) has, however, greatly increased the theoretical knowledge of the phenomena of reaction and immunity.

#### Phytotoxins.

The most important phytotoxins are:

1. Ricin.
2. Abrin.
3. Crotin.

Ricin is a deadly poison, of which the smallest fractions of a milligram are sufficient to kill rabbits. Like bacterial toxins, ricin requires for its action an incubation period of at least twenty-four hours. The typical postmortem findings consist of redness and swelling of Peyer's patches. Ricin is a hemotoxin; if mixed, as an emulsion, with red blood-cells, the erythrocytes sink to the bottom and are agglutinated.

Ehrlich succeeded in immunizing animals against ricin by first giving it to them per os in increasing doses for a long period of time, and later on by subcutaneous injection. The antitoxic serum thus produced neutralizes the poisonous action of ricin both in vivo and in vitro.

Abrin, a vegetable poison, is obtained from jaquiritia (*Abrus precatorius*) and in its action closely resembles ricin, but is less poisonous. It is a marked irritant of the conjunctiva and was at one time employed in cases of trachoma.

Roemer found that by repeated instillation of abrin into the same conjunctival sac, no reaction was ultimately obtained (local immunity), while the conjunctiva of the other eye retained its susceptibility. If the instillation was continued for a long period of time, a "general immunity" was attained which extended to the conjunctivæ of both eyes. As a result, in the serum of such animals antiabrin could be demonstrated.



Croton is the seed of *croton tiglium*, a substance less poisonous than either ricin or abrin. It does not agglutinate, but produces hemolysis of rabbits' red blood cells. Toward the red blood cells of other species (*e.g.*, bird), it is entirely inactive. The immunization of rabbits is readily brought about by subcutaneous injections. Their serum neutralizes the hemotoxic action in vitro.

### The Zootoxins.

Most important of the animal toxins are

- |                                  |                                  |
|----------------------------------|----------------------------------|
| 1. Phrynylysin (toad poison),    | } Simple hemotoxins.             |
| 2. Arachnolysin (spider poison), |                                  |
| 3. Snake poison,                 | } Lecithin producing hemotoxins. |
| 4. Scorpion poison,              |                                  |
| 5. Bee poison,                   |                                  |

The one striking characteristic of toxins, that an immunity can be raised against them, is also possessed by these poisons. Aside from this fact they present many variations from the true class of toxins. Most of these poisons are complex, *i.e.*, they contain more than one toxin and all are hemotoxic.

Toad poison is obtained by rubbing up the skins of the *Bombinator igneus*; spider poison by trituration of the living "cross spiders" (*Epeira diadema*) in three or four times the amount of physiological salt solution containing toluol.

The toad and spider poisons contain simple hemotoxins, that is to say, by the mixture of small amounts of this toxin with erythrocytes absolutely serum-free, hemolysis of the latter takes place. Not all species of blood are affected alike. The red blood corpuscles of sheep, goats, and rabbits are especially adapted for experiments with phrynylysin, while rabbits', rats', and human blood is more suitable for arachnolysin. Immunity of rabbits is easily attained.

### Snake Poisons.

The most familiar poisonous snakes are the Cobras (*Naja*) of India and Indo-China which belong to the family of Colubridæ, the European viper, and the American rattlesnake; the last two being of the Viperidæ species. The poisons of these two families show great individual differences. Thus, those of the Colubridæ group are decidedly thermo-resistant (temperatures as high as 100° C.) while the viper's poison is entirely destroyed at a temperature varying between 80 to 85° C., and markedly weakened at 70°C.

Snake poisons, as a rule, produce both local reactions at the point of the bite, and severe general disturbances.

The cobra bite is only slightly painful. A characteristic feeling of stiffness extends from the point of infection over the entire body. In several hours a rapidly increasing weakness sets in terminating in deep coma and death.

The viper bite incites a very severe local reaction. The point of infection is red, extremely painful and swollen. Convulsions, hemorrhages, followed by delirium which finally changes into stupor are manifest, and death takes place in one to three days. If the poison gets into the circulation directly, death is likely to occur in a few minutes.

The prognosis of a snake infection depends largely upon the situation of the bite. The greater the blood supply of the infected area the more dangerous is the result. Bites received through the clothing are relatively less dangerous, as a great part of the poison remains adherent to the clothing.

Snake poisons act primarily upon the nervous system and blood, although they exhibit a number of other toxic and ferment properties. Thus viper toxin occasions immediate coagulation of the blood by its action upon the vascular endothelium and has for this reason been called by Flexner and Noguchi, "Hemorrhagin."

*Furthermore, all snake poisons have a hemolytic power.*

Cobra hemolysis represents one of the most interesting of Cobra biological phenomena, and since it may possibly be employed Hemolysis. in clinical methods of examination its action will be here reviewed.

Cobra hemotoxin is characterized by its power of dissolving the red blood corpuscles of certain kinds of animals (ox, sheep and goat) only in the presence of serum. Other red blood cells do not require any serum for their hemolysis (dog, guinea-pig, man, rabbit, horse). If the red blood corpuscles of the first group of animals washed free of their serum are mixed with cobra poison, no hemolysis takes place. On subsequent addition of any fresh serum, hemolysis is in evidence. (Flexner, Noguchi.)

The agent which activates the hemolytic substance belongs undoubtedly to the class of lipoids. Of these, lecithin stands pre-eminent. It is, however, by no means certain whether that is the only or the most important activator.

Some sera exhibit this activating influence only when first heated. In their unheated state they are entirely inactive. Other sera act in a manner decidedly the reverse. Kyes and Sachs mention that this depends altogether upon the nature of the lecithin union. The following table shows the various combinations and their resultant action.



Combination		Power of serum to activate the hemolysis		
Serum	Red cells	If serum is unheated	If serum is heated at	
			56° C.	65 to 100° C.
Horse.....	Ox.....	+	+	+
Horse.....	Horse.....	+	+	+
Ox.....	Horse.....	+	—	+
Ox.....	Ox.....	—	+	+
Sheep.....	Ox.....	—	+	+
Sheep.....	Sheep.....	—	—	+ weak hemolysis.
Human.....	Ox.....	—	—	+
Human.....	Human.....	+	+	+
Rabbit.....	Ox.....	+	+	+
Guinea-pig.....	Ox.....	+	—	+
Guinea-pig.....	Rabbit.....	+	—	+

+Signifies hemolysis.

—Signifies no hemolysis.

### Cobra Hemolysin Test.

1. *Washing of Erythrocytes.*—The blood is collected into sterile flasks containing sterile glass beads. It is shaken and thus defibrinated to prevent coagulation. The defibrinated blood is next centrifugalized and the serum separated and drawn off with a pipette. The red blood cell sediment is mixed with physiological salt solution and again centrifugalized. This procedure is repeated several times until all the serum is removed. The red blood cells are used in a 5 per cent. suspension; *i.e.*, 1 part of washed erythrocytes suspended in 19 parts of saline.

2. *The Activating Agent.*—As an activating agent 0.2 c.cm. of serum or a 0.1 per cent. lecithin solution is employed. The lecithin can be kept as a stock solution consisting of 1 g. lecithin in 100 c.cm. of methyl alcohol. A 0.1 per cent. solution of the stock mixture is made by mixing 0.1 c.cm. of the solution with 9.9 c.cm. of physiological salt solution.

3. The snake poison hemotoxin is resistant toward heat so that it may be heated to almost 70° C. without interfering with its activity. Cobra poison contains the greatest amount of hemotoxin. While 1 mg. of cobra toxin hemolyzes 1 c.cm. of 5 per cent. horse's red blood cells in five to ten minutes, a similar amount of viper toxin requires thirty minutes for the same action.

V. Dungern and Coca explain this type of hemolysis by the existence of a ferment within the snake poison which breaks up the lecithin with the liberation of oleic acid. This acid has long been known as a hemolytic agent. The necessity for adding lecithin or serum to certain species of blood is explained by the variability in the lecithin content of the erythrocytes, or a variability in the lecithin union.

Cobra Toxin Activation in Tuberculosis. Calmette notes in the blood of tuberculous patients more than the normal amount of lecithin; for that reason their serum can be used in very small doses to activate the cobra hemolysin. By this means he attains a diagnostic reaction for tuberculosis. On examination of the blood of 177 tubercular individuals he has found:

78 per cent. of positive reactions in the first stage of tuberculosis.

57 per cent. of positive reactions in the second stage of tuberculosis.

70 per cent. of positive reactions in the third stage of tuberculosis.

Szaboky has confirmed these findings, but not enough control examinations of normal individuals or of other infections had been made to firmly establish the diagnostic value of the test.

At the instigation of the author, Alessandrini repeated the work of Calmette but came to different conclusions. The reaction was not found specific for tuberculosis; it was given by various other diseases. Furthermore Alessandrini could not confirm the hypothesis that hemolysis was dependent upon the lecithin or lipid content of the serum. By simple mixture of cobra poison and horse's erythrocytes, he found that the resistance of the cells toward any traumatism becomes greatly diminished so that hemolysis can be attained even by hypotonic salt solutions in concentrations which under normal circumstances are entirely ineffective for horse's red cells.

The hemolysis of snake poison can be overcome or interfered with by the addition of large amounts of normal serum, cholesterin, and small amounts of snake poison serum.

Much and Holzmann have recently described the so-called "Psycho-The Psycho-reaction" which can be explained thus—Normal serum, when added to a mixture of cobra extract and human red blood-cells will not interfere with consequent hemolysis. If, however, the serum is obtained from patients suffering from depressive mania, circular insanity or dementia præcox, and added to the cobra extract and human red blood-corpuscles, the expected hemolysis does not take place. One would naturally suppose that this fact would be employed for clinical diagnosis, but unfortunately it has been generally proven by most authorities that it is altogether impossible to do so for the simple reason that it is not absolutely specific. Bauer has found the same reaction with navel blood. It is probable that the interference with hemolysis is brought about by an increase in the cholesterin of the serum—a greater possibility in diseases of the central nervous system than in any physiological or other pathological condition.

Cobra Immunity. In immunizing laboratory animals one cannot start with inoculations of the unaltered snake poison.

Phisalix and Bertrand begin with subcutaneous injections of a toxin heated to 75° C. and after two days use one-half of the minimal lethal dose of the unaltered toxin.

Calmette weakens the cobra poison by the addition of an equal amount of 1 per cent. gold chlorid, and after four such injections with increasing amounts at each time, the pure toxin in very small doses is employed.



In the same manner Calmette immunized horses and obtained highly antitoxic sera. He tested the strength of these sera, as follows:

1. *Upon Rabbits*.—Each animal received an injection of 2 c.cm. of the serum into the vein of one ear, and after two hours 1 mg. of toxin into the vein of the other ear. A control animal was similarly treated with toxin only. The latter animal died in a half hour, while the former remained alive.

2. *Upon White Mice*.—Diminishing amounts of serum were mixed in test tubes, with 0.0001 gm. of toxin (in 1 per cent. solution) and the mixtures injected into the mice. The amount of serum which should completely neutralize the toxin must not be more than 0.03 c.cm.

According to Calmette one can judge the efficiency of an immune serum by its antihemolytic power, inasmuch as the hemotoxic and neurotoxic actions run parallel. This is denied by Noguchi.

The scorpion and bee poisons display properties similar to those of the cobra poison. They also combine with lecithin to produce hemolysis.

Thus far, it has been shown that the lipoids, especially lecithin are actively associated in the hemolysis of erythrocytes; whether the toxin combines with the lipoids and forms a toxolipoid (toxolecithid) which is hemotoxic, or whether as v. Dungern believes, the hemolytic action is due to the fatty acid derived from the lecithin by the ferment action of substances contained in the poison, has not been definitely proven.

Quite recently it has been thought that pernicious anemia and paroxysmal hemoglobinuria are closely associated with such toxolipoids.

Tallquist obtained from a *Bothriocephalus latus*, a hemotoxic  
Pernicious poison of a lipid nature which experimentally produced a  
Anemia. blood picture characteristic of pernicious anemia. But it  
would be incorrect to associate all forms of pernicious anemia  
with tape-worm poison; more probable is it that hemotoxins are formed  
within the organism itself.

In *paroxysmal hemoglobinuria* a hemotoxin of very peculiar  
Paroxysmal properties is found circulating in the blood.  
Hemoglo- It can be demonstrated as follows.  
binuria.

1. *Ehrlich's Method*.—One of the patient's fingers is tightened by means of a small tourniquet and kept immersed in ice-cold water for half an hour. Some blood is then collected into a capillary pipette from the finger thus tied, and as a control, blood from a finger of the other hand is drawn off. This is allowed to clot and then centrifugalized. The results are that the serum from the finger held in the ice water is tinged red from dissolved hemoglobin while the control serum is normally pale.

2. *Donath-Landsteiner's* method repeats Ehrlich's experiment in vitro.



The patient's and the control individual's serum are each mixed with washed human erythrocytes in various proportions. It does not matter whether the red blood cells are obtained from the patient or normal individual. The mixtures are allowed to remain for one-half to one hour in the ice box and then from one to three hours at a temperature of  $37^{\circ}$  C. The serum from the paroxysmal hemoglobinuria patient shows hemolysis.

A control tube containing the same ingredients, in the same proportions and maintained at *either* cold *or* warm temperatures, but not at both in succession as above, exhibits no hemolysis.

The hemolytic process in this disease is of a complex nature. In the cold, one element combines with the erythrocytes, and at high temperature another unfolds hemolytic tendencies. Some sera lacking entirely or not having enough of the second element in the serum, demonstrate no hemolysis. But on addition of some normal serum hemolysis occurs. It can therefore be concluded that the second factor which acts in the heat is present within normal serum, while the first substance, the specific one, is found only in the blood of those suffering from paroxysmal hemoglobinuria; (and according to Donath and Landsteiner in 10 per cent. of cases of general paralysis). It is, in addition, the author's opinion, that similar toxic substances exist in the blood of epileptics and idiots.

Not all cases of paroxysmal hemoglobinuria possess this characteristic hemotoxin. In some it is only found periodically.

No explanation has as yet been offered for these varying phenomena. Attempts have been made to ascertain whether the hemotoxin is stimulated by an external agent or by infection (lues, malaria, trypanosomiasis) or whether it is of endogenous origin. The answer is still for the future to disclose.

### The Antiferments.

Ferments are very closely allied to toxins in their biological structure. By the immunization of animals with ferments in as pure a form as possible, antiferments can be demonstrated. Just like antitoxins, antiferments can neutralize their respective ferments *in vitro*. As to their presence, it is quite important to know that they are found in normal serum in certain small quantities (together with antitoxins). The difference in their presence in a normal serum and that in an immune, is purely a quantitative one.

The antiferments thus far demonstrated are

Antilabferment.

Antipepsin.

Antitrypsin.

Antisteapsin.

Antifibrinferment.

*It is difficult to obtain by immunization an antiferment serum of very high*



*strength.* Probably the normal organism is so regulated that it compensates for any increased amount of antiferment.

Till recent times the demonstration of antiferments bore no clinical interest. The antibodies of the proteolytic enzymes first began to attract attention when the inhibitory influence which blood serum has upon the autolysis of organs was proven. It was Jochmann and Müller who showed in connection with their studies of the proteolytic ferments of leucocytes, that apart from these, the serum itself possesses an inhibitory influence upon the leucocyte ferment. This is found to be especially marked in diseases associated with great destruction of leucocytes. Following them, Marcus, as well as Brieger and Trebing, discovered a restraining influence in the serum upon the action of pancreas trypsin and proved that the so-called antitrypsin was considerably increased in carcinoma patients. Bergmann and Meyer, also working along these lines, then demonstrated that the wrongly called "carcinoma reaction" was by no means specific for carcinoma, but was found in a large number of other diseases. It cannot, as Brieger later announced, even be considered as a criterion for cachexia Reaction. (cachexia reaction).

Undoubtedly, the already normal antiproteolytic power of the serum can be considerably increased in animal experimentation by a group of well-known proteolytic agents, and especially by leucocyte ferment and pancreatic trypsin. To differentiate between antileucocyte and antitrypsin ferment in the narrow sense of the word, is impossible. The one "immune serum" (*sit venia verbo*, if one can speak of immune serum in this sense) neutralizes the other antigen. Clinically a high antitryptic titer of the serum is found in about 90 per cent. of carcinoma patients, and is almost regularly observed in infections with high fevers as typhoid, severe articular rheumatism, sepsis, etc. In pneumonia there is found during the infection a marked change from an excessively high to a low titer. In Morbus Basedowii (as well as in experimental thyroid feeding) it is almost the rule to find a high antitrypsin content, but one must always keep in mind that even few normal individuals show a similar increase.

The clinical diagnostic importance of the antitrypsin titer is slight in comparison with its experimental increase. In accord with the findings in Basedow's disease, and in thyroid feeding it may be considered as an outcome of increased proteid destruction (hyper-production of proteolytic ferments in the tissues?). Leucocyte ferment has been found of practical use in the treatment of cold abscesses, *i.e.*, in processes where lymphocytosis and failure to produce polynuclear leucocyte ferment is present. On the other hand antitrypsin or antileucocyte ferment or even normal serum is employed to counteract inflammatory processes, *i.e.*, to neutralize the excessive production of the leucocyte ferments, with apparent success (Leucoantifermentin, on the market). According to recent findings, the antitrypsin titer of the mother's blood increases markedly during the period of labor, while that of the fetus remains unaltered.

There are two methods for the antitrypsin determination. The first was devised by Jochmann and Müller for proving the presence of leucocyte ferment and its antiferment, and then similarly employed by Marcus in the study of pancreatic ferments. Its principle depends upon the digestive action of proteolytic ferments upon serum albumin. When a drop of trypsin is placed upon a Löffler's serum plate, after a little while, a clear spot appears where the trypsin was brought into contact with the plate.



If to this trypsin an amount of serum is previously added, which fully neutralizes the digestive action, no clear zone appears upon Löffler's plate.

The details of this procedure are as follows: The ferment solution consists of 0.1 gm. trypsin, well shaken with 5 c.cm. of undiluted glycerin and 5 c.cm. of distilled water, then left in an incubator for a half hour at 55° C., then again shaken and filtered.

The serum is mixed in small test-tubes or upon a glass slide with varying amounts of the trypsin; thus 1 loopful of serum is mixed with 1/2, 1, 2, 3, 4, etc., up to 20 loopfuls of the trypsin solution and of each of these mixtures one loopful is placed upon Löffler's plate. (Ox serum plate, should be three days old). The plates are then placed into the incubator for twenty-one hours at 55° C. The presence or absence of the clear zones determines the quantities of ferment which respectively have not or have been neutralized by the one drop of serum (e.g., 1:6 means that in the mixture of 6 loopfuls of trypsin and 1 loopful of the serum for examination the digestive power of the trypsin was still interfered with).

The inequality in the strength of the Löffler plates, their variability in the degree of alkalinity, the measurement by loopfuls, all, might prove to be sources of error which may greatly influence the results. Thus the latter can only be taken as approximate, relative values.

The second, more exact and satisfactory method was introduced primarily by Gross and Fuld for presenting the action of trypsin, and was modified by v. Bergmann together with Bamberg and Meyer for the determination of antitrypsin. Numerous workers have found it thoroughly reliable. Its principle is based on the digestion of a clear casein solution. If the entire amount of casein is digested, no more is left to be precipitated by the addition of acid and therefore the solution remains clear. If, however, casein has been left undigested, the addition of acid will produce a turbid solution or even a white precipitate.

The necessary reagents are:

1. *Casein Solution*.—One gm. of casein is dissolved under slight heating in 100 c.cm. of N/10 NaOH; this solution is next neutralized by N/10 HCl, litmus being used as indicator, and diluted with physiological salt solution up to 500 c.cm. (If sterilized, it can be kept for a long while.)

2. *Trypsin Solution*.—0.5 gm. of trypsin (purissimum Grubler) is dissolved in 50 c.cm. of normal NaCl + 0.05 c.cm. of normal sodium hydrate solution and then diluted with physiological saline up to 500 c.cm.

3. *Acid Solution*.—Five c.cm. of acetic acid + 45 c.cm. of alcohol + 50 c.cm. of water.

First the titration of the trypsin solution is undertaken in order to find out how much trypsin is required to fully digest a constant quantity of casein. Gradually increasing amounts of trypsin (from 0.1 to 0.6 c.cm.) are placed in six test-tubes and to each 2.0 c.cm. of casein are added. These tubes are placed in an incubator at 37° C. for one-half hour, and then several drops of the acid solution are placed into each tube. The first tube, and all those above it that remain absolutely clear, contain enough trypsin to fully digest the 2.0 c.cm. of casein.

Now comes the second part of the test.

In each of eight to ten test-tubes are placed 2 c.cm. of the casein solution and 0.5



c.cm. of a 2 per cent. dilution of the serum for examination; to these is next added the trypsin solution in successively increasing amounts, beginning with the smallest quantity which in the first part of the test was sufficient to completely digest the given amount of casein. Salt solution is then added to each of the test-tubes so that all contain an equal quantity of fluid, and the mixtures placed in an incubator at  $37^{\circ}$  C. for one-half hour. At the end of this time, several drops of the acid are added to each tube. Those tubes which become cloudy or show a precipitate designate the amounts of trypsin solution which have been neutralized by the 0.5 c.cm. of diluted serum. For example:

In the first part of the test it was found that the tube containing 0.4 c.cm. of trypsin was the first to remain clear, in other words was sufficient to fully digest 2 c.cm. of the casein solution. In the second part of the test the lower limit of the added trypsin dilution was 0.4, and it was found that the tubes containing 0.4, 0.5, 0.6, and 0.7 c.cm. of trypsin, for example, now gave precipitates and only 0.8 remained clear. This indicates that part of the formerly sufficient amount of trypsin was now neutralized by the antitrypsin of the added serum so that digestion was interfered with. Thus the antitrypsin titer in this case is 0.8.

Recently the above method of trypsin titration has been applied to the determination of the presence of pancreatic ferment in the intestinal secretions, feces, and stomach contents.

## CHAPTER X.

### AGGLUTINATION.

#### BACTERIAL AGGLUTININS. HEMAGGLUTININS. TRANSFUSION TESTS.

The Phenomenon of Agglutination.

If the serum from an immunized animal, or a patient convalescing after an infection, be mixed with a suspension of the bacteria which were involved in the production of said conditions, a peculiar phenomenon takes place. In the former diffusely cloudy liquid, small granules and clumps appear which sink to the bottom of the test-tube and leave a supernatant clear fluid. On microscopic examination, the sediment presents bacteria, (which have remained alive as is demonstrable by making cultures of same). This same observation can be made with perhaps more flattering results when the experiment is performed in a hanging drop. The bacteria are seen to lose their motility, adhere to each other, finally gravitate toward larger groups and arrange themselves in clumps. The phenomenon thus described was discovered by Gruber and Durham, and is called *agglutination*; while substances which cause this, *agglutinins*.

If instead of the immune serum or that of the convalescent patient, normal serum is employed and the above test repeated, it will be seen that agglutination likewise occurs. The reaction is, however, somewhat incomplete; the clumps are smaller, and formed much more slowly. *If a quantitative determination with different dilutions* of both sera is made, the power of agglutination disappears with the normal serum at a low dilution, while the immune serum remains perfectly active at even much greater dilutions. Thus the main difference between the agglutinating normal and immune serum is a quantitative one depending upon the amount of agglutinins present. Whether any qualitative difference exists between the normal and immune agglutinins is doubtful. It is, however, of no practical significance.

If instead of homologous bacteria, different (heterologous) bacteria are employed, *e. g.*, cholera vibrio and typhoid serum, agglutination also takes place, if the typhoid serum is used in concentrated or only slightly diluted form; but in moderate or great dilutions, no agglutination occurs. Normal serum will agglutinate the cholera vibrio in the same strength as the immune typhoid serum. In other words the typhoid serum contains more agglutinins for its homologous bacteria than a normal serum, but it has only the same titer of agglutination as a normal serum for heterologous bacteria.



*The agglutination reaction is specific in the respect that high dilutions of serum will agglutinate only its homologous bacteria and leave the heterologous ones uninfluenced. Agglutination becomes non-specific, when concentrated or low dilutions of serum are employed.*

The relative specificity just described is of great clinical diagnostic value. For example, given a serum suspicious of Value of Ag-typhoid, the question is to establish this absolutely. One glutination. immediately proceeds to make a suitable dilution of the unknown serum and mixes with it known typhoid bacilli. A similar dilution of normal serum is made as a control and mixed with the same amount of typhoid bacilli. If agglutination occurs with the unknown serum and not with the control serum, the former must have come from a typhoid patient. If the bacteria are not agglutinated, the serum was not of typhoid origin.

In an equal manner can the identity of unknown bacteria be established by the use of known sera. Thus, when certain bacteria have been isolated and information is wanted as to whether they are typhoid, an emulsion of these is made and mixed with a typhoid serum in suitable dilution, and a similar amount of bacteria is mixed with a normal serum of like dilution. Agglutination occurring in the first of these mixtures and not in the second proves the typhoid character of the unknown bacteria. In this manner the agglutination test can be used for identification of any antigen.

The practical application of agglutination has been greatly used in cases of typhoid fever. Here agglutinins are very easily stimulated in the course of the disease and generally they can be demonstrated in the serum seven to ten days after infection. The agglutinins remain not only during the active stage of the disease, but also during the convalescing period. Widal, the Parisian clinician, was the first to adopt this agglutination reaction for the serum diagnosis of typhoid. It is thus commonly known as the Widal reaction.

Technique of Aggluti-  
nation. The technique of the reaction is as simple as its principle. This accounts for its wide adoption. It may either be performed macroscopically or microscopically (orientation test).

### The Macroscopic Agglutination Reaction.

For this reaction it is necessary to have

1. The immune serum and a normal control serum;
2. A homogeneous bacterial emulsion.

The production of a homogeneous bacterial emulsion offers slight technical difficulties.

It can be obtained in the following ways:

*a. Bouillon Culture.*—Many bacteria, like typhoid, paratyphoid, dysentery, coli, etc., grow very easily in broth. Such a fresh (twenty-four hours), diffusely turbid culture can be employed readily for agglutination purposes. In place of live bacteria, dead may also be used—a fact which has greatly added to the practical application of the test.

For preparing the latter, 0.5 per cent. of phenol or 1 per cent. of formalin (40 per cent.) are added to the twenty-four hour bouillon cultures. The result is, that a sediment of bacteria is formed from which the supernatant fluid should be carefully poured off. The bacterial suspension is kept on ice and thoroughly shaken before use.

Ficker has in this way prepared standard emulsions of dead typhoid and paratyphoid bacilli which are sold by Merck under the name of "Ficker's Diagnosticum."

For Widal's test, a small quantity of the patient's blood is collected in a capillary tube and the end closed with sealing-wax. The blood is allowed to clot, and the serum to separate off. The separation of the latter can be hastened by centrifugalization.

In practice, the Widal test as performed with Ficker's diagnosticum, is arranged as follows:

Bacillus suspension	Dilution of serum	Physiological salt solution
Tube 1, 0.5 c.cm.	.....	0.5 c.cm.
Tube 2, 0.5 c.cm.	0.5 c.cm. of 1:10	.....
Tube 3, 0.5 c.cm.	0.5 c.cm. of 1:50	.....
Tube 4, 0.5 c.cm.	0.5 c.cm. of 1:100	.....

One of four results may be obtained.

1.	2.	3.	4.
Positive reaction	Doubtful reaction	Negative reaction	Worthless reaction
1. No agglutination.....	No agglutination.....	No agglutination.....	Agglutination.
2. Marked agglutination.	Marked agglutination	Very slight agglutination.	Agglutination.
3. Marked agglutination.	Very slight agglutination.	No agglutination.....	Agglutination.
4. Slight agglutination...	No agglutination.....	No agglutination.....	Agglutination.

It is very advisable to make control tests with normal serum. After mixture of the various ingredients the tubes are placed in the incubator at 37° C. for two hours. Then the results are read off; the first tube must show absolutely no agglutination, otherwise (as seen in Division No. 4 above) the entire test is of no significance. The cause for such spontaneous or pseudo-agglutination occurring in tube 1 may be found either in the bacterial emulsion or NaCl solution. The grade of agglutination is estimated by the size of the agglutinated clumps and the rapidity with which they are formed. The mild grades of agglutination are frequently overlooked by the beginner. For typhoid a positive reaction is one where agglutination



takes place in the dilution of 1:100; a positive result in the dilution of 1:50 can only be considered as probably positive.

As has been said, broth cultures may be used for the agglutination test if the bacteria grow diffusely and regularly within the bouillon. This is not the case, however, with all bacteria, as for example, the cholera vibrio which produces a thin pellicle upon the surface of the broth.

b. *Agar Cultures*.—Kolle and Pfeiffer have advised instead of broth the use of agar cultures. The bacteria are washed off, and an even emulsion made in physiological salt solution, or in a dilution of the serum for examination.

The details of the procedure are as follows:

Into a row of test-tubes is placed 1 c.cm. of various dilutions of the serum for examination. e.g., 1:10, 1:50, 1:100, 1:200, 1:500. A normal serum is similarly diluted as a control. One other test-tube is to contain 1 c.cm. of saline only.

A full loop of an eighteen to twenty-four hours old agar culture is evenly and finely rubbed up in each of the above test-tubes as follows:

The test-tube is held almost horizontally in the left hand between the thumb and index finger; a platinum loop between the thumb and index finger of the right hand is filled with the bacteria from the agar culture, and placed in the tube containing the serum dilutions. The bacteria are then gently and thoroughly rubbed up on the moistened wall of the tube but not within the fluid. By rolling the test-tube slightly, a part of the rubbed up bacteria is washed into the fluid and the remaining bacterial mass is again triturated. This process is repeated until all the bacteria are washed into the fluid. Thus, a homogeneous suspension is obtained.

The author has found this method of Peiffer and Kolle most accurate.

It is worthy of note in this connection, that the controls show no clumps or granules. (Pseudo-agglutination). There are some bacteria which can be evenly emulsified only with great difficulty, while others are very easily agglutinated even by normal serum. In either case the test is not conclusive.

*For the hanging-drop method*, blood is collected in a Wright capsule or a small test-tube; 6 to 8 drops of blood suffice. The blood is allowed to clot or the serum is hastened by centrifugalization. Four loopfuls of broth or saline (or equal amounts as measured by a Wright pipette) are placed on each of two slides. To one of these one loopful of the serum (or one equal part as measured by the Wright pipette) is added and thoroughly mixed. From this mixture one loopful or equal measure is mixed with the broth or normal saline upon the second slide; thus making serum dilutions of 1:5 on the first slide and 1:25 on the second slide. A loopful of typhoid culture is placed on the center of each of two cover slips. To the first is added one loopful of the serum dilution 1:25, and to the second is added one loopful of the serum dilution 1:5 thus making a dilution of 1:50 and



1:10 respectively. Each cover slip is inverted over a hollow slide protected by vaseline, and examined microscopically. A control with normal serum and also one with culture alone should be made.

For the identification of bacteria only *highly agglutinating animal sera* can be employed. Rabbits, goats and horses, of Agglutinating Sera, are most suitable for such experiments. The best results are obtained when the animals are immunized intravenously by repeated injections with gradually increasing doses of dead bacteria (killed at 60° C.). Usually two to three injections of 1/4 to 1 agar culture of bacteria suspended in saline solution suffice to give an agglutinating titer of 1 to 5000. The serum should be withdrawn eight to ten days after the last injection. With typhoid bacteria one may attain a strong agglutinating serum in a rabbit by the intraperitoneal injection of 1 c.cm. of a twenty-four hours' live broth culture. This is to be repeated in 7 to 10 days. As a matter of course, the titer of the serum should be tested from time to time, because the height of the antibody curve can only reach a certain point. When a sufficient strength is obtained the animal is bled. It is not possible to produce equally strong agglutinating sera for all bacteria.

The agglutinins belong to the class of the more resistant serum substances. By the addition of one drop of pure carbolic acid they can be preserved on ice for a long time. Heating variously affects the different bacterial agglutinins. The agglutinins for pest and tubercle bacilli are destroyed at 56° C. while other bacteria are not influenced by even higher temperatures. The animal from which the agglutinating serum has been obtained also influences to a great degree the resistance toward heat. Thus the typhoid agglutinating serum derived from the horse is much more resistant than that obtained from the rabbit.

### The Microscopic (Orientation) Agglutination Test.

This method is especially of use, when only small amounts of culture or serum are obtainable. Also, if agglutination is employed for the quick recognition of bacteria, as for example, when it is desirable to know whether a blue colony on a Conradi-Drigalski-agar plate is typhoid or not.

In such a case a drop of the immune serum in the dilution of 1:50 or 1:100 is placed upon a cover-glass held with a Cornet's forceps, and a small part of the bacterial colony for identification carefully mixed with this serum. As controls, a mixture is made with salt solution and with normal serum. If agglutination occurs, small granules or clumps can readily be seen with the naked eye by holding up the cover-glass against the light. The control glasses on the other hand should show only a homogeneous turbidity. These changes are still more evident if the mixture is examined microscopically in the form of a hanging drop. (Described, p. 108.)



### Group Agglutination.

On testing the titer of a strongly agglutinating typhoid serum, and a strongly agglutinating cholera serum, against typhoid, paratyphoid, colon and cholera bacteria, the results will be the following:

Agglutination titer	Of typhoid serum	Of cholera serum
Against typhoid.....	1 : 2000	1 : 10
Against paratyphoid.....	1 : 100	1 : 10
Against bacter. coli.....	1 : 25	1 : 10
Against cholera.....	1 : 10	1 : 3000

The cholera serum acts strictly in accordance with the rules stated above for specific agglutinins, *i.e.*, marked agglutination with homologous bacteria; very weak, with heterologous. The typhoid serum on the other hand, although it fulfills the same requirements in the main, nevertheless it manifests some important differences when mixed with heterologous bacteria. It has practically no influence upon the cholera vibrio with which the typhoid bacillus is not at all related; agglutination of 1 : 10 can be attained even by a normal serum. The colon bacillus which closely resembles the typhoid, morphologically, but which has very different biochemical properties, is more strongly agglutinated, 1 : 25; while the paratyphoid bacillus, very much like the typhoid bacillus both morphologically and biologically, is agglutinated even in larger dilutions, 1 : 100. This entire phenomenon is an expression of the biological relationship of the various bacterial groups and is known as *group reactions*.

*An understanding of group reactions is to be found in a more complete conception of specificity.* From this source we have learned that the difference in antibodies is influenced by the dissimilarity of the injected antigen. For example, the difference between the cholera and typhoid agglutination is caused by the difference existing in the protoplasmic structure of the respective bacteria. As these bacteria, however, are not constituted of a distinct chemically defined substance, but made up of a mixture of various substances, there may be a number among them which can act as antigens. If, figuratively speaking, there are five different elements in the body of the typhoid bacillus which can act as agglutinogens, *i.e.*, antigens, these should be able to form according to the law of specificity five different agglutinins. On mixing a typhoid serum with typhoid bacilli, one brings together five distinct antigen antibody combinations and consequently complete and thorough action results of this union. A biological relationship of bacteria implies the exist-



ence of some common protoplasmic constituents. Expressed in the same figurative manner the colon bacillus can be said to have antigen number 1 in common with the typhoid and paratyphoid bacillus and the paratyphoid may have antigen numbers 1 and 2 in common with the typhoid bacillus. As a result, the typhoid serum will react with colon bacilli by virtue of their common agglutinin number 1, and with paratyphoid bacilli through its agglutinins numbers 1 and 2. The other "partial agglutinins" remain inactive on account of the missing suitable agglutinogens.

The existence of such partial antigens and partial antibodies is for some bacteria more than of mere theoretical importance. It is even possible that a strong colon serum will agglutinate no colon bacilli other than that particular strain employed for the production of the serum. Such being the case with a number of micro-organisms, the sera made at present both for diagnostic and therapeutic purposes are polyvalent (multipartial). By *polyvalent serum* is meant one which is produced either by immunizing animals with many different strains of the same bacterium, or a mixture of sera obtained from different animals immunized with various strains.

The practical importance of partial agglutinins is recognized Castellani's in the diagnosis of mixed infections. Castellani found that by Test. the mixture of an immune serum with its corresponding bacteria, the agglutinins for these as well as the partial agglutinins for the heterologous bacteria are absorbed. On the other hand, if the same serum be mixed with the heterologous bacteria, the agglutinins for the homologous group are quantitatively retained.

A practical example will make this clearer.

The serum of a patient agglutinates typhoid as well as paratyphoid bacilli, in a dilution of 1:100. This may indicate one of three possibilities:

- a. Patient is infected with typhoid, but has formed an exceptionally large number of partial-agglutinins for paratyphoid bacilli.
- b. Patient is infected with paratyphoid bacilli, but has formed at the same time many partial-agglutinins for typhoid.
- c. Patient has a mixed infection of typhoid and paratyphoid and therefore formed agglutinins for both.

A decision in regard to the above may be reached according to the following method given by Castellani:

Four rows of test-tubes are arranged, each row containing three tubes with 1 c.cm. of serum dilutions 1:10, 1:50, 1:100 respectively. In each of the first and second rows, 1 loopful of typhoid bacteria is emulsified.

In each of the third and fourth rows, 1 loopful of paratyphoid B. bacilli is emulsified.

The tubes are placed in the incubator for two hours, absence or presence of agglutination in each test-tube noted, and after centrifugalization (which may become unnecessary if the bacteria are strongly clumped or grouped at the bottom of the tube), the supernatant liquid is transferred to other test-tubes and kept in the same order.

Then each of the first row receives 1 loopful of typhoid bacilli,  
each of the second row receives 1 loopful of paratyphoid B. bacilli,



each of the third row receives 1 loopful of typhoid bacilli,  
each of the fourth row receives 1 loopful of paratyphoid B. bacilli,

All are once more placed in the incubator for two hours.

a. If typhoid exists the agglutination titer in the second part of the test will become weaker for the typhoid bacilli in the first row, and weaker for the paratyphoid B. bacilli in the second and fourth rows. The titer in the third row remains the same.

b. If paratyphoid exists, the agglutination titer for typhoid in the first and third row becomes less, that for paratyphoid in the fourth row diminishes, while the titer in the second row for paratyphoid remains the same.

c. If a mixed infection exists, the agglutination titer in the first and fourth row diminishes and in the second and third row remains the same.

In this connection a few exceptions may be mentioned:

A serum which is kept for a long time frequently loses part Agglutinoids. or even all of its agglutinating titer. Whereas it formerly agglutinated in the strength of 1:1000, it may now become inactive in dilutions even of 1:10. The first thought that arises in explanation of this is that the serum has perhaps degenerated and the agglutinins were destroyed. If, however, further dilutions are made, 1:100 may show mild, while 1:500 strong agglutination. This, first of all, demonstrates that agglutinins are still present, although diminished in amount, and second, that another substance has arisen which in the stronger concentrations interferes with agglutination. A simple experiment explains this.

If the test-tube containing the serum dilution 1:10 and the non-agglutinated bacteria be centrifugalized, the serum removed and the bacteria mixed with a known strongly agglutinating serum, it will be found that the bacteria have become inagglutinable. Substances of certain kinds have combined with the bacteria and prevented them from undergoing agglutination. These substances are strongly specific, acting only upon homologous bacteria. Their origin can also be demonstrated.

An agglutinating serum which is heated to 65° or 70° C. loses its agglutinating power but the substance interfering with the subsequent agglutination has remained. Ehrlich explains the situation as follows: He claims that agglutinins are built complexly; that they possess a binding (haptophore) group by means of which they unite with the bacteria (agglutigen) and a second group (ergophore or agglutino-phore) by virtue of which agglutination results. If serum is kept for a long period of time, or exposed to high temperature, many of the ergophore groups are rendered inactive, while the haptophore groups being more resistant remain and unite with bacteria. Agglutinins possessing only their haptophore groups are known as agglutinoids. They combine with the bacteria, and still do not agglutinate them, but at the same time prevent other agglutinins from acting. If this old agglutinoid and agglutinin containing serum is diluted, so few of both of these substances remain that the bacteria can absorb both, allowing the relatively few agglutinins to manifest their activity.

It is important to note in this respect that occasionally even a fresh, highly valent serum will present a tendency toward interfering with the agglutination processes. This is also explained by the existence of agglutinoids—a fact as yet not definitely proven.

Another finding, only encountered in exceptional cases, is the existence of the so-called *non-agglutinable strains* of bacteria. These give all the



characteristics of the general class of bacteria to which they belong, but are not agglutinated by their respective serum; as, for example, a strain of typhoid bacilli, which are not agglutinated by any typhoid serum. The only positive proof that they are typhoid bacilli is the ability to produce by their injection into animals an active immunity against fully virulent typhoid bacteria.

Non-agglutinable strains of bacteria can be isolated especially from the lower animals. At times, however, they regain their agglutination property when they are grown in artificial media and frequently subplanted. Possibly, the reason that the bacteria become inagglutinable at all, is that they undergo immunization within the organism against the existing agglutinins. By growing bacteria in agglutinating serum for a certain time, one can obtain non-agglutinable strains.

1. Agglutinins for *typhoid* and *paratyphoid* A and B, can, not infrequently, be demonstrated in the patient's serum as early as the third day, but as a rule, at about the beginning of the second week of the disease. Moreover, they remain within the serum for several weeks after the illness and disappear only gradually. A positive agglutination test does not, however, mean the existence of the corresponding disease. *A healthy bacillus carrier can also have an agglutinating serum.* Some cases of icterus catarrhalis even give a positive Widal test. But in order to assign to this last a correct explanation, one must remember that typhoid bacilli may remain in the gall-bladder for years and thus lead to catarrhal inflammation and stone formation.

Partial agglutinins from coli infections must always be considered. Some authorities mention a positive Widal, in connection with endocarditis maligna, sepsis, malaria, phthisis, and miliary tuberculosis.

An absence of the agglutination test, especially at the early part of an illness, should not influence a negative diagnosis of typhoid too greatly, inasmuch as many cases are known where the reaction appeared for the first time during the period of convalescence. In the employment of this test as an aid for the differential diagnosis between several bacterial infections, it is best to titrate the serum to its limit, as the higher titer for one class of bacteria generally speaks in favor of the infection by the same. Paratyphoid serum agglutinates typhoid bacilli only slightly, while true typhoid agglutinates both typhoid and paratyphoid bacteria with almost equal strength. In severe and difficult cases, Castellani's test should be performed. Paratyphoid B. serum always gives the limit of its agglutinating titer both with the pathogenic mouse typhoid and hog cholera bacillus.

2. *Cholera*.—Only rarely has the agglutination test been employed with the serum of patients infected with cholera. On the other hand, if cultures from a cholera stool are made upon a plate, the identification of the suspicious colonies grown here is regularly conducted by means of this test. For



this purpose, it is very specific, as group reactions almost never take place. Strong agglutinating sera are easily obtained by immunization of animals.

3. *Epidemic, Cerebrospinal Meningitis*.—Agglutination in this disease serves mainly for the identification of suspicious meningococcus cultures. As has been shown by Wassermann and Kutscher, some strains are agglutinated only after a long period (twenty-four hours) and at higher temperatures as 56° C.<sup>1</sup>

4. *Dysentery*.—The agglutination property is employed both for testing the serum, and identifying cultures. The Flexner type of bacillus produces agglutinins more readily than the Shiga-Kruse. It is also agglutinated more readily. Only positive reactions in dilutions of 1:30 are of diagnostic consideration. Occasionally, partial agglutination takes place with heterologous dysentery strains, typhoid and colon bacteria.

5. *Pest*.—The reaction is very specific, but of slight significance, as it appears only on the ninth day; occurring with a serum dilution of 1:3, it is considered of positive diagnostic value.

6. *Malta Fever*.—In most instances the serum gives the agglutination reaction with the micrococcus melitensis. Normal serum may give the reaction in dilution 1:30, so that higher dilutions only are of aid in diagnosis.

7. *Staphylo-, Strepto- and Pneumococci*.—Clinically, the agglutination test is never employed in these cases.

8. *Tuberculosis*.—Here the agglutination test is associated with the difficulty of obtaining a homogeneous tubercle bacillus suspension. This, however, is overcome in one of two ways.

a. *Arloing-Courmont's Method* (1898).—The tubercle bacilli are obtained in the so-called "homogeneous culture" form. S. Arloing first grows the bacteria on potatoes for a long time, and then transplants them in glycerin bouillon which is shaken daily for five minutes. After a number of subcultivations, a culture is obtained after several months. This strain grows rapidly in a few days and diffusely clouds the broth.

Such a culture diluted with physiological saline solution is used for the test. Here small test-tubes are preferable and the ingredients should be mixed in the following proportions:

- 2 drops of serum + 10 drops of culture (1:5)
- 1 drop of serum + 10 drops of culture (1:10)
- 1 drop of serum + 15 drops of culture (1:15), etc.

The tubes are well shaken and placed in the incubator. According to Arloing and Courmont, a positive reaction even in the dilution of 1:5 speaks for tuberculosis. Best results are by this means obtained in incipient and mild tubercular cases; those which are farther advanced do not react.

b. *Method of Koch*.—Koch filters the ordinary tubercle bacillus bouillon cultures, dries the remnants upon the filter, and rubs them up in an agate mortar with N/50 NaOH to a dilution of 1:100. The solution is centrifugalized and enough weak HCl is added until the reaction is only slightly alkaline. The dilution is then brought

<sup>1</sup> Frequently, during even the first days of the disease, the patient's serum in a dilution of 1:10 gives the agglutination test. This is rare with higher dilutions of the serum as 1:50. It usually takes some time before the agglutination becomes evident.



up to 1:3000 by the addition of 0.5 per cent. phenol in normal saline, and kept for twenty-four hours in the incubator.

A somewhat simpler procedure is to dilute new tuberculin B. E. to 1:100 with 0.5 per cent. of carbolic saline solution, centrifugalize this for six minutes and then dilute to 1:1000. The solution thus obtained can be preserved in the ice-box for fourteen days. Just before using, a still further dilution of 1:10 is made.

The agglutination test has not been generally adopted as a method of diagnosis. The technique is rather difficult, and the results not absolutely reliable. The reason for this is that high agglutination values are rarely met with, and slight ones are found even in normal individuals. Then, too, the methods of tuberculin diagnosis are so much simpler that they have been given the preference.

Koch himself advised the agglutination test, not as a means of diagnosis, but rather as an aid in tuberculin therapy. He found that during the treatment of tuberculosis with new tuberculin the agglutinative power of the patient's serum increased. He therefore took this as an index of the acquired immunity. Further study, however, convinced him that the agglutination cannot thus be interpreted, so that at the present day tuberculosis agglutination has no practical application.

10. *Glanders*.—Highly valent sera can be obtained, according to Kleine, by intravenous immunization of donkeys and goats. The serum serves for identification of the glanders bacilli. Kleine prepares a standard bacterial emulsion in the following manner: Four well grown glanders cultures are killed at 60° C. and the mass of bacteria triturated in 2 c.cm. of 1/2 per cent. carbolic-saline solution. This is then diluted in a measuring glass so that 40 to 50 c.cm. of carbolic-saline solution are added for each culture. The entire mixture is filtered through paper and 3 c.cm. are used in each test. Normally, horses may have an agglutination titer up to 1:400. Glanders infected animals react as high as 1:2000. Injections of mallein increase the agglutination titer. Experiences in this respect with the human being are still scanty.

Hemagglu- tinins, Immune and Normal.	Just as injections of bacteria produce bacterial agglutinins, injections of erythrocytes stimulate the formation of hemagglutinins which cause the red blood cells to congregate in clumps.
---	---

At times the presence of hemagglutinins is masked by the simultaneous existence of hemolysins which dissolve the red blood corpuscles. If, however, the immune serum is heated to 56° C. the complement is destroyed, thus interfering with the action of the hemolysin and allowing the agglutinins to exhibit their action. In other instances as during the immunization of rabbits with dog's erythrocytes, hemagglutinins are formed in such great quantities that by mixing the immune rabbit's serum with the dog's erythrocytes so strong an agglutination occurs that the hemolysins can no longer attack the clumped erythrocytes. The hemolysis can be demonstrated only if clumping is prevented mechanically, by thorough shaking of the mixture.

Also under normal circumstances, *i. e.*, without immunization, the serum of one animal species can to a certain degree agglutinate (and hemolyze) the red blood cells of another species. The quantity of these normal hemagglutinins is comparatively small.



Still more interesting is the observation that there are hem-Isohemag- agglutinins against the red cells of different animals even of  
glutinins. the same species, so-called Isohemagglutinins or Isoagglutinins.

These have thus far been demonstrated in the bloods of dogs (Von Dungern) steers and rabbits (Ottenberg and Friedman). Isoagglutinins in the human serum were discovered independently by Landsteiner and Shattock in 1900. At first the occurrence of isoagglutination was regarded of pathological significance, but soon it was shown that the phenomenon occurred with a large percentage of normal bloods. In fact all human bloods can be divided into four sharply defined groups according to the way in which they interagglutinate. The groupings can be explained by assuming the existence of two agglutinins of which the first group possessed both, the second one, the third one, and the fourth neither. In each case the cells are susceptible only to that agglutinin which does not exist in the individual's own serum. Thus:

The serum of the first group, designated as group I, possesses the power of agglutinating the red cells of members of all the other groups but the red cells of members of group I are not agglutinated by any human serum. This group includes about 50 per cent. of all persons examined.

The serum of the members of the second group agglutinates the red cells of members of the third and fourth groups. The cells of members of the second group are agglutinated by sera of individuals of groups I and III. The serum of group III agglutinates cells of persons belonging to members of the second and fourth groups; its cells are agglutinated by sera of the first and second groups.

The fourth group, whose members are relatively rare, is characterized by possessing no agglutinin for human red cells and by its cells being agglutinable by the sera of all other groups.

The group characteristics are permanent for each individual throughout his life. When concentrated, the agglutinins act almost instantaneously; when diluted they act more slowly. Agglutination occurs in the cold as well as at high temperatures. The peculiar groupings are not only permanent with the individual but they are hereditary. Von Dungern and Hirschfeld have conclusively proved that agglutinins are hereditary and follow the Mendelian law. This observation had, however, been made long before this in a paper by Epstein and Ottenberg (1908).

With the recently increasing popularity of blood transfusions, the phenomena of isoagglutination and hemolysis, the two being very closely related, have attained a more practical significance. In selecting donors for a transfusion, agglutination and hemolysis tests should always, when time permits, be made before operation. These tests *in vitro* are usually a safe guide as to conditions *in vivo*. That donor should be chosen who

belongs to the same group as the patient, that is where no interagglutination or hemolysis exists. This is advisable not only so as to get the best results from the transfusion, but also in order to avoid any untoward symptoms or intoxications that are associated with the intravascular agglutination or hemolysis (rise of temperature, dyspnea, edema, hemoglobinuria). The more important of these two factors is not as yet clear. If for a given transfusion a donor belonging to the same class cannot be obtained, it is safer to use a person whose serum is agglutinative toward the patient's cells than one whose cells are agglutinated by the patient's serum.

The materials necessary for the agglutination and hemolysis

Transfusion tests are as follows:

Tests. (1) Sterile syringes or needles for puncturing the vein; (2) 1 per cent. sodium citrate solution in 0.85 per cent. salt solution; (3) 0.85 per cent. salt solution; (4) a test-tube rack having two narrow test-tubes ( $4 \times 1\frac{1}{2}$  in.) for each donor (numbered); one partly filled with the sodium citrate solution, one empty.

From each donor 1 to 2 c.cm. of blood are aspirated from a vein of the elbow; several drops of this blood are allowed to flow into his tube with the sodium citrate solution, the rest is collected in his dry test-tube. This is also done to the recipient, but from him 3 to 4 c.cm. of blood are necessary so as to have sufficient serum for a number of donors (10-15).

The sodium citrate tubes are centrifugalized and a sediment of the red blood cells obtained; the supernatant fluid is pipetted off and the red cells made up approximately to a 10 per cent. suspension with the normal salt solution.

The serum tube is also centrifugalized so that clear serum is separated off. The clot should not be disturbed too energetically as it is best to get absolutely clear yellowish serum not blood tinged.

The following mixtures are then made with each donor's blood, preferably within 12-24 hours of the time of collecting the blood:

(a) 3 parts or units of donor's serum and 1 part or unit of recipient's red cell emulsion.

(b) 3 parts or units of recipient's serum and 1 part or unit of donor's red cell emulsion.

*Controls:*

(c) 3 parts or units of donor's serum and 1 part or unit of donor's red cell emulsion.

(d) 3 parts or units of recipient's serum, 1 part or unit of recipient's cell emulsion.

(e) 3 parts of saline, 1 part of donor's red cells.

(f) 3 parts of saline, 1 part of recipient's red cells.

These mixtures are made in very small test-tubes ( $3 \times \frac{3}{8}$  inch). The



quantity comprised by each "part" or "unit" varies according to the amount of blood or serum obtained from each donor and recipient. If sufficient, it is best to work in drops (each drop being about 0.05 c.cm.), thus mixing 3 drops of serum (0.15 c.cm.) and 1 drop of the red-cell emulsion (0.05 c.cm.). If the serum is not sufficient, a smaller arbitrary unit may be used in the form of Wright's pipettes (4 to 5 millimeters caliber) fitted with rubber nipples, drawn out to a length of 2 to 3 inches, and marked off about 1 inch from the tip with a blue pencil, this distance comprising the unit.

The tubes are placed in the incubator for three hours and then in the ice box for 24 hours. Agglutination when it occurs does so rather promptly, within 15 to 30 minutes after the mixtures have been made. It is recognized macroscopically by the clumping of the red blood cells into small floccules which later on appears like a distinct clot. A hanging drop preparation of such a mixture shows the same phenomenon; usually this is unnecessary as the macroscopical appearance is characteristic. If one is in doubt, however, microscopical examination should be made. Hemolysis, if pronounced, is observed in an hour or even less, but certainly after the three hours' incubation; the finer grades of hemolysis are detected after the tubes have remained in the ice box 12 to 24 hours. The control tubes must show no agglutination or hemolysis.

In cases where for any reason the taking of blood from the vein is not allowed or impossible (as in infants), Wright's method of working with small quantities (Epstein and Ottenberg) should be resorted to.

The skin at the bed of the finger nail or of the lobe of the ear is pricked deeply with a Hagedorn needle. For the red blood-cell suspension several drops of blood are allowed to flow or taken up with a dropper and expelled into a tube with sodium citrate solution. The red blood cells are washed and diluted as before.

For the serum, three or four Wright's capsules (see under opsonins) are filled with blood which is allowed to clot, and centrifugalized. Each capsule is nicked with a file, allowing the capsule to be broken open and the serum pipetted off.

For making the mixtures Wright's pipettes (four to five millimeters caliber) fitted with rubber nipples are used. With a blue pencil an arbitrary point is marked off upon the drawn out extremity of the pipette. Three volumes of serum (a bubble of air between each volume), and one volume of cell suspension are drawn into the same pipette. These ingredients are mixed by running them gently out and then drawing up again into the pipette. The entire mixture is then drawn into the body of the pipette and the tip is sealed in a flame. Each pipette serves as a little tube.

The time and manner of incubation, and the observation for hemolysis follow the rules mentioned above.

This method is especially valuable when a very great number of donors are to be examined. If, however, a sufficient amount of serum is obtainable the editor prefers the first method as it is a good deal simpler; then, too, the total amount in each test is the same, thus allowing better of comparison as to the intensity of hemolysis.



## CHAPTER XI.

### PRECIPITINS.

In the former chapter, the phenomenon of agglutination was explained as a clumping of bacteria occurring when serum is mixed with its corresponding bacteria. In 1897 R. Kraus described a phenomenon, very closely allied to the one just mentioned. He found that when an immune serum, for example, of cholera, typhoid, or pest, is mixed with the clear, sterile filtrate of the respective bouillon cultures of their bacteria (instead of the bacteria themselves), the clear solution becomes turbid, and a precipitate forms. This reaction is known as precipitation; the elements within the immune serum, precipitins; while the substances (antigen) with which the precipitin reacts and which originally stimulated the production of the precipitin, *precipitinogen*.

Like all biological reactions, the phenomenon of precipitation is not limited to bacterial immune sera and culture filtrates, but is observed when any animal, vegetable or bacterial soluble proteid substance is mixed with the serum of an animal which has been immunized against the particular proteid material in question.

Tschistowitsch and Bordet were the first who called attention to these non-bacterial precipitins. Bordet (1899) found that the blood serum of rabbits treated with the serum of chickens gave a specific precipitate when mixed with chicken serum. Tschistowitsch demonstrated a similar reaction with the sera of rabbits treated with horse's and eel serum.

The biological structure of the precipitins is strongly analogous to that of agglutinins. Many authorities, in fact, consider them identical. Whatever has been said in regard to the effects of heating and addition of acids or alkalies upon agglutinins, applies equally to precipitins. Moreover, they also are composed of two groups, a binding (haptophore) and a functionally active (ergophore) group. If the latter is missing, they are known as precipitinoids, and can interfere with precipitation just as agglutinoids do with agglutination.

In speaking of precipitation, it has always been customary to differentiate between bacterial and proteid. For practical purposes this division is superfluous inasmuch as the bacterial precipitins are nothing more than precipitins of bacterial proteids.

### Bacterial Precipitin Reactions.

For the production of precipitating sera, animals, preferably rabbits, are injected either with broth cultures or salt solution emulsions of agar cultures of the bacteria. Five to six injections of gradually increasing quantities are given intraperitoneally or intravenously, at intervals of from five to six days. The dose varies with the virulence of the bacteria.

The intravenous method frequently gives the stronger serum but the mode of administration depends also on the pathogenic properties of the microorganism in question. The immunized animals should be bled about seven to twelve days after the last injection of bacteria. The filtrates of bouillon cultures and the various forms of bacterial extracts will also, when injected, produce precipitins. The serum from individuals undergoing an infection, or convalescing from one, contains precipitating bodies against the respective infective agent.

Inasmuch as the precipitin reaction consists in the formation of a precipitate, it is important that *both of the ingredients (precipitin and precipitinogen) be absolutely clear and have no tendency to spontaneously become turbid, or form a precipitate.*

In order to get a clear serum one should avoid withdrawing the blood during the period of digestion of the animal, because it is chylous at such a time. In man the best occasion for obtaining the blood is in the morning before breakfast. As for animals, it is advisable to give them no solid food (or milk) for twenty-four hours previous to venesection. Then a very minute quantity of blood is withdrawn and immediately centrifugalized in order to ascertain whether the serum is clear or not. If it is satisfactory, larger amounts may be collected. The presence of erythrocytes and bacteria causes a serum to be turbid. Simple sedimentation or centrifugalization suffices to overcome this.

If in spite of these precautions turbidity still persists, recourse may be had to filtration through paper or bacterial filters, preferably new ones. This method should, however, be used as a last resort, because filtration always tends to diminish the strength of a serum.

Bacterial precipitinogens are prepared by filtration either of bouillon cultures or bacterial extracts. The filtrates must be absolutely clear; also sterile, as frequently the precipitin reaction requires a long period of time. If bacteria are present they may grow quickly, and produce turbidity. After a time the precipitinogen loses its property of combining with precipitins and forming precipitates. In such a case the precipitinogen can still be employed for immunization purposes.



A constant amount of precipitinogen is placed in each of a row of test-tubes, and to these are added diminishing amounts of the immune serum.

Technique of the Precipitin Reaction. A set quantity of serum and varying amounts of precipitinogen can also be employed. The result of the reaction depends to a very large extent upon the quantitative relationship of these ingredients. *If relatively too much precipitinogen exists, a precipitate will not form.* An already formed precipitate will dissolve on the addition of more precipitinogen.

The explanation of this peculiarity is unknown. Since colloidal substances, however, at times give similar reactions, many authorities have classed the precipitins among them.

The technique of a precipitation test is best seen in the following table:

Cholera bouillon filtrate	Cholera serum	Physiological saline sol.	Result	
			After 4 hours	After 24 hours
5.0 c.cm.	1.0 c.cm.	.....	Very cloudy.	Clear; marked sediment at bottom.
5.0 c.cm.	0.5 c.cm.	0.5 c.cm.	Cloudy.	Clear with moderate sediment at bottom.
5.0 c.cm.	0.1 c.cm.	0.9 c.cm.	Faint cloud.	Clear; slight sediment.
5.0 c.cm.	0.05 c.cm.	0.95 c.cm.	Clear.	Clear; no sediment.
5.0 c.cm.	.....	1.00 c.cm.	Clear.	Clear; no sediment.
.....	1.0 c.cm.	5.0 c.cm.	Clear.	Clear; no sediment.
.....	0.5 c.cm.	0.5 c.cm.	Clear.	Clear; no sediment.
.....	0.1 c.cm.	5.9 c.cm.	Clear.	Clear; no sediment.
.....	0.05 c.cm.	5.95 c.cm.	Clear.	Clear; no sediment.

A parallel row of tubes with normal serum should be included.

If highly valent sera, such as are obtained by immunization with bacterial extracts, are employed, precipitation may result soon after mixing the two constituents. The precipitins are strongly specific, although it may be said that just as in agglutination, *there exists in precipitation a certain degree of "group reactions."*

The precipitation test has no clinical diagnostic value. It demonstrates nothing more than the agglutination test, is more difficult of execution and associated with greater sources of error. Only occasionally is it of service to prove the presence of soluble bacterial substances within exudates or organ fluids.

Diagnostic Value of Bacterial Precipitation.

Porges and v. Eisler have employed the precipitation test as a means for the differentiation of capsule-bacteria where the method of agglutination is associated with certain difficulties. The precipitinogen was produced by filtration of four-weeks-old

bouillon cultures of pneumococci, rhinoscleroma, and ozoena bacilli. The immune serum was obtained from rabbits which had received four to five subcutaneous injections of the respective bacterial suspensions.

Fornet has recently advocated the precipitation test as an aid in the clinical diagnosis of typhoid fever. Although his attempts have not been attended with practical success, the principles of the reaction deserve discussion on account of their originality.

Fornet believed that it should be possible to demonstrate in the blood of typhoid patients the presence of the antigens (precipitinogens) which stimulate the antibodies, long before the latter themselves become evident. He actually was able to obtain turbid mixtures when he combined precipitating typhoid serum with the serum of typhoid patients. In many cases he obtained these results before the appearance of the Gruber-Widal reaction.

The method which he has recently employed is known as the "ring test."

Small test-tubes 8 cm. in height and 0.5 cm. wide, are placed in rows of twenty each in a small black test-tube rack so arranged by the help of Ring Test. side stands that the tubes are inclined at an angle of about  $45^{\circ}$ . Across the back of the rack is attached a strip of dark cloth as a background to facilitate the detection of any precipitate. The immune (or convalescent) serum is placed in tubes in concentrated and diluted form 1:5 and 1:10 with normal saline, and then the serum for examination in concentrated and similar dilutions is carefully floated on top of the immune serum. The mixtures are allowed to stand undisturbed at room temperature for two hours, and if the reaction is positive a whitish ring at the point of contact of the two sera makes its appearance. A control test-tube of normal serum plus immune, and another of normal plus the unknown serum in the same dilutions as those employed in the test, must remain negative.

Besides in typhoid fever, the ring test is also evident in scarlet fever, measles and syphilis.

For the precipitation test in syphilis, the serum from patients with manifest luetic symptoms is employed as precipitinogen, Syphilis and the serum from individuals with general paresis acts as precipitating agent. The ring test must be carried out strictly in accordance with the rules given by Fornet, but even so, its diagnostic value for syphilis is still doubtful. Plaut claims that normal serum gives the reaction just as often as luetic serum; this is strongly denied by Fornet.

Theoretically, it is questionable whether these precipitates and rings are similar in origin to bacterial precipitates, or whether physical-chemical causes are at the bottom of the former phenomenon. In accordance with the latter view several other reactions have been recently recommended for the serum diagnosis of syphilis.

*a.* Porges and Meier noticed that luetic sera are capable of producing flocculent precipitates from lecithin solutions. Porges soon found the same occurrence with solutions of bile salts.

Many additions and modifications have been made to the Porges' Reaction.



reaction since it was first recommended. According to the most recent publication, the reaction is carried out as follows:

The requirements are:

1. One per cent. solution of sodium glycocholate (Merck) in distilled water.
2. The patient's serum which must be absolutely clear, and heated for one-half an hour at 56° C.

Two-tenths of each of the above are placed into a narrow test-tube 6 to 7 mm. in diameter, and allowed to rest for sixteen to twenty hours at room temperature. A positive reaction consists of the appearance of distinct coarse flocculi, which as a rule, collect near the surface. Mere turbidity or faint precipitates are considered as negative.

The original Porges method of employing lecithin was not at all specific, the reaction being present in tuberculosis, carcinoma, and other infectious diseases. As for the new modifications, nothing has been brought forward in their support.

This reaction belongs to the same general class of precipitation tests for lues, but is very much simpler than any of the others. Two-tenths c.cm. of absolutely clear, fresh (at the most, two hours old), active serum is mixed with 0.6 c.cm. of distilled water, in a small test-tube 7×0.5 cm. Sera containing hemoglobin or lipoids are not suitable for this reaction. The mixtures are allowed to stand at room temperature. In several hours, at the latest fifteen, a thick flocculent precipitate 2 to 4 mm. high appears at the bottom of the tube. Kreibich's analysis showed it to consist of fibrin globulin.

Apparently this substance is increased in luetic serum and precipitated by the distilled water in which it is insoluble. Klausner's reaction is by no means specific for syphilis as it is in evidence in starvation, typhoid fever, measles, scarlet fever, pneumonia, and other diseases, as well as during health. Nevertheless it must be said that it is found more frequently, earlier and much stronger in lues than in any other condition.

Klausner states that in fresh cases of lues the best reaction is seen in about seven to nine hours, while in older cases a weak reaction appears in twelve hours. Mercury influences the test in that the interval until the precipitate becomes marked, is prolonged and later on the precipitate becomes fainter.

In spite of its simplicity, Klausner's reaction has not been generally adopted for clinical work, as the Wassermann reaction with its far greater accuracy has replaced it.

### Proteid Precipitins.

While bacterial precipitation is interesting from a biological standpoint but bears no practical significance, proteid precipitation represents one of the most important practical aids in forensic medicine. *By this means the differentiation of various proteids can be easily and definitely determined, a problem which was left unsolved by chemistry.*

The phenomenon of protein precipitation is absolutely analogous to



that of bacterial precipitation. If a clear proteid solution ( $a$ ) is mixed with the clear serum ( $a'$ ) of an animal immunized against the above proteid ( $a$ ), turbidity and precipitation will occur; while if a mixture of the serum ( $a'$ ) is made with a non-homologous proteid say ( $b$ ), or a mixture of the proteid ( $a$ ) with the serum ( $b'$ ) of an animal immunized against  $b$ , no precipitation takes place. Graphically expressed it looks thus:—

$a + a' =$ precipitation.

$b + a' =$ no precipitation.

$a + b' =$ no precipitation.

$b + b' =$ precipitation.

In other words, *a precipitating immune serum reacts only with its homologous proteid. The precipitin reaction is specific.*

Forensic Use of Albumin Differentia-  
tion. It is greatly to the credit of Wassermann and his co-workers A. Schütze and Uhlenhuth, who recognized that this specificity of precipitins was of great medico-legal value.

For example, a bloody shirt is found in the home of a man charged with murder; the prosecution sees in that the proof of crime, while the defendant pleads that the stains belong to the blood of a sheep; the proof as to their source is of the utmost deciding evidence; and while chemical or microscopical examinations are of little or no use, serum diagnosis wins the day.

The blood-stained clothing is extracted in water, part of the extract is mixed with  $a$ , the serum of a rabbit immunized against human serum and another part is mixed with  $b$ , the serum of a rabbit immunized against sheep's serum. If the mixture  $a$  shows a precipitate, it can be definitely stated that the blood stain contained serum derived from a human being; while if mixture  $a$  is clear and  $b$  shows the precipitate, it is strongly corroborative of the presence of sheep's serum.

Blood Relationship. This example suffices to indicate the value of this biological fact. In addition the reaction is made use of in the determination of the nature of meats (detection of horse meat substitution for beef).

Furthermore, this method has explained a number of scientifically interesting problems. Just as group agglutination demonstrated the close relationship existing between various bacteria, so also serum precipitation proves a distinct relationship between the different species of animals (horse and donkey, dog and fox, hare and rabbit, ape and man, etc.).

Thus the serum of a rabbit immunized against human serum precipitates not only human serum but also that of monkeys; the serum of a chicken immunized against rabbit's serum precipitates not only that, but also hare's serum. In order, however, to differentiate between rabbit's and hare's serum, Uhlenhuth advises the immunization of a rabbit with hare's serum. The serum of such an immunized rabbit precipitates



only hare's serum and not rabbit's, for the reason that "Isoprecipitins," *i.e.*, precipitins against the same kind of animal, are, as a general rule, not developed. Similarly the differentiation between human and ape's serum can be accomplished by the immunization of apes with human serum.

Attempts have also been made by means of the precipitation reaction to determine the origin of albumin in urine, and the foreign proteids circulating in the blood of artificially fed infants.

The technique remains the same, independent of the purpose it is employed for. It consists in the mixing of the clear precipitating serum and the clear proteid, or albumin precipitinogen.

Strongly precipitating antisera against proteid solutions are prepared by methods analogous to those employed for the production of Proteid production of antibacterial sera. The use of rabbits is generally advised. The sera or proteid solutions should be Antisera. sterile. Filtration through small porcelain filters may be necessary. The injections may be made subcutaneously, intraperitoneally, or intravenously. The subcutaneous route has no advantage unless the substances to be used are contaminated; but then, larger quantities are necessary. The intravenous path is the one of choice. A single injection of a large dose (15 to 20 c.cm.), or three injections of moderately large doses (5 to 10 c.cm.) on three successive days may be given and the animal killed after 7 to 10 days. These methods have the advantage that a precipitating serum is obtained in a short period of time. They do not however yield as strong a precipitating serum as the following slower procedure. One c.cm. of the solution is injected four or five times at intervals of six days. The dosage may also be increased at each successive inoculation, for instance, beginning with 2 c.cm. of an animal serum and increasing gradually through 3, 5 and 8 c.cm. to possibly 15 c.cm. at the last injection. The animals should be weighed from time to time and if considerable loss of weight ensues during immunization, the intervals between injections should be increased.

It is advisable to inject five or six animals at the same time, and by different methods, inasmuch as rabbits vary greatly in their individual power to produce precipitins and moreover, because some die after the third injection. Frequently only one serviceable serum is obtained, even though the immunization of five rabbits was undertaken.

Beginning on the sixth day after the last injection, one should, at regular intervals of one or two days, remove a small quantity of blood from the vein of an ear and test the strength of the serum. As soon as it is found to be satisfactory the animal should be bled and its serum preserved on ice, with precautions for sterility. The rules given above for obtaining



a clear serum should be kept in mind. If the precipitating value of the serum is insufficient, more injections may be given before the animal is finally bled.

If the serum is not withdrawn at the proper time, its strength begins to diminish and further injections no longer stimulate new antibodies. It is even possible for the entire precipitin action of the serum to disappear.

**Titration.** The following method of titration of the strength of a serum is the simplest. One c.cm. of various dilutions (1:10, 1:100, 1:1000, 1:10000) of the proteid under examination (precipitinogen) is placed into different test-tubes and 0.1 c.cm. of the precipitating serum is added to each. The tubes should not be shaken, but it is occasionally necessary to place them in the incubator for one hour before any turbidity or precipitate appears. The least amount of proteid solution which still distinctly shows a precipitate is taken as the titer of the serum.

For medico-legal purposes, Uhlenhuth advises the use of only Uhlenhuth's highly valent sera.

**Method of Proteid Differentiation.** He considers an antiserum as efficient if 0.1 c.cm. of it, when mixed with its respective serum in the dilution of 1:1000, produces a distinct turbidity, either at once or in one to two minutes at the latest; three to five minutes is the limit for an indication of turbidity in the dilutions of 1:10000 and 1:20000.

Like in all other biological reactions, control tests, here two in number, are of the utmost importance. One tube must contain 0.1 c.cm. of the precipitating serum mixed with 1 c.cm. of saline, another 0.1 c.cm. of the precipitating serum mixed with a heterologous serum in the dilution of 1:200 and 1:1000. Both of these tubes should show absolutely no precipitate after twenty minutes. In this way the specificity of the precipitin is determined; and it must be remembered that it is the quantitative specificity which counts.

In the process of the determination of the nature of meats, it is especially necessary to ascertain exactly the precipitating titer against bovine and pig's serum possessed by the rabbit's precipitating serum directed against horse's serum.

When clear solutions are at hand the precipitin reaction is comparatively simple. Frequently, however, the test must be performed with old and dirty blood stains, or all kinds of prepared sausage, so that the first and important task is to obtain a clear solution.

In dealing with blood, milk, or seminal stains, the parts of the clothing involved are excised, divided into very minute shreds, and placed in a test-tube with a small amount of 0.85 per cent. of salt solution. If the material is not too old, extraction for one hour is usually sufficient, otherwise it may necessitate a period of twenty-four hours or more. Stains upon solid material such as steel, wood, stone, etc.,



are carefully scraped off, and suspended in physiological salt solution. To obtain a clear solution the extract must be passed through filter paper or eventually the lilliputian bacterial filter.

In the examination of meats or other food stuffs, it is best to remove the material for examination from the center of its thickest part, as this portion has been least exposed to the methods of preservation, especially the high temperatures. Three hours' extraction is usually sufficient; the fresher the meat, the shorter this period. Very much salted meats are best washed with distilled water, previous to extraction. Inasmuch as a great deal of fat interferes with the reaction it is advisable to remove it beforehand by extraction with ether and chloroform for twenty-four hours (Miessner and Herbst).

Before performing the actual test with the unknown blood stain, it is best to try out the entire reaction with a similar but known blood stain in order to make sure whether all the ingredients are in good working order. In laboratories equipped for medico-legal examinations, stains made upon linens from the blood of man, ox, pig, horse, etc., are always kept in readiness for such preliminary tests.

Uhlenhuth indicates a set of rules to be observed whenever the reaction is undertaken. They are here cited in their original form, as practice has shown them to be of great service.

"In order to obtain sufficient extract for the test, a small amount of the material is placed in a test-tube containing 5 c.cm. of normal salt solution. This must not be shaken. After one to two hours, 2 c.cm. are poured off into another tube and gently shaken. If a persisting froth appears upon the surface of the fluid, it can be taken as proof that sufficient extraction has occurred, and the rest of the fluid is thereupon also transferred to this tube. If no froth appears the 2 c.cm. should be returned into the first test-tube and the extraction continued until repeated tests finally show the presence of froth. It is preferable not to disturb the sediment at the bottom of the test-tube. The extract eventually obtained may have to be filtered, if not absolutely clear.

Such an extract is, as a rule, stronger than that required for the test, *i.e.*, 1 : 1000. If one drop of a 25 per cent. nitric acid solution is added to 1 c.cm. of a 1 : 1000 serum dilution and then heated, a faint opalescence appears. Enough saline should therefore be added to the final extract so that the nitric acid test corresponds to that given by a dilution of 1 : 1000.

The following mixtures are then made:

Test solution	Precipitating serum from rabbit	Normal rabbit's serum	Normal saline	Result	
				After five minutes	After twenty minutes
1 c.cm. 1 : 1000	0.1	.....	.....	Opalescence.	Turbidity and sediment.
1 c.c.m 1 : 1000	.....	0.1	.....	Clear.	Clear.
.....	0.1	.....	1 c.cm.	Clear.	Clear.

The result should be read after twenty minutes at room temperature. As a further control a similar row of tubes should be made with the extract of the non-bloody part of the clothing in order to show that the latter alone does not give the reaction.

Even putrid or otherwise chemically changed proteids may still give the precipitin reaction.

The precipitation test only determines the animal species from which the proteid originates, but cannot prove whether it comes from the blood, semen, milk or other proteid body. In order therefore to make a medico-

legal diagnosis of "human blood stains," chemical evidences "Origin" must in addition be brought forward, that the stain really and "Con- consists of blood. Obermeyer and Pick have further shown stitutional" that besides animal specificity ("origin specificity"), pre-Specificity. cipitation also demonstrates the "constitutional specificity" of proteids.

If instead of employing pure animal or plant proteids for the immunization of animals, variously changed albumins are used (heated albumins, acid albumins, formaldehyde albumin, etc.) the organism reacts by producing antibodies of a characteristic nature, different from those developed after inoculation with the pure albumin. For example, the serum of a rabbit immunized for a long time with horse's serum (normal immune precipitin) will produce a precipitate when mixed in vitro with the pure horse's serum and not when added to the latter, heated, even if the normal immune serum is of very high titer. On the other hand, if a rabbit is injected with horse's serum which has been changed by being diluted and boiled for a short time, the immune serum thus obtained will react not only with normal horse's serum but also with heated serum and a group of its decomposition products with which the normal immune serum ordinarily never induces a precipitate.

This fact is of practical application. In meat substitution, it is very popular to boil the sausage in order to make detection of the substituted meats more difficult. With the aid, however, of precipitins produced by immunization with heated proteids, this fabrication is more easily detected than if a normal immune serum were used.

While animal specificity is not destroyed when the albumins are modified in the above manner or changed by tryptic digestion or oxidation, Obermeyer and Pick have demonstrated that their specificity is lost when an iodine, nitro or diazo group is inserted in the proteid molecule. Immunization with such transformed proteid compounds, *e.g.*, xanthoprotein, can produce a precipitating serum which will react with every xanthoprotein even in homologous animals. These authors conclude that species specificity is probably dependent upon a certain aromatic group of the proteid molecule.



It is interesting to note that the proteid contained in the lens of the eye belongs to this class of modified proteids which possess constitutional, but no species specificity. A serum produced by immunization with lens substance will react with the proteid derived from the lens of any animal, but with no other animal proteid.

In conclusion, the origin of the precipitate formed during the Origin of precipitation reaction is of interest. When a very strong Precipitate. precipitating serum is employed, the precipitinogen is so greatly diluted that it no longer gives any of the chemical reactions for proteids, but nevertheless yields a heavy precipitate when the precipitating serum is added. This surely cannot come from the small trace of proteid in the precipitinogen. Furthermore, if the immune serum is diluted, the formed precipitate becomes comparatively weaker and disappears entirely if dilution is increased. It is, therefore, generally considered that the precipitate originates from the immune serum.

## CHAPTER XII.

### BACTERIOLYSINS AND HEMOLYSINS (CYTOLYSINS).

If a guinea-pig is immunized with living or dead bacteria, for instance cholera or typhoid, and then to test its immunity is injected with a single fatal or many fatal doses of living bacilli, the animal remains alive; whereas a normal-control animal, not treated beforehand, succumbs to a similar inoculation. In order to determine the forces to which the immunized animal owes its protection, Pfeiffer undertook the following experiment: Two guinea-pigs, one immunized and another normal, were simultaneously injected intraperitoneally with living cholera vibrios, and the peritoneal exudate was withdrawn from time to time and examined microscopically in hanging-drop preparations. (The method of withdrawing the peritoneal fluid with capillary pipettes and other technical details will be described below.)

A very striking phenomenon occurred. While the cholera Pfeiffer's vibrios in the peritoneal exudate of the normal animal retained their form and motility and increased in number continuously until the animal succumbed to the infection, the bacteria in the peritoneal exudate of the immunized animal behaved quite differently; they first began to lose their power of locomotion, then their form changed, they broke up into evenly small shining masses, so-called "granula," and finally, after several minutes these also disappeared. Guinea-pigs injected with the peritoneal exudate from these infected immune animals remained healthy, and nutrient media inoculated with material from the same source remained sterile.

The above experiment is named after its discoverer, Pfeiffer, and the phenomenon itself, "bacteriolysis."

Bacteriolysis is a strictly specific process. If an animal which is immune to cholera is inoculated with typhoid bacilli, the bacteria markedly increase, as in a normal animal. The process by which this bacteriolytic force takes place is clearly demonstrated when a mixture of living cholera vibrios and blood serum of a guinea-pig, which has been actively immunized against cholera, is injected into the peritoneal cavity of a normal guinea-pig and, as a control, normal serum mixed with living cholera vibrios is inoculated into a second guinea-pig. Here the exudates on examination from time to time show that in the peritoneal cavity of the animal injected with the immune cholera serum, the same phenomena of bacteriolysis occur as described above, leading to the sterilization of the peritoneal cavity, and protection of the animal from illness. In the control animal,



however, the normal serum has no influence upon the bacteria, so that they increase rapidly and kill the animal.

It is evident then, that the bacteriolytic power resides not only in the actively immunized animal, but that it may also be transmitted to other animals by means of the former's serum. Bacteriolysis, therefore, is not a property of the tissues of the actively immunized animal, but is to be traced to specific antibodies, "Bacteriolysins" which circulate in the blood serum and body fluids.

From the above experiment it must be assumed that the phenomenon of bacteriolysis, like agglutination and precipitation, can be demonstrated also in vitro. The earlier investigations in this connection, however, were unsuccessful. Bordet was the first to obtain conclusive results and also to elucidate the cause of previous failures.

While agglutination in vitro and bacteriolysis in vivo were Bacteriolysis readily produced by mixing living bacteria with old immune in Vitro. serum, bacteriolysis in vitro did not occur under similar circumstances. But when *freshly drawn* blood serum or exudate of an immune animal was used, bacteriolysis took place in vitro also. (In fact, granule formation can be directly observed by the microscope.) When the serum becomes old—and twenty-four hours is sufficient to cause the change, it loses its bacteriolytic powers. It seems at first glance as if bacteriolysins may be active outside the body also, but that here they lead only an ephemeral existence. This view, however, is not quite correct; for "inactive" serum, which has become "ineffective" in vitro, can again produce bacteriolysis, if it is utilized to passively immunize healthy animals. Something must exist in the organism, which supplements the inactive bacteriolysins and restores their activity. This "reactivating substance" is independent of the immunizing process, since it is to be found in normal animals also. Furthermore, inasmuch as not only cholera and typhoid immune sera, but also all other immune sera and not only guinea-pig's serum but even rabbit's, horse's, and human serum may in like manner be reactivated, it is evident that the reactivating agent lacks specificity. On account of this peculiar quality of supplementing the inactive bacteriolytic serum so that it can develop its real effectiveness, Ehrlich called the reactivating substance "*Complement*." Accordingly, the complement is a normal non-specific substance which is found in the body fluids (particularly abundant in the blood serum) of every organism; its existence is evidenced either by the activation or reactivation of bacteriolytic antibodies.

Bordet demonstrated that the apparent ease with which the bacteriolysins lose their activity is to be traced not to these bodies, but to the complement. If a small amount of fresh normal serum is added to bacteriolytic serum which has become inactive, reactivation occurs in

vitro, that is to say, the bacteriolytic serum regains its ability to dissolve bacteria. The bacteriolytic power of fresh immune serum, depends, therefore, upon the fact that it contains not only bacteriolysins but also complement; the failure of old immune serum to produce bacteriolysis is accounted for by the lack of complement, while its capacity for reactivation is explained by the still present bacteriolysins.

As the above-described experiments indicate, *bacteriolysis is a complex process, which is produced by the interaction of two substances; one, the bacteriolysin, is formed through an immunizing process, and accordingly is a specific antibody of great stability, while the other, the complement, is a normal non-specific and very labile serum substance.*

The stability of the immune bacteriolysin is evident in its resistance to heat, whereas the complement is thermolabile. If freshly drawn immune serum is heated to 56° C. for one-half hour, the complement is, as a rule, rendered ineffective, while the bacteriolysin is not in any way injured; it retains its specificity, and the degree of its affinity to antigen remains unchanged. Bacteriolysins are interfered with by temperatures above 60° C. only.

Concerning the finer mechanism of bacteriolysis there are two opposing views, that of Bordet and of Ehrlich. Without considering too closely the remarkable researches of these two investigators, the synonyms for bacteriolytic antibodies usually found in the literature will be reviewed.

In attempting an explanation for bacteriolysis, Bordet has recourse to certain phenomena in staining technique. There are some substances which can be stained only when prepared in a definite way by means of another substance, a so-called mordant ("Beize") which itself is not a stain. According to Bordet, the specific substance produced by immunization represents a kind of mordant which "sensitizes" the bacteria to the action of the second normal non-specific substance; the latter is really the active agent in causing the dissolution of bacteria and is called by Bordet "alexin"—an older term used by Buchner—in contradistinction to "substance sensibilatrice."

Ehrlich, on the other hand, advocates a more chemical conception of the essential process of bacteriolysis. He believes that the substance formed by immunization, which, for the sake of brevity, is called the immune body, is characterized primarily by the fact that it has two binding groups. One of these has a chemical affinity for the bacterial cell and is, therefore, known as the "cytophile group," the other is characterized by its binding affinity for complement and is, therefore, known as the "complementophile" group. Also because of its two binding groups (receptors) the immune body itself is called amboceptor, that is, double receptor.

Thus, according to Ehrlich, bacteriolysis takes place in the following way: The cytophile group of the amboceptor, which is strictly specific for its antigen, attaches itself to the antigen, for instance the cholera vibrio; while the complementophile group binds the complement. The complement must be regarded as a sort of digesting (proteolytic) ferment. Although it is always present in normal serum, it is not effective, because bacteria have no affinity for it. Only through the medium of the amboceptor (Zwischen-Körper, intermediary body), can complement bind itself to bacteria and dissolve them.



The specificity of the bacteriolytic process depends, therefore, on the specificity of the cytophile group, while the complementophile group possesses no or, strictly speaking, only slight specificity; it adapts itself to the complements of very many though not quite all kinds of animals.

Recent experiments have proven that the complement consists of two different parts, the middle piece and the end piece.

### Technique of Bacteriolytic Experiments.

To determine the occurrence of bacteriolysis there are two methods of procedure:

1. Pfeiffer's experiment.
2. The bactericidal plate method.

#### I. The Pfeiffer's Experiment.

The essentials of Pfeiffer's experiment have been described at the beginning of this chapter. Briefly, it consists in injecting intraperitoneally, in a normal animal, bacteriolytic immune serum mixed with living bacteria. The resulting bacteriolysis is studied microscopically by withdrawing small amounts of peritoneal exudate from time to time. If this experiment is performed with various dilutions of immune serum, and if it be determined at what dilution bacteriolysis fails to occur, then the bacteriolytic titer is evident.

The details can best be understood by taking a practical example. It is desired to find the bacteriolytic titer of the serum of a patient recovering from typhoid fever by means of the Pfeiffer experiment.

To accomplish this task the following ingredients are needed:

1. A strain of bacillus typhosus of known virulence for guinea-pigs.
2. Patient's serum, sterile, and free from complement.
3. Guinea-pigs of 250 gms. weight.

A preliminary experiment must be performed in order to determine the virulence of the typhoid strain.

#### TESTING THE VIRULENCE OF STRAIN.

Guinea-pig No. 1.	1/II '09 One loopful of a typhoid agar culture suspended in 1 c.cm. of bouillon, injected intraperitoneally.	2/II dead.
Guinea-pig No. 2.	1/II '09 One-half loopful of same.....	2/II dead.
Guinea-pig No. 3.	1/II '09 One-fifth loopful of same.....	2/II dead.
Guinea-pig No. 4.	1/II '09 One-eighth loopful of same.....	2/II sick. 4/II dead.
Guinea-pig No. 5.	1/II '09 One-tenth loopful of same.....	2/II sick. 3/II well.

As far as the Pfeiffer experiment is concerned the titer of virulence in this case is  $1/5$  of a loopful of an agar culture because this dose is fatal within twenty-four hours. In order, however, to make sure of excluding all individual variations, which can and occasionally do occur, it is advisable to use not the titer dose, but its fifth or tenth multiple, that is, in this case, one loopful.

Doses larger than one loopful should be avoided, so that if any particular strain of typhoid bacilli is not sufficiently virulent, necessitating the use of larger doses, the virulence must first of all be increased. This is done by passing the organism through animals such as guinea-pigs.

The method is as follows: A very large dose of the culture, for example To Increase the surface of an entire agar tube, is injected intraperitoneally. Every the Virulence. animal succumbs to this enormous dose. The bacteria-laden exudate from the abdominal cavity, which, of course, must be removed under sterile precautions, is then inoculated into a second guinea-pig and when it dies, into a third, and so on. As a rule, after passing through one or two animals the bacterial strain (which must be grown pure from the cadaver) becomes more virulent, as can be proven by titration. Very often the virulence is increased exclusively for the species of animal used and occasionally this is associated with a decrease in virulence for other species. After a series of passages through animals, the strain reaches a maximum strength beyond which it cannot be increased. The degree of virulence varies with the type of bacteria. Typhoid and cholera usually reach only a moderate virulence ( $1/10$  to  $1/20$  loopful); the bacteria of the hog cholera group can acquire a distinctly higher virulence; for instance, *B. paratyphosus*,  $1/100$  to  $1/1000$  of a loopful, while the streptococcus and pneumococcus reach the highest figures,  $1/10,000$  to  $1/1,000,000$  of a loopful.

For the Pfeiffer's experiment with cholera or typhoid, the most suitable strains are those of such a virulence that  $1/5$  to  $1/10$  of a loopful injected intraperitoneally kills in twenty-four hours.

The serum to be investigated is freed of its complement by Technique of heating in a water-bath for one-half hour at  $56^{\circ}$  C. Then a Pfeiffer's series of dilutions are made in bouillon (not in salt solution) Experiments. for instance  $1/10$ ,  $1/100$ ,  $1/1000$ , etc. A c.cm. of each dilution is put into a test-tube (a sterile pipette should be used) and rubbed up with a standard loopful of an 18 to 24 hour agar culture of typhoid bacteria. Finally the contents of each test-tube are injected intraperitoneally in a guinea-pig of 250 gms. weight.

Inasmuch as small amounts are apt to be lost when aspirating the fluid with the syringe as well as when pouring the bacterial emulsion into a watch glass, it is better to rub up two loops of the culture in 2 c.cm. of bouillon instead of 1 loop in 1 c.cm., and then withdraw only 1 c.cm. for use in the experiment.

The following controls should be prepared:

1. Dilutions of the serum of a normal person (or animal of the same type) + typhoid culture.



2. Dilutions of immune serum + a heterologous culture.
3. (a) Bouillon + typhoid-culture.  
(b) Bouillon + heterologous culture.

The study of the bacteriolytic phenomena follows the inoculation. For this purpose capillary pipettes to withdraw the peritoneal exudate are prepared according to the directions of von Issaeff.

A thin glass tube is heated in a Bunsen flame almost to the melting point, then removed from the flame and immediately drawn out with a sudden jerk. Very fine capillary pipettes can thus be made.

The removal of the exudate is accomplished as follows: a small cut is made with scissors through the skin of the guinea-pig's abdomen; the capillary pipette, the large end of which is kept closed with the index finger, is forced into the abdominal cavity with a single push. The pressure of the finger is next relaxed and the tube slowly withdrawn. In order to avoid injuring the intestines, the precautions usually advised in intraperitoneal inoculations should be observed here. The author has found Friedberger's method of holding the animal very serviceable (see Fig. 5). The procedure is absolutely painless, moreover, the ordinarily sensitive guinea-pigs withstand the operation almost without uttering a sound.

It is best to withdraw the exudate immediately after the injection and then at intervals of five to ten, twenty, and thirty minutes, etc. Observations are made directly in hanging-drop preparations. Stained specimens are less reliable and instructive because, according to the investigations of Radziewsky, the findings are dependent upon the kind of coloring matter used. Bacteria which are in the process of dissolution soon lose the power of being stained by methylene blue, while they retain their affinity for carbol-fuchsin and aqueous solution of gentian violet. Granules are demonstrated only incompletely in stained preparations.

The prognosis for the animal *quoad vitam* is unfavorable, if bacteriolysis does not occur; good, if it does. Yet there are exceptions to the latter rule, a subject to which reference will be made later on. Now that the most important technical details of the Pfeiffer phenomenon have been considered, the following protocol will more clearly illustrate the exact procedure.

*Titration of a bacteriolytic serum (after Pfeiffer).*

Guinea-pig 1-4-07 No. 1.	One loopful of typhoid culture.	+0.1 typhoid serum.	Beginning of bacterioly- sis after 10 minutes; after 30 minutes exu- date was sterile.	Animal remained alive.
	in 1 c.cm. of bouillon intraperi- toneally.			
Guinea-pig 1-4-07 No. 2.	One loopful of typhoid culture.	+0.01 typhoid serum.	Beginning of bacterioly- sis after 15 minutes; after 20 minutes only isolated, non-motile bacteria, many gran- ules; after 40 minutes sterile.	Animal remained alive.
	in 1 c.cm. of bouillon intraperi- toneally.			
Guinea-pig 1-4-07 No. 3.	One loopful of typhoid culture.	+0.001 typhoid serum.	Beginning of bacterioly- sis after 15 minutes; after 20 minutes nu- merous granules and also many non-motile bacteria; after an hour, no bacteria at all.	2/4 animal slightly ill. 3/4 animal recovered and re- mained alive.
	in 1 c.cm. of bouillon intraperi- toneally.			
Guinea-pig 1-4-07 No. 4.	One loopful of typhoid culture.	+0.0001 typhoid serum.	After 20 minutes gran- ules and many motile bacteria; after 1 hour motile bacteria very numerous.	2/4 animal found dead.
	in 1 c.cm. of bouillon intraperi- toneally.			
Guinea-pig 1-4-07 No. 5.	One-fifth loopful in 1 c.cm. of bouillon intra- peritoneally.		After 20 minutes gran- ules and many motile bacteria; after 1 hour motile bacteria very numerous.	2/4 animal found dead.
Guinea-pig 1-4-07 No. 6.	One loopful of typhoid culture.	+0.1 normal serum.	After 15 minutes many granules, also isolated motile and non-motile bacteria; after 20 min- utes motile bacteria; after 30 minutes motile bacteria very numer- ous.	2/4 animal found dead.
	in 1 c.cm. of bouillon.			
Guinea-pig 1-4-07 No. 7.	One loopful of bacteria, para- typhosus. (Vir- ulence 1/10 loopful.)	+0.1 typhoid serum.	Complete similar find- ings.	2/4 animal found dead.
	in 1 c.cm. of bouillon.			

The bacteriolytic titer of the tested serum in this case would lie between 0.001 c.cm. and 0.0001 c.cm. and could be exactly determined by further tests which would take into consideration the intermediate doses.



On close study of the above experiment, it will be noted that even in those cases in which the animals died of the infection, bacteriolytic phenomena were not altogether absent. They occurred particularly in the beginning and were incomplete. This can be considered as evidence of the fact that even normal animals possess a certain supply of bacteriolysins which are, however, readily exhausted. This amount of normal bacteriolysin in serum varies greatly with the species of animal; thus the sera of man and rabbit contain very little normal bacteriolysins for cholera and typhoid, while horse's serum is well supplied with the same.

According to Kolle, a loopful of virulent cholera vibrios is destroyed in the peritoneal cavity of a guinea-pig, by

0.005 to 0.01 c.cm. of normal horse's serum.

0.01 to 0.02 c.cm. of normal ass serum.

0.02 to 0.03 c.cm. of normal goat's serum.

0.1 to 0.3 c.cm. of normal rabbit's serum.

The protective action of bacteriolytic sera differs very essentially from that of antitoxic sera. For the latter, the law of multiple proportions holds true; a stronger dose of toxin is neutralized by a proportionately larger amount of antitoxin; to bacteriolytic sera this rule does not

Endotoxin. apply. If the bacteria are increased beyond a certain quantity, their dissolution can indeed be accomplished by the addition of sufficient amounts of bacteriolysins, but the animal dies nevertheless. Its peritoneal cavity examined during life or postmortem may be absolutely sterile. Pfeiffer's explanation for this phenomenon is that the endotoxins within the bacteria are liberated by bacteriolysis and kill the animal. Fatal results from endotoxin follow in a similar manner when dead instead of living bacteria are injected.

Since endotoxins can continue their effective action in spite of the serum, it is evident that the usual bacteriolytic serum lacks the power to neutralize the poisons of the endotoxins. Many investigators have attempted to supply this deficiency. (This will be considered later.)

While bacteriolysis may take place without any resulting protective action, on the other hand a serum may be curative in spite of the *absence* of bacteriolysis. This is well demonstrated in Metchnikoff's experiment.

A marked leucocytosis in the abdominal cavity of a guinea-pig is produced by the intraperitoneal injection twelve hours previously of 5 to 10 c.cm. of aleuronat solution or sterile bouillon. Pfeiffer's experiment is then performed. As a rule, bacteriolysis occurs up to a certain point, particularly when cholera vibrios are used; most of the bacteria, however, retain their form and are taken up by the leucocytes.

Metchnikoff used this experiment to uphold his theory of the signifi-

cance of phagocytosis. Pfeiffer maintained that bacteriolysis was the most important protective weapon of the immune organism against bacterial invasion. According to Metchnikoff and his followers among whom Bail in particular must be mentioned, bacteriolysis in the abdominal cavity is only an exceptional phenomenon (test-tube experiment in vivo); its occurrence is made possible by the circumstance that the abdominal cavity is as a rule almost free of wandering cells, and that the few which are present are so injured by the severity of the infection that they disintegrate. If their number increases, bacteriolysis does not occur, or at least is only slight. Likewise, bacteriolysis is incomplete in the presence of cells, for instance in the blood, spleen, liver and subcutaneous tissue, etc.

A detailed consideration of this much mooted problem does not fall within the compass of this book. It is sufficient to have pointed out the great questions of fundamental significance which hinge upon the discussion of the Pfeiffer experiment, questions which concern the essential features of antibacterial immunity. It can be readily understood, therefore, why the phenomenon of bacteriolysis has been so much studied, although its practical significance is only limited.

The Pfeiffer experiment can be used for the differentiation of bacteria as well as for the demonstration of bacteriolysins in serum. It serves as a control for the agglutination reaction.

The Practical Application of the Pfeiffer Experiment. Pfeiffer and Kolle, Brieger and others, have used bacteriolysis as a method of estimating the immunity obtained by active protective immunization against cholera and typhoid in man. It must, however, be questioned whether it is admissible to draw conclusions as to the degree of active immunity from the height of the bacteriolytic titer of the serum, inasmuch as animals are found which possess no active immunity and still have sera of high bacteriolytic powers.

*The most important practical use of the Pfeiffer experiment lies in the identification of suspected cholera cultures.* In Germany, the Pfeiffer test made with the vibrios obtained in pure culture from the suspected patients is required for the official diagnosis of the first cases of cholera.

The serum used for this purpose should be at least strong enough in amounts of 0.0002 c.cm. to cause the disintegration of the bacteria in one hour, when a mixture of one loopful of an eighteen-hour agar culture of cholera in 1 c.cm. of nutrient bouillon is injected into the peritoneal cavity of a guinea-pig.

For this experiment four guinea-pigs of 250 gms. weight are used.



Animal	Culture	Serum	Method of injection	Result in cholera cases
No. 1.....	One loopful of an 18 hours' growth of culture suspected to be cholera in 1 c.cm. of bouillon.	0.001 c.cm. cholera serum=5 times the titer dose.	Intraperitoneally.	After 20 minutes or at the latest 1 hour, bacteriolysis occurs; animal remains alive.
No. 2.....	One loopful of an 18 hours' growth of culture suspected to be cholera in 1 c.cm. of bouillon.	0.002 c.cm. cholera serum=10 times the titer dose.	Intraperitoneally.	After 20 minutes or at the latest 1 hour bacteriolysis occurs; animal remains alive.
No. 3 (control).	One loopful of an 18 hours' growth of culture suspected to be cholera in 1 c.cm. of bouillon.	0.01 c.cm. of normal serum=50 times the titer dose of the immune serum.	Intraperitoneally.	Increase in number of bacteria; animal dies.
No. 4 (control of virulence of culture).	One-fourth loopful of an 18 hours' growth suspected of being cholera in 1 c.cm. of bouillon.		Intraperitoneally.	Increase in number of bacteria, animal dies.

In cases of subsiding cholera, the Pfeiffer experiment is performed with the serum of the patient in dilutions of 1 to 20, 1 to 100 and 1 to 500.

Bacteriolysis with typhoid organisms is less typical than with cholera. For diagnostic purposes the test is resorted to, only when the agglutination reactions are doubtful. When bacteriolysis also gives uncertain results, an animal is immunized with the typhoid suspected bacteria and its serum tested for its power of agglutinating or destroying definitely known typhoid bacteria and eventually the immunized animal may be injected with virulent typhoid bacilli.

Bacteriolysis is even more unsatisfactory with bacillus paratyphosus B. and the related hog cholera group of organisms.

While with typhoid bacteria the onset of bacteriolysis offers a favorable prognosis for the animal, guinea-pigs inoculated with bacteria of the paratyphoid-hog-cholera group die in spite of complete bacteriolysis. Death always takes place late (from three to six days), while the control animals succumb in about twenty-four hours. Bacteriolysis has also been observed with the bacillus of dysentery and with the tubercle bacillus; but thus far, these phenomena have gained no clinical significance. Bacteriolysis does not occur in anthrax, pest and the various diseases due to cocci.

## II. Bactericidal Plate-culture-method.

(Plattenverfahren) *according to Neisser and Wechsberg.*

For the determination of the bactericidal titer of a serum, *Neisser and Wechsberg recommended the so-called bactericidal plate-culture method.* The principle of it is as follows: the serum to be tested is inactivated; different amounts of this inactivated serum are mixed with a definite constant quantity of bacteria, and a constant quantity of active normal serum is added as complement. This mixture is left in the thermostat sufficiently long to permit the occurrence of bacteriolysis. Now, to determine whether and to what degree death of bacteria resulted from the effect of the reactivated bacteriolysins (or of some bactericidal substance otherwise unknown), agar is added, the mixture plated, and the number of colonies counted.

Stern and Korte recommend this procedure for clinical purposes, as a substitute for the Pfeiffer test in the diagnosis of typhoid fever. They point out the sparing of animals as one of its advantages. On the other hand, this method consumes much more time and its results are less trustworthy. It has not found a place, therefore, in clinical practice.

The Tech- of the patient, and that of a person not ill with typhoid as  
nique of control, are inactivated for one-half hour at 56° C. and 1 c.cm.  
the Method. of each in decreasing dilutions is poured into sterile test-tubes.

To each is added 0.5 c.cm. of a twenty-four-hour typhoid bouillon culture diluted in bouillon to 1:5000 or 1:10000. For reactivation 0.5 c.cm. of fresh normal rabbit's serum in a dilution of 1 to 12 in physiological saline is added and the whole thoroughly shaken. The tubes are then placed in the thermostat for three hours. The entire contents of each mixture is plated on agar, and after eighteen to twenty-four hours the plates are to be examined. That particular plate is considered to indicate the extreme limit of the bacteriolytic action of the serum in which there is still evident a very great decrease in the number of colonies as compared with the innumerable colonies found on the control plates.

Certain other controls are necessary:

1. One tube containing culture and complement.
2. One containing culture and inactivated immune serum in the highest concentration used.
3. The same with inactivated normal serum instead of immune serum.
4. Complement without culture and immune serum to test its sterility.
5. Immune serum without culture and complement to test its sterility.
6. One tube containing only culture, to be plated immediately.
7. One tube, containing only the culture, to be plated after standing in the thermostat for three hours.



Töpfer and Jaffé pour a thin layer of agar into a petri dish and let it harden. Upon this the culture-serum-agar mixture is poured, and after hardening is covered with another thin layer of agar. In this way the formation of a film of culture in the water of condensation is avoided.

A practical example is appended to illustrate the plate culture method.

Culture	Serum	Complement	Result (poured after remaining 3 hours in the thermostat)	
			Normal serum	Immune serum
0.5 c.cm. 1/5000 typh.	1/100 c.cm.	0.5 c.cm. 1 : 12 rabbit's.	0 colonies.	Many thousand.
0.5 c.cm. 1/5000 typh.	1/500 c.cm.	0.5 c.cm. 1 : 12 rabbit's.	100 colonies.	Many thousand.
0.5 c.cm. 1/5000 typh.	1/1000 c.cm.	0.5 c.cm. 1 : 12 rabbit's.	Many thousand.	Many thousand.
0.5 c.cm. 1/5000 typh.	1/5000 c.cm.	0.5 c.cm. 1 : 12 rabbit's.	∞	Many thousand.
0.5 c.cm. 1/5000 typh.	1/10000 c.cm.	0.5 c.cm. 1 : 12 rabbit's.	∞	500
0.5 c.cm. 1/5000 typh.	1/20000 c.cm.	0.5 c.cm. 1 : 12 rabbit's.	∞	200
0.5 c.cm. 1/5000 typh.	1/30000 c.cm.	0.5 c.cm. 1 : 12 rabbit's.	.....	0
0.5 c.cm. 1/5000 typh.	1/40000 c.cm.	0.5 c.cm. 1 : 12 rabbit's.	.....	5
0.5 c.cm. 1/5000 typh.	1/50000 c.cm.	0.5 c.cm. 1 : 12 rabbit's.	.....	60
0.5 c.cm. 1/5000 typh.	1/100000 c.cm.	0.5 c.cm. 1 : 12 rabbit's.	.....	800
0.5 c.cm. 1/5000 typh.	1/200000 c.cm.	0.5 c.cm. 1 : 12 rabbit's.	.....	∞
Control I	—	0.5 c.cm. 1 : 12 rabbit's.	Many thousand.	
0.5 c.cm. 1/5000 typh.				
Control II and III	1/100 c.cm.	—	∞	∞
0.5 c.cm. 1/5000 typh.				
Control IV	—	0.5 c.cm. 1 : 12 rabbit's.	0	
Control V	1/100 c.cm.	—	0	
Control VI	—	—	Many thousand.	
0.5 c.cm. 1/5000 typh.				
immediately poured.				
Control VII	—	—	∞	
0.5 c.cm. 1/5000 typh.				
poured after 3 hours.				

In addition to the results which one would expect, this experiment shows one striking point. With the normal serum the tube which contains the largest amount of normal bacteriolysins shows, on plating, the fewest germs. The greater the dilution of the serum the more prolific is the bacterial growth. The titer of the normal serum in this case lies between 1/100 and 1/500. The controls show that the serum and complement are sterile, and that the inactive normal serum is ineffective. During the three hours in the thermostat the bacterial suspension has become stronger. The retarded growth in the complement culture tube can be traced probably to the presence of normal bacteriolysins.

With the immune serum, on the other hand, results are quite different. Where the most concentrated serum is used, the bacterial growth is still rather profuse; only the moderate doses show a true bactericidal action and the small doses are altogether ineffective. The titer of this serum is between 1/30,000 and 1/40,000.

Neisser and Wechsberg explain this phenomenon by the so-called "*deviation of the complement.*" They assume that in the serum of higher

concentration there are so many amboceptors that the bacteria cannot bind them all. The amboceptors remaining free attach themselves to the complement by means of their complementophile group just as the already bound amboceptors have done. Thus, a part of the complement is deviated from the bacteria and only an incomplete bacteriolysis takes place.

The theory of complement deviation does not in the opinion of the author withstand critical examination. Particularly the evidence brought forward by Bordet and Gengou that the affinity of complement for the bacterium + amboceptor complex (Sensitized bacterium) is considerably greater than for free amboceptor, militates against the view of Neisser and Wechsberg.

It is possible that agglutination may account for the phenomenon of deviation of the complement in that the agglutinated masses of bacteria afford a more resistant barrier to the action of the bacteriolysins. The author has now and then observed an analogous phenomenon in hemolytic experiments; strong doses of hemolysin were less effective than moderate ones, and in these cases the momentary hemagglutination was readily visible. Also, by titrating bactericidal sera in animal experiments, it has been found that moderate doses often afforded the greatest protective action.

For the practical application of the plate culture method, knowledge of the following data is important, as it is necessary to consider the difference between the bactericidal titer of sera of normal and of typhoid patients. According to Korte and Steinberg the bactericidal titer was

	Of normal cases	Of typhoid cases
Under 100 in.....	74 per cent.	0.0 per cent.
Between 100 and 1000 in.....	8.6 per cent.	3.3 per cent.
Between 1000 and 10,000 in.....	15.4 per cent.	15.1 per cent.
Between 10,000 and 100,000 in.....	2.0 per cent.	23.3 per cent.
Over 100,000 in.....	0.0 per cent.	58.3 per cent.

In typhoid fever the bactericidal titer does not run strictly parallel either with agglutination or the Pfeiffer experiments. It falls toward the end of the disease and is low during convalescence.

Besides its use in typhoid fever, the plate culture method has been employed for experimental purposes in cholera and dysentery; in these diseases, however, it possesses no clinical diagnostic significance.

Concerning bacillus paratyphosus B, the views of different authorities are widely at variance. While some obtained very good results, similar to those found in typhoid fever, Töpfer and Jaffé could demonstrate no bactericidal power whatever in vitro. This difference can be explained only by variations in sera.



## Hemolysins.

An animal that is injected with the red blood cells of a different species develops in its serum antibodies which are biologically analogous to bacteriolysins and differ from them only in that they cause disintegration of erythrocytes instead of bacteria. These antibodies are therefore called hemolysins, or to be more precise immune-hemolysins, since they arise through a process of immunization. The breaking up of the red blood corpuscle, hemolysis, is recognized by the naked eye. The hemoglobin passes from the erythrocytes into the surrounding fluid (serum or physiological salt solution) and colors it red. The previously opaque blood lakes and becomes transparent. Immune-hemolysins like bacteriolysins belong to the class of amboceptors. They are relatively thermostable in that they withstand a temperature of from 56° to 58° C. without being injured; they require complement for the development of their hemolytic action. Furthermore, immune-hemolysins, like all amboceptors, are specific, *i.e.*, the serum of a rabbit immunized against horse's blood can dissolve only the blood of a horse and not that of a hen or cow. On the other hand, group reactions occur here also; for instance the immune-hemolysin produced in a rabbit against horse's blood is likewise active against donkey's blood.

Just as various antitoxins, agglutinins, precipitins and bacteriolysins can be found in normal serum, so also can normal hemolysins of amboceptor structure be discovered in the blood of many animals.

While normal hemolysins come into play in only a few reactions, as in several modifications of the Wassermann test, the significance of immune-hemolysins is extraordinarily great. These antibodies, discovered by Bordet, and independently by von Dungern and Landsteiner, were carefully studied by Ehrlich and Morgenroth and many others. Such researches have, first of all, greatly advanced the subject of immunity in its theoretical aspects, in that they have created the possibility for the discovery in minute detail the finer relationship which has explained some of the phenomena occurring in bacteriolysis. Furthermore, the studies of hemolysins led to the discovery of the complement fixation method, a procedure of exceptional practical value.

As far as the technique for obtaining immune-hemolysins is concerned, the rules which hold for every process of immunization are naturally to be followed here also. It is not possible, however, to immunize every kind of animal against every type of red blood corpuscle. Rabbits, goats, horses and chickens are the ones which are best adapted to supply hemolytic sera. An animal produces a better hemolysin the remoter its relationship to the

animal from which the erythrocytes for injection are taken. The blood to be injected can be employed in just the condition in which it flows from the vein. Nevertheless it is as a rule defibrinated, to prevent coagulation. The simplest and most practical way of doing this is to place some glass beads into a bottle or Erlenmeyer flask and then sterilize it by dry heat. The blood coming from the vein is allowed to flow into one of these flasks and then it is repeatedly shaken for several minutes. This suffices to defibrinate the blood and thus prevent coagulation.

The production of hemolysins depends entirely upon the red blood corpuscles. The presence of the serum is not only superfluous, but even harmful, as experience has shown that dangerous reactions may follow the injection of foreign serum.

Before injecting, therefore, the erythrocytes are washed. For this Washing of purpose a few cubic centimeters of defibrinated blood are poured into Red Blood a centrifuge tube and the level of the fluid marked on the tube. An Corpuscles. equal or double this amount of 0.85 per cent. saline is added, and the tube rapidly centrifugalized. The erythrocytes fall to the bottom, while the upper layers of the tube consist of diluted serum more or less tinged with hemoglobin. The fluid is carefully decanted, fresh saline added, the tube gently shaken, and again centrifugalized. If this is done two to three times the erythrocytes can be freed of the last traces of serum; finally, by adding saline up to the mark made at the beginning of the experiment, the erythrocytes are obtained in the normal concentration, just as in the blood, but completely free of serum.

Immuniza- The washed, defibrinated blood can be injected subcuta-  
tion. neously, intravenously, or intraperitoneally. With the subcutaneous and intraperitoneal methods in a rabbit, injections of from 5 to 20 c.cm. are necessary at intervals of five to six days. Far larger quantities should be given to bigger animals, like goats and sheep. Subcutaneous injections often cause infiltrations and occasionally abscesses. The author therefore uses the intravenous method in rabbits.

A suspension of washed blood corpuscles is diluted four to five times with physiological saline; 0.5 to 1.0 c.cm. of this fluid is slowly injected into the ear vein every five to six days. Three injections are almost always sufficient for procuring a good serum. The animals sustain the first two injections with ease, but the third and following ones are not altogether without danger. This is supposed to be akin to anaphylactic phenomena. It is therefore advisable to immunize several animals simultaneously, so that in case one dies there is another to replace it. Furthermore there are such marked individual variations in the ability to produce hemolysins that it is best to have several animals to choose from. Beginning on the sixth day after the third injection, blood should be withdrawn for the determination of the hemolytic strength and this process repeated daily until the titer has reached a satisfactory height and then the animal should be bled. If only a small amount of hemolysin is needed, the animal can be allowed to live; it will gradually lose its titer completely and will act apparently like a normal



animal. Nevertheless, an essential difference exists. For if the animal previously immunized is again injected, hemolysins reappear after a short incubation period, whereas in a normal animal a prolonged immunization is necessary. Hemolysins, therefore, exist to a certain extent in a preformed state in the cells of an immunized animal. If a stimulus to immunization occurs, the hemolytic substances are thrown off into the circulation, while in a normal animal the formation of hemolysins by the cells must first take place.

If a great amount of hemolysin of the same titer is needed, it is best to bleed the animal to death. For the preservation of hemolysins the author recommends the following procedure which he has found very trustworthy. One to 3 c.cm. of serum obtained sterile are poured into sterile tubes, which are closed with non-absorbent cotton. The tubes are placed into a water bath at 56° C. for one-half hour to inactivate the serum and are then covered with sterile rubber caps. (These are sterilized by placing them in a 1 per cent. sublimate solution for forty-eight hours.)

An immune hemolysin must answer both qualitative and quantitative determinations; qualitative, whereby is proven that the serum can hemolyze only the red blood cells which serve as antigen or to a slight degree those of related animals, and that it has only the effect of a normal serum upon the erythrocytes of other animals. The quantitative estimation supplies the only means for the absolute differentiation between a normal and an immune serum. In complement fixation where hemolysis bears an active part, it is the quantitative use of the hemolysin which decides the result of the reaction. The immune serum must therefore be "titrated."

If fresh active hemolytic immune serum is used, a constant quantity of blood serving as antigen is mixed with decreasing quantities of the immune serum and the mixtures placed in the thermostat. Results like the following will be obtained.

Antigen blood	Hemolytic serum of immune rabbit	Result after 2 hours
1 c.cm. of 5% sheep's blood.....	1 c.cm. of active serum, 1 to 10	Hemolysis.
1 c.cm. of 5% sheep's blood.....	1 c.cm. of active serum, 1 to 20	Incomplete hemolysis.
1 c.cm. of 5% sheep's blood.....	1 c.cm. of active serum, 1 to 50	Incomplete hemolysis.
1 c.cm. of 5% sheep's blood.....	1 c.cm. of active serum, 1 to 100	No hemolysis.

On the basis of this experiment the titer of the hemolytic serum for sheep's blood would lie between 1/10 and 1/20. But this is incorrect, as it was pointed out previously that by immunization only the amboceptors are increased and the complement remains unchanged. Each of the above

dilutions decreases therefore not only the amount of hemolysin, the quantitative estimation of which is the object of the experiment, but also the complement. Inasmuch as the latter was not at first increased, a point is soon reached where there is no complement at all in the diluted fluid; as a result hemolysis cannot occur, for only the combination of hemolysin + sufficient complement can exhibit any hemolytic action. *Correct titration consists therefore in allowing varying quantities of hemolysin with a constant amount of complement to act upon a constant quantity of red blood cells. The simplest method of accomplishing this is first to destroy the complement by inactivation of the hemolytic serum, then to make the desired dilutions, and finally to add, to all, the same amount of normal serum as complement.* The normal serum of an animal of the same species as that which provided the immune serum can under no circumstances serve as complement. On the contrary, foreign sera are much more suitable; and guinea-pig's serum is especially recommended as complement when immune rabbit's serum is used. Not every complement serves equally well for any immune serum.

A very good hemolytic system and one which is almost exclusively used for the complement fixation reaction, is sheep's blood as antigen, rabbit's immune hemolysin as amboceptor and normal guinea-pig's serum as complement. The preparation of these ingredients should be carried out as follows:

1. *Sheep's Blood*.—This should be defibrinated and washed. Washing is necessary because fresh sheep's blood contains complement; if the blood is a few days old, washing is even more important.

Although serum which is not fresh does not contain sufficient active complement to cause the danger of superfluous complement, it nevertheless contains substances which interfere with hemolysis. Probably the existence of "complementoids" is the disturbing factor. It must be assumed that complement is composed of two biologically different parts, as is the case with toxins and ferments. One is the haptophore group, which has affinity for the complementophile group of the amboceptor and is the more stable of the two. The other corresponds to the energy group of the toxins (toxophore element) and of the ferments. Just as after the destruction of the toxophore group there remain only innocuous toxoids whose single perceptible activity consists in their ability to neutralize antitoxins, so also, after the destruction of the weakly resistant energy elements of the complement, there remain complementoids which lack the ability to activate a bacteriolytic or hemolytic amboceptor, although by virtue of their uninjured haptophore groups they bind the complementophile groups of the amboceptors. In this way they usurp the place of whatever active complement may still be present, rendering the latter inactive, and as a result hemolysis is absent or incomplete.

Following the technique of Ehrlich and Morgenroth, a 5 per cent. suspension of washed red blood corpuscles is employed to test a hemolysin.



A pipette, closed at the top by pressure of the index finger, is thrust to the bottom of the washed erythrocytes contained in the centrifuge tube; a definite amount, for instance 1 c.cm., is withdrawn and allowed to flow into a graduate. For diluting purposes (in this case up to 20 c.cm.) only isotonic or weakly hypertonic NaCl solutions may be used. If water, hypotonic or strongly hypertonic salt solutions are employed, the red blood cells disintegrate. This is not a true biological hemolysis, but depends upon physical basis. 0.85 per cent. saline is most suitable for the majority of erythrocytes (man, rabbit, guinea-pig, ox, sheep). When, instead of an isotonic salt solution, an isotonic sugar solution is made, the red cells are retained in their proper form, but the addition of hemolysin and complement produces no hemolysis. The presence of salt is indispensable for hemolysis as well as agglutination.

Undiluted, unwashed, defibrinated blood if removed sterile can be kept several days in the ice-box. The "Frigo" apparatus is unsuited for this purpose, because the thawing of the frozen blood breaks the capsule of the red blood corpuscle. The deterioration of the preserved blood is recognized by the large hemoglobin content of the serum or the violet color of the blood.

Occasionally blood left in an ice-box becomes dark. This is due to the lack of oxygen. When the 5 per cent. suspension is made and thoroughly shaken, the red color returns. Such blood of course is perfectly serviceable.

Still, it is best not to keep blood longer than four days. Blood older than that, even if apparently unchanged, possesses a lowered resistance and can give a far higher titer in hemolysin tests than fresh blood.

2. The rabbit's hemolysin must have been inactivated for one-half hour at 56° C. Dilutions are made with physiological saline.

3. Guinea-pig's complement is obtained by bleeding to death a healthy normal animal.

The blood is allowed to flow directly into a centrifuge tube and then to clot; the clear serum is obtained by centrifugalization. For titration of hemolysin it is best to use a constant dose of complement as 1 c.cm. of a 1/10 dilution. Complement can be kept for twenty-four hours in the ice-box. When older than this it suffers a distinct decrease in efficiency as complementoid is produced. (See above.) In the "Frigo," complement may be kept for weeks. Stern, however, does not recommend complement preserved in "Frigo" for use in complement fixation tests, as its affinity for amboceptor is noticeably decreased.

One c.cm. of each of the three reagents (each so diluted with saline that the desired dose is contained within 1 c.cm.) is mixed and 2 c.cm. of 0.85 salt solution is added to make the total volume up to 5 c.cm.

The following controls are absolutely necessary.

1. A test showing that hemolysin in strong dosage but without complement is ineffective;
2. A test indicating that without hemolysin complement in the dosage used is ineffective;
3. A test which shows that the NaCl solution is isotonic.

The three reagents must be thoroughly mixed by careful shaking of the tubes which are then placed in the thermostat at 37° C. and hemolysis watched for. The duration of the observation is a matter of personal preference. Only, the length of time must always be mentioned. One must say, for instance, that the titer of this hemolysin is 1:800, using 0.1 c.cm. of complement under observation for one-half hour, or it is 1:500 with 0.1 complement under observation for two hours. The time in which hemolysins work is very different. While many hemolysins of the same titer act in a few moments, others require two hours. The author has made it a rule to read the result after two hours' observation, but he notes the progress of the reaction every one-half hour in order to determine whether it is a slowly or rapidly acting hemolysin.

The following chart demonstrates the titration of a hemolysin as a preliminary experiment to the complement fixation method.

	Antigen	Amboceptor	Comple- ment	0.85 % Saline	Result of hemolysis		
					After ½ hr.	After 1 hr.	After 2 hrs.
1.	1 c.cm. of 5% sheep's blood	1 c.cm. dilution 1:10	1 c.cm. dilu- tion 1:10	2 c.cm.	complete	complete	complete
2.	"	" 1:100	"	"	complete	complete	complete
3.	"	" 1:250	"	"	complete	complete	complete
4.	"	" 1:500	"	"	Incom- plete	Almost complete	complete
5.	"	" 1:750	"	"	Almost_0	Incom- plete	complete
6.	"	" 1:1000	"	"	0	Incom- plete	complete
7.	"	" 1:1500	"	"	0	0	Incom- plete
8.	"	" 1:2000	"	"	0	0	0
Control I	"	" 1:10	.....	3 c.cm.	0	0	0
Control II	"	.....	1 c.cm. dilu- tion 1:10	"	0	0	0
Control III	"	.....	.....	4 c.cm.	0	0	0

Determining the end reaction is a source of difficulty for the beginner. Between the extreme "0" *i.e.*, entire absence of hemolysis, where the appearance of the tube corresponds to that of control III representing a suspension of red blood cells diluted with isotonic saline, and the other extreme "complete," *i.e.*, complete hemolysis, where every trace of corpuscular elements has disappeared and a fluid looking like dilute red-wine



remains, there are many intermediate stages. These intervening grades of reaction are represented by the terms almost 0, incomplete, almost complete, and similar expressions. The meaning of the terms is self-evident. How any particular tube is to be designated is of course a subjective question since the so-called transitional stages are numerous.

A few hours after the reaction is ended, a remarkable difference may be noted between the tubes in which hemolysis has occurred and those in which hemolysis has been incomplete or totally absent. In the last mentioned, the red blood cells have sunk to the bottom and above them remains a clear fluid which consists of pure saline or diluted serum (complement + immune serum) and is colored accordingly. If the supernatant fluid is richer in hemoglobin than that of the corresponding control, it is evident that some of the erythrocytes were hemolyzed and their hemoglobin set free. If the erythrocytes have collected at the bottom apparently in the same quantity as in the control tube, and form there a large deposit, a trace of hemolysis or almost 0 would be the terms used in reporting the result. Tube 7 after two hours showed incomplete hemolysis, *i.e.*, compared with control III it was noticeably clearer, but not completely transparent. After twenty-four hours there was a small mass of undissolved red blood cells at the bottom of the test-tube and above it a deep red fluid which was only slightly different from that in the tubes where the erythrocytes were completely dissolved. If this sediment should become so small that on shaking only a cloudy turbidity is produced, the result would correspond to the designations "very small sediment," "occasional erythrocytes at the bottom of the tube," or "almost complete hemolysis."

In the tubes containing inactive hemolysin without complement (control I, and in complement fixation reactions) hemagglutination can occur because the agglutinins which also exist in the serum remain active. Hemagglutination is recognized by the shaking the sediment: the erythrocytes are not equally distributed, but remain in clumps or strings and soon sink to the bottom again.

For many purposes it is desirable to titrate the complement content of a serum. The method is the same as that used in hemolysin titration, with the difference that a fixed amount of hemolysin and varying quantities of complement are employed.

The titer of this complement when employed with a hemolysin of 1/1000 strength and allowed to stay in the incubator for two hours would be 0.04 c.cm.

The complement content of the serum of a healthy guinea-pig is fairly constant. During illness the titer usually is decreased. Among healthy people the complement titer shows marked individual variations.

For hemolysis a definite quantitative relationship between hemolysin and complement is necessary.

Antigen	Amboceptor	Complement	NaCl solution	Result after two hours
1. 1 c.cm. 5 % of sheep's blood.	1 c.cm. hemolysin dil. 1 : 1000.	1 c.cm. 1 : 10 (= 0.1).	2.0 c.cm.	Complete.
2. 1 c.cm. 5 % of sheep's blood.	1 c.cm. hemolysin dil. 1 : 1000.	0.8 c.cm. 1 : 10 (= 0.08).	2.2 c.cm.	Complete.
3. 1 c.cm. 5 % of sheep's blood.	1 c.cm. hemolysin dil. 1 : 1000.	0.6 c.cm. 1 : 10 (= 0.06).	2.4 c.cm.	Complete.
4. 1 c.cm. 5 % of sheep's blood.	1 c.cm. hemolysin dil. 1 : 1000.	0.4 c.cm. 1 : 10 (= 0.04).	2.6 c.cm.	Complete.
5. 1 c.cm. 5 % of sheep's blood.	1 c.cm. hemolysin dil. 1 : 1000.	0.2 c.cm. 1 : 10 (= 0.02).	2.8 c.cm.	Incomplete.
6. 1 c.cm. 5 % of sheep's blood.	1 c.cm. hemolysin dil. 1 : 1000.	1.0 c.cm. 1 : 100 (= 0.01).	2.0 c.cm.	0
7. 1 c.c.m 5 % of sheep's blood.	1 c.cm. hemolysin dil. 1 : 1000.	—	3.0 c.c.m.	0
8. 1 c.c.m 5 % of sheep's blood.	—	1 c.cm. 1 : 10 (= 0.1).	3.0 c.c.m.	0
9. 1 c.c.m 5 % of sheep's blood.	—	11	4.0 c.c.m.	0

On the basis of the two titrations outlined above, it is estimated that at least 0.04 c.cm. complement is necessary to activate 0.001 c.cm. of hemolysin. If less complement is used with the same amount of hemolysin, hemolysis does not occur or else it is incomplete. If the quantity of hemolysin is increased for instance threefold, then it will be found that 0.02 c.cm. of complement suffices to produce hemolysis. Vice versa with an excess of complement the hemolysin titer of 0.001 c.cm. may be reduced. However, there are narrow limits to this mutual compensatory action.

### Cytotoxins, Cytolysins.

The hemolysin bodies are characteristic and important members of a general class of substances known as cytotoxins, especially investigated by Metchnikoff and his co-workers.

Just as the immunization with erythrocytes led to the production of lytic amboceptors which in connection with complement destroyed and dissolved their antigens, so in a similar manner, various substances more or less specific for their antigens have been produced through immunization with leucocytes "Leucocidin," with nerve tissue, "neurotoxin," with spermatozoa, "spermatoxin," and kidney tissue, "nephrotoxin." The proof of their action, particularly of neuro-, nephro-, and hepatotoxin is not simple. As all these cytotoxic sera have at the same time a hemolytic action, it is not easy to decide to what extent the changes in the organs observed after the injection of the cytotoxic substances are dependent upon the action of hemolysins. It must further be taken into consideration that none of these sera are absolutely specific for the organ in question. This is not surprising, inasmuch as there are widespread common group characteristics (common receptors) among the different organs serving as antigens, and only very few groups of a specific nature. The hopes which, at the beginning, were placed upon the study of cytotoxins particularly with the expectation that they would tend to become diagnostic and therapeutic methods for the treatment of malignant tumors, have as yet been unrealized. The entire field of cytotoxins urgently requires further investigation.



## CHAPTER XIII.

### THE METHOD OF COMPLEMENT FIXATION.

Its principle. Antituberculin. Ehrlich's side-chain theory. Serum diagnosis of syphilis, and diseases caused by animal parasites.

It has already been demonstrated that neither bacteriolysis nor hemolysis can take place without the presence of complement. The question therefore arises whether this complement is the same in both of these reactions or whether normal serum possesses different complements. In order to solve this, a number of very complicated experiments have been carried out by Ehrlich and Morgenroth, Metchnikoff, and Bordet and Gengou. Ehrlich and Morgenroth endeavored to show that not only do the complements of different animals of the same class vary, but that numerous complements exist within one individual serum (conception of the multiplicity of complements). Metchnikoff believed that each serum contained at least two complements, the microcytase and the macrocytase, thus enlisting the supporters of a dualistic theory. Bordet and his school, on the other hand, although agreeing with the idea that the complement varies in different animals, deny its multiplicity and contend that any given serum contains but one alexin, or complement—the theory of unity of complement. It would be superfluous to cite all the experimental data supporting these opinions, but nevertheless a review of the classical experiment of Bordet and Gengou which corroborated the existence of only one complement, thus offering the fundamental principle for the establishment of the most important method of serum diagnosis, namely, complement fixation, would not be out of place.

The Principles of Complement Fixation. Bordet and Gengou mixed in a test-tube typhoid bacteria (antigen), inactivated typhoid immune serum (amboceptor) and normal serum (complement). Union of the bacteria and immune serum first took place followed by absorption of, and coalescence with, the bacteriolytic complement contained in the normal serum. As a result, bacteriolysis occurred and the bacteriolytic complement was used up during this process. Bordet and Gengou reasoned that if the bacteriolytic and hemolytic complements were identical, then in the above mixture of typhoid bacteria, immune serum and normal serum, the hemolytic as well as bacteriolytic complement should be absent, while if the plurality of complement exists, the hemolytic complement should still be present. Accordingly, after a certain interval, washed erythrocytes and inactivated homologous immune serum were added and

hemolysis looked for. No hemolysis took place, thereby attesting to the fact that the bacteria in the first part of the test had "fixed" ("held in check") not only the bacteriolytic but also the hemolytic complement. Bordet and Gengou thereupon named this test "complement fixation" or "complement binding" (La fixation d'alexine).

With the aid of this experiment Bordet and Gengou were able to prove a number of theoretically important points. They demonstrated that *absorption of complement was not necessarily accompanied by bacteriolysis*. For example, the anthrax and pest bacteria when mixed with their respective homologous immune sera show no or only very incomplete bacteriolysis. The erroneous conclusion thus reached, to the effect that these sera contained no amboceptors, was disproved by Bordet and Gengou, who demonstrated that, 1, these sera contained amboceptors in spite of the absence of bacteriolysis, 2, the complement was absorbed, although no bacteriolysis took place.

During the process of immunization, amboceptors were found far more frequently than bacteriolysins. These two terms must not be considered as synonymous.

*Amboceptor signifies a more generic term, and one must differentiate between amboceptors of cytolytic and non-lytic properties.* Whether the difference here really depends upon the different nature of the amboceptor, or upon the construction and constitution of the antigen, is not solved.

The fixation of the complement precedes the act of bacteriolysis. The important requirement for the fixation is an antigen which has been sensitized by the attachment of the amboceptor, thus increasing the affinity toward the haptophore group of the complement. Antigen alone, or even amboceptor alone, binds the complement only very slightly or not at all. Whether the zymotoxic (energy) group of the complement manifests its activity (bacteriolysis) or not (absence of bacteriolysis) is materially indifferent for the complement fixation.

Through complement fixation, as introduced by Bordet and Gengou, one is enabled to prove the presence of specific antibodies when the antigen is known or reversely, an unknown antigen provided the specific antibody is given. This method of serum diagnosis can be widely employed, as the majority of bacteria and immune sera (with the exception of pure antitoxic sera)<sup>1</sup> when mixed homologously, give a positive reaction—the absence of hemolysis, proving the absorption of complement by the union of the antigen and its specific amboceptor. This reaction is strongly specific. If bacteria are mixed with an inactive heterologous immune serum, or with a heated normal one, not in concentrated form (normal amboceptor), and complement is added, the latter will not be fixed but remains to be taken up by the subsequently added red blood cells, and its immune serum, causing hemolysis. Hemolysis indicates that the mixed bacteria and serum are not homologous, as the complement is left free, and given a chance to unite with the added erythrocytes and hemolytic amboceptor. In the case where the bacteria are known, *e.g.*, typhoid bacilli, the occurrence of hemolysis indicates that the examined serum contains no

<sup>1</sup> Even antitoxic sera are said by Nicolle to give complement fixation reactions.



typhoid amboceptors. If the serum is known (*e.g.*, meningococcus serum) the occurrence of hemolysis proves that the bacteria under examination are not meningococci. The absence of hemolysis will in the first case point out that the unknown serum contains typhoid amboceptors, *i.e.*, is a typhoid serum; while in the second case the absence of hemolysis would bear definite evidence in favor of meningococci. The accompanying figures, 15 and 16, represent schematically the positive and negative complement fixation test.

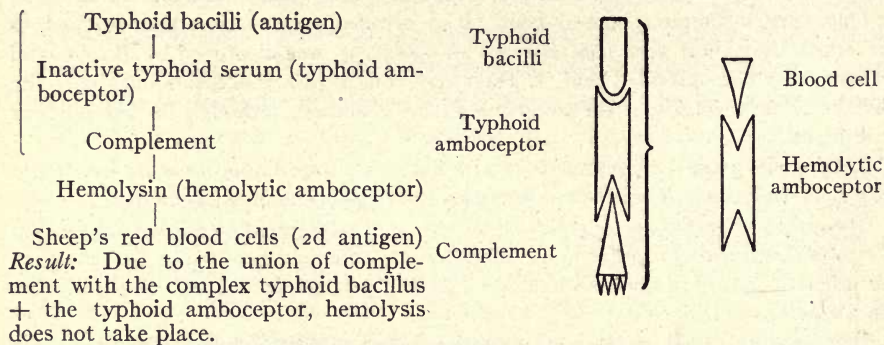


FIG. 15.

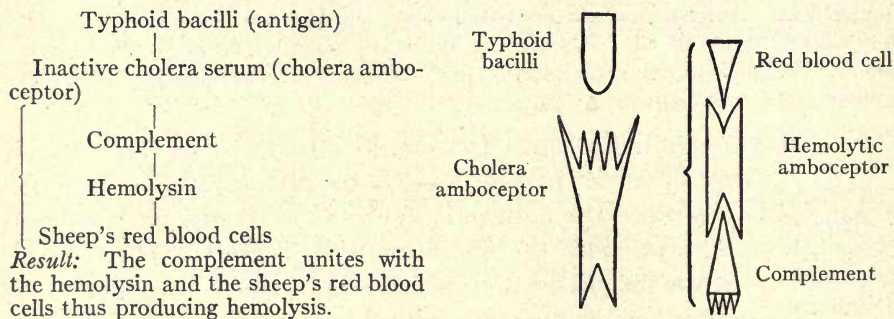


FIG. 16.

Gengou further showed that not only cellular antigens can stimulate the formation of amboceptors, but that during the course of immunization with proteids in solution (milk, serum, etc.), complement binding amboceptors are also formed in addition to the precipitins. Citron has therefore proposed the term "antigenophile," to designate the "cytophile" group of the amboceptor.

Widal and Lesourd were the first to make practical application of the complement fixation property. They found that the Bordet-Gengou reaction could be obtained far more frequently and earlier with the serum of typhoid patients than the agglutination test. Nevertheless, this entire complement fixation method remained unheeded for several years.

Moreschi (at Pfeiffer's institute), while conducting some theoretical studies concerning the nature of anticomplements, *i.e.*, such substances which tend to neutralize the action of complements, discovered anew, that by the mixture of a soluble proteid with its antiproteid serum the existing complement disappeared. This, as has been seen, can be explained by the presence within the immune serum of bodies similar to Gengou's amboceptors. Moreschi, however, stated that the complement disappeared because it was thrown to the bottom mechanically, by the occurrence of precipitation. Such a physical explanation for the complement fixation reaction led a number of authorities to the belief that the positive Bordet-Gengou reaction was in reality no amboceptor action, but a result of a similar precipitation process. Wassermann and Bruck, Liefmann, Wassermann and Citron, and later on Moreschi himself realized that this physical explanation was incorrect, inasmuch as complement fixation took place even if all precipitation was prevented by heat or other influences. Furthermore, complement binding of an unspecific nature can be produced by the mixture of glycogen or peptone with serum, a procedure wherein surely no precipitation plays any part. Finally Moreschi showed that there were strongly precipitating sera which nevertheless did not exhibit the Bordet-Gengou phenomenon.

Thus was definitely established that the complement fixation was entirely independent of either bacteriolysis or precipitation.

Following Moreschi's researches, Neisser and Sachs continued Gengou's studies and advised this demonstration of the proteid amboceptors as a control to the precipitation method for the differentiation of proteids. Its action is so much finer, and more delicate than the precipitin test that even the minutest traces of proteid can be recognized.

With the encouraging results of Neisser and Sachs in mind, Wassermann attempted by the use of highly immune antibacterial serum to discover any soluble bacterial proteids which may exist in the blood, derived from the respective bacteria invading the organism at the onset of an infection. Practical application proved that not enough of these proteids existed free in the circulation, but that they were probably bound by the tissue cells.

Wassermann and Bruck then employed the complement fixation test with the idea of demonstrating the existence of the respective antigens in the diseased organs. Tuberculous glands and lungs served as material for this experiment. They were able to obtain complement fixation when an extract of tuberculous organs as antigen was mixed with a tuberculous serum (manufactured by the Höchst Farbwerke). If instead of the latter, the serum from tuberculous individuals was substituted, no positive complement fixation reaction was obtained. On the other hand, the

Antituber- reaction was given if the human tuberculous serum employed  
culin. came from an individual who had received therapeutic inocu-  
lations of tuberculin. In other words, the serum of treated  
individuals contained, in contrast to the untreated ones,  
amboceptors against a soluble tuberculous substance also present in the  
extract of tuberculous glands. Wassermann and Bruck identified this  
substance as tuberculin, because the sera of the treated individuals gave  
the same positive results if a solution of old or new tuberculin was used in-



stead of the extract of tuberculous organs. Thus, the latter contained tuberculin while the sera of the tuberculin-treated individuals contained amboceptors designated by Wassermann and Bruck as "antituberculin." The name antituberculin has not been a well chosen one, because it creates the impression among many as being an antitoxin. It is better to speak of it as antituberculin amboceptors.

Since, according to Wassermann and Bruck these antituberculin amboceptors were not supposed to be formed spontaneously in tuberculous individuals, but only in those treated with tuberculin their demonstration could be of no apparent diagnostic value. On the other hand, their existence greatly furthered the understanding of Koch's tuberculin reaction, as most tuberculous individuals who had antituberculin amboceptors in their serum did not respond to the subcutaneous injection of tuberculin.

Wassermann and Bruck, moreover, showed that a mixture of tuberculin with an extract from tuberculous organs bound complement. From this they concluded that the extract likewise contains antituberculin amboceptors. Thus reasoning they developed their tuberculin theory.

The difference in the reaction observed in a normal and tuberculous individual after inoculation with tuberculin, can be fully explained by the Theory of presence of antituberculin amboceptors in the tuberculous focus. By Wassermann virtue of their specific affinity, the amboceptors attract the injected and Bruck. tuberculin toward them. The tuberculin and antituberculin unite, and absorb the complement from the circulating blood stream, since the complementophile group of the amboceptor is free and unbound. By virtue of the fresh complement which is an actively lytic ferment, and the attracted leucocytes, a partial destruction and casting off of the tuberculous focus results. Upon this depends the therapeutic effect of the tuberculin. During a prolonged treatment with tuberculin, the body produces an excess of antituberculin amboceptors so that finally some appear free within the blood serum. When this is the case the tuberculous organism loses its power to react toward tuberculin, as the latter is neutralized in the blood-stream at a point away from the local focus. No therapeutic effect is any longer obtained from the tuberculin injections, so that they can, for a time, be suspended. The aim of tuberculin therapy should be to work with small doses so that only a focal reaction is obtained and the hyperproduction of antituberculin amboceptors be postponed as long as possible.

Numerous exceptions were at once taken to this theory and its experimental data, the most important of which can here be mentioned.

"Summier-  
ung's Ein-  
wand" (Ex-  
ception  
taken on  
Ground  
of Summa-  
tion of  
Antigen.)

Weil and Nakayama disagreed with the proof of the existence of "antituberculin" in the organ extracts, on the basis that Wassermann had overlooked the effect of a summation of antigen. This is best explained as follows: Complement is bound not only by antigen + amboceptor, but also by large doses of antigen itself dependent upon the normally present amboceptors existing in the serum employed for complement

Old tuberculin	Complement	Erythrocytes	Hemolysin	Result
0.05	0.1	1 c.cm. 5%	Twice the hemolytic titer.	Hemolysis.
0.1	0.1	1 c.cm. 5%	Twice the hemolytic titer.	Hemolysis.
0.15	0.1	1 c.cm. 5%	Twice the hemolytic titer.	No hemolysis.

0.15 old tuberculin is thus sufficient of its own accord to bind complement. In their experiment, Wassermann and Bruck found that, for example,

Tuberculin	Extract of tuberculous organs	Complement	Erythrocytes	Hemolysin	Result
0.1	0.1	0.1	1 c.cm. 5%	Twice the hemolytic dose.	Complement fixation.
0.1	.....	0.1	1 c.cm. 5%	Twice the hemolytic dose.	Hemolysis.
.....	0.1	0.1	1 c.cm. 5%	Twice the hemolytic dose.	Hemolysis.

This, however, in no way proves the existence of "antituberculin" in the extract of tuberculous organs, as it is perfectly possible and even probable that 0.1 of the organ extract contains 0.05 c.cm. at least of tubercle bacillus substance (tuberculin) which, when added to 0.1 of tuberculin used for antigen, is sufficient to give an amount of tuberculin perfectly capable, as has been seen, of binding complement by its own activity.

In order to overcome this possibility one must work with such small but at the same time maximum amounts of antigen and antibodies, that at least double the quantity of each of these reagents does not, of its own accord, bind complement. For tuberculin this is estimated as follows:

Tuberculin	Complement	Hemolysin	Erythrocyte	Result
0.2	0.1	2×Hemolytic dose.	1 c.cm. 5%	No hemolysis.
0.18	0.1	2×Hemolytic dose.	1 c.cm. 5%	No hemolysis.
0.15	0.1	2×Hemolytic dose.	1 c.cm. 5%	No hemolysis.
0.14	0.1	2×Hemolytic dose.	1 c.cm. 5%	Incomplete hemolysis.
0.12	0.1	2×Hemolytic dose.	1 c.cm. 5%	Complete hemolysis.
0.1	0.1	2×Hemolytic dose.	1 c.cm. 5%	Complete hemolysis.

Twelve-hundredths is the maximum non-binding or hemolytic dose. For the complement fixation test where the object is to demonstrate anti-tuberculin amboceptors, the maximum amount of antigen to be used is therefore 0.06 T. or one-half of the maximum non-binding dose.

In the same way the hemolytic dose, and the dose of the antibody should be estimated.



Organ extract	Complement	Hemolysin	Erythrocyte	Result
0.2	0.1	2×Hemolytic dose.	1 c.cm. 5%	0
0.16	0.1	2×Hemolytic dose.	1 c.cm. 5%	Complete hemolysis.
0.1	0.1	2×Hemolytic dose.	1 c.cm. 5%	Complete hemolysis.

The non-binding dose is 0.16. The amount, however, to be employed in the complement fixation test must be 0.08 c.cm. of organ extract.

If on mixing 0.06 T. and 0.08 extract, complement fixation still appears, then this summation of antigen can be disregarded and an antigen antibody reaction must be considered. For, even granting that 0.08 of extract does for its greater part, *e.g.*, 0.06 at the most, consists of tuberculin, then this amount + 0.06 of the tuberculin in the antigen only makes 0.12 of tuberculin, a quantity not sufficient to fix the complement. De facto, complement fixation does occur when the above test is carried out with proper dosage, so that most probably it is occasioned by the biological antigen antibody reaction. *As a general rule for all complement fixation tests, the dose of each ingredient employed should never be more than 1/2 of its maximum quantity that does not of itself bind complement.*

A second exception, taken by Weil and Nakayama as well as

Other by Morgenroth and Rabinowitsch relates to the activity of the  
Exceptions. complement when it combines with tuberculin and anti-tuberculin.

They claim that by this union the complement's lytic function is entirely lost. Morgenroth and Rabinowitsch even go so far as to deny the existence of antituberculin in the blood of tuberculous individuals.

The author also undertook a minute study of this question and came to the following definite conclusion.—There are some tuberculous individuals who spontaneously develop antituberculin amboceptors, a fact to be expected because it has for a long time been known that on and off tuberculin can be liberated in the organism of tuberculous individuals. As a natural consequence antibodies will be formed, and most probably by those tissue cells in the neighborhood of the liberation of the tuberculin, *i.e.*, the focus of infection.

Before proceeding, however, to the author's conception of the tuberculin theory it is necessary to review Ehrlich's principles of immunity upon which the ideas of antibodies and their specificity are based.

Ehrlich's idea of the biological structure of cells is that they consist of two parts, a central functioning radicle ("Leistungs-kern") upon which depends the specialized activities of the cells, as for example, a glandular or nerve cell, and a multiplicity of side chains or receptors (a term borrowed from Ehrlich's Theory.

the chemistry of the benzol group), by means of which the cell enters into chemical relation with food and other substances brought to it by the circulation. These receptors are exceedingly numerous, as the nutritive substances upon which the cell depends for its maintenance are very varied. Besides these general receptors the special cells also have different and special side chains; then, too, there exist very great quantitative differences among the latter; and finally it must be added that the selective activity of the cells depends upon the variability of these receptors.

When an infection occurs, pathological material is brought to the cell bodies instead of physiological normal substances. Certain of these poisonous products find suitable receptors in all of the cell groups, others fit only into distinct groups of cells, while a third class are not taken up at all. The organism which possesses no receptors for any of the pathological agents cannot assimilate any deleterious substances and is therefore immune. Lack of amboceptors is therefore a natural form of immunity. The organism, having only a special group of cells for the reception of certain pathological matter, will make use of these cells for the binding and assimilation of the toxic material. For example, the nerve cells alone have receptors for tetanospasmin; no matter how or when the poison is introduced into the organism the nerve cells will absorb it. As this toxin is poisonous for the central atom group (Leistungskern) of the nerve cell, the latter is destroyed. The union between the nerve cell receptors and the tetanospasmin toxin is only the preliminary act for the cell destruction; the actual death of the cell being caused by the action of the toxophore group of the poison upon the functional radicle of the cell. If, however, such receptive side chains are possessed not only by the brain but also by other cells, *e.g.*, connective-tissue cells, the tetanospasmin will in part be bound by the latter. The toxophore group of the toxin does not have any harmful effect upon the functional radicle of these cells, and thus no toxic effects will be incurred by the union, and the nerve cells remain unaffected.

The number of receptors which cells possess for tetanospasmin, for example, are limited and after their junction with the tetanospasmin, are rendered useless and inactive. By the normal reparative mechanism of the body, new receptors are generated. This reparative process does not as a rule stop at a simple replacement of lost elements, but according to the hypothesis of Weigert tends to overcompensation. The receptors eliminated by toxin absorption are reproduced in an excess of the simple physiological needs of the cell. Continuous and increasing dosage of the toxin soon leads to such excessive production of receptors that the latter find no more room to be attached to the cell, but are cast off and circulate free in the blood. They still, however, retain their property of being able to combine with tetanospasmin.



If such an organism is injected with tetanospasmin the latter toxin is bound by the free receptors in the serum, and thus the respective "sessile" receptors attached to the cells are prevented from coming in contact with the poison. Inasmuch as the free receptors possess no functional radicle which can be injured, the toxin remains entirely innocuous for the individual. Such protective bodies lend to the organism its attained immunity and are known as antitoxins. Their function can be compared to lightning rods.

v. Behring well expresses their action when he states that the same elements, which attached to the cells render the body susceptible to toxic substances, when circulating freely in the blood serve to protect it.

The antibodies against toxins and ferments are of the simplest form. They possess only a binding group which has an affinity toward the haptophore group of the toxins and ferments. They, therefore, belong to the class designated by Ehrlich as "haptines" of the first order.

To the haptines of the second order belong the agglutinins and precipitins. They possess besides a haptophore group also an agglutinophore or precipitinophore group by virtue of which agglutination or precipitation takes place.

Belonging to the haptines of the third order are the class of amboceptors which have in addition to the haptophore group also a complementophile group for their union with the complement.

These hypotheses of Ehrlich greatly simplify the explanation of many serum reactions as well as many of the phenomena associated with the action of tuberculin. In all probability the healthy cells which exist in the tuberculous focus and which are capable of reaction, produce the antituberculin.

Christian and Rosenblatt offered experimental evidences for this statement. They demonstrated that tuberculous guinea-pigs, in whom antituberculin was produced by tuberculin injections, showed a diminution of antituberculin in the blood when tuberculous glands were removed by operation.

The antituberculin production by the cells is a transitory action arising only when tuberculin has spontaneously or artificially reached the circulation. Following this stage of activity there comes a period of quiescence during which no free antituberculin can be found in the serum. The cells, however, are supplied with a great many more sessile receptors than usually: they possess a higher affinity toward tuberculin and produce antituberculin much more readily than normal cells.

This also explains why the smallest amounts of tuberculin produce a reaction in tuberculous and not in the normal individuals. In the former, the cells in the zone surrounding the tuberculous focus are abundantly supplied with receptors, so that on the injection of tuberculin, its action

appears almost concentrated at this point. Occasionally the sessile receptors are\* relatively scarce and the first injection excites no reaction. By the time of the second or third inoculation these sessile amboceptors have so increased that a positive reaction is apparent when the same or even a smaller dose is injected. This phenomenon of increased sessile receptors explains the reappearance of subsided subcutaneous, cutaneous, or ophthalmic reactions after renewed injections of tuberculin.

To recapitulate the biological phenomena associated with a positive tuberculin reaction, it may be said that the tubercle bacilli, or portions of their body substances existing in the infected focus, stimulate the adjacent cells to produce a great number of sessile receptors. When the tuberculin is injected for the first time, these sessile receptors at once take up the tuberculin and as a result, the production of antituberculin in the focus is further stimulated.

A part of the tuberculin has already been attracted by the receptors of the cutis or subcutis cells (intracutaneous reaction) or the cells of the mucous membrane (ophthalmic reaction) and here too has stimulated the production of antibodies (antituberculin). It is only a quantitative difference in the number of receptors which actually differentiates a normal from a tuberculous individual. Thus is explained that even in non-tuberculous individuals a local reaction may be obtained if the dose of tuberculin injected is large enough; a focal reaction, however, will be given only by a tuberculous subject. The greater the number of sessile antituberculin receptors that have been formed in the tuberculous focus, the greater becomes the affinity of these cells toward the tuberculin; so that with the second, third, and subsequent tuberculin injections, focal reactions (*i.e.*, antituberculin productions) are more easily stimulated.

As for the origin of the fever, it is probable that a pyrotoxic substance is formed by the union between tuberculin, antituberculin and complement. This poison first isolated *in vitro* by Citron will again be referred to and belongs to the class of anaphylotoxins (Friedberger) or toxopeptids (M. Wassermann and Keyser).

Finally the antituberculin receptors become so numerous that they are detached from the cells and become free receptors. This period, however, is only transitory, as is corroborated by the difficulty connected with the demonstration of these antibodies in the focus. This free antituberculin combines with the tuberculin (spontaneously formed or injected) and attracts the complement, or the complement producing phagocytes. Uncombined complement has no effect on the tissues. It is different, however, with the phagocytes. These can without any additional help act directly upon the infected focus. If the tuberculin treatment is continued,

\* A sessile receptor is one which is still attached to its cell and not yet free in the blood.



a period arises during which the antituberculin bodies are so greatly accumulated in the local focus that they ultimately escape into the blood stream. This freely circulating antituberculin neutralizes any freshly injected tuberculin, so that such patients become refractory against even the largest amounts of it. (Tuberculin immunity.) Tuberculin immunity is not, however, in all cases to be identified with a strong antituberculin content in the serum as is demonstrated by the complement fixation method. For example, it is very difficult to produce antituberculin bodies by treatment with S. B. E., although by its use an immunity against B. E. is easily attained.

The question as to how great a rôle the antituberculin bodies play, and their exact interpretation is very complicated. The present status of our knowledge may be expressed as follows.

The administration of the various tuberculin preparations to tuberculous patients results in the formation of antibodies within their serum, which with the respective tuberculin as antigen will give the phenomenon of complement fixation in vitro. The different tuberculins are not equally efficient as antigens. Thus B. E. is the best stimulant of antibodies. Furthermore the antibodies obtained from the soluble tuberculin preparations are not identical with those from the insoluble products. Some sera fix complement only with old tuberculin, others react only with new tuberculin. The tuberculous serum of Meyer and Ruppel contains besides these two antibodies another group which gives the complement fixation test with an alcoholic extract of tubercle bacilli as antigen (Citron and Klinkert). It may be assumed that with other antigens, other complement binding antibodies will be discovered. The presence of these antibodies is of itself no criterion for the existence of a tuberculin immunity. There are tuberculous subjects who are not susceptible to tuberculin and at the same time possess no in vitro demonstrable antibodies; reversely, there are very highly susceptible tuberculous individuals with antibodies in their serum. The explanation for the last class of cases has been furnished by Citron. The author demonstrated that the antituberculin contained within the serum in certain instances raises the susceptibility against tuberculin. Thus there is a hypersusceptibility of a humoral form analogous to serum anaphylaxis, besides the hypersensitiveness depending upon the increased number of sessile receptors.

To offer an explanation for the first class of cases (*i.e.*, tuberculous subjects with no antituberculin and not susceptible to tuberculin) it has been assumed that besides complement binding agents there are also directly neutralizing or antitoxin-like bodies within the serum. Pickert and Löwenstein could demonstrate that the serum of some tuberculous patients had the property when mixed with tuberculin to so neutralize the latter

that it could no longer be used for the cutaneous reaction. They named this antibody "antikutine," and proved its existence as follows:

Twenty days after the last tuberculin injection the serum of the patient is withdrawn and mixed with tuberculin in 1, 2, 5 and 10 per cent. tuberculin serum mixtures. These are kept in the incubator for two hours, then in the ice-box for twenty hours and then used for the cutaneous reaction. The abrasions made with the v. Pirquet borer must be at least 3 cm. distant from each other, intervened by control spots. By gently stretching the skin of the forearm one can apply ten reactions over this area. Only a distinct papule formation should count as a positive reaction and observation should be continued for six days. Just as antituberculin amboceptors may be found in the serum of patients who have not had any specific treatment, so also can these antikutine bodies originate spontaneously.

Marmorek described still another antibody, found in a horse immunized with living young tubercle bacilli. This serum when mixed with the urine from patients with active febrile tuberculosis will fix complement. Examinations by Citron and Klinkert showed that here one is probably dealing with an antibody brought about by immunization against altered tuberculous tissue.

In former times a negative tuberculin reaction after a prolonged treatment was stamped as a cure of the tuberculosis, a fact obviously incorrect; for no matter how successful the tuberculin therapy may be, it cannot always be considered as a complete curative procedure. In general, that method should be adopted which makes the individual non-susceptible to the largest doses of tuberculin. It was found in practice that those patients having the greatest amount of antituberculin in their serum usually offered a better prognosis.

Serum Diagnosis of Syphilis.	The experiences gained by the employment of the complement fixation test in tuberculosis lead to its application in the study of syphilis. The difficulties in this disease were greater, inasmuch as there were no bacteria or preparations like tuberculin which could be used as antigen.
------------------------------------	--

Syphilitic human organ extracts were employed, with the idea that these would contain the specific virus. The serum of monkeys previously immunized with such extracts, when mixed in vitro with the latter, gave complement fixation. This experiment is not, however, conclusive; the positive reaction may be due to anti-human proteid amboceptors produced at the same time by the injection of the human serum contained in the organ extract. The experiment was changed and the syphilitic organ extracts from apes were used so as to exclude this error. Even in this way complement fixation was attained. Later on it was found unnecessary to inject the monkeys with the extracts since after ordinary infection their serum would give complement fixation. In this manner it was almost definitely established firstly, that these extracts contained



a substance specific for syphilis which could with most probability be considered a luetic antigen, and secondly that infected apes possess antibodies against this antigen.

The next step was to try the reaction in man. The first experiments of Wassermann, Neisser, Bruck and Schucht did not give the hoped-for returns. Although the reaction was obtained with human serum, the percentage of positive results was so small (see next chart) that its practical value as a means of diagnosis offered no great help. Only in general paralysis did the expectation seem promising. In about 80 per cent. of all cases Wassermann and Plaut were able to demonstrate the luetic antibodies in the cerebrospinal fluid.

Schütze's experiments in tabes led him to the same findings. Citron has obtained a much smaller percentage of positive reacting cerebrospinal fluids in tabes.

As it seemed that the means of diagnosis was not to be established by the demonstration of the syphilitic antibody, Neisser and Bruck believed that better results may possibly be achieved by the discovery of the luetic antigen in the serum through complement fixation.

This attempt too was unsuccessful. No antigen could be found, but the extracts of red blood cells from syphilitic individuals when mixed with the serum of highly immunized monkeys gave a positive complement fixation. Neisser and his co-workers concluded therefrom that the erythrocyte extract contained the luetic antigen. Citron soon demonstrated that the extracts of normal individuals gave a similar reaction and what was more important, that this so-called blood antigen existed in the blood entirely uninfluenced by mercurial treatment. Since these experiments, not much importance has been attached to this reaction.

Meanwhile the author working at the Kraus clinic proved by a large series of experiments that luetic antibodies were present in almost all cases of lues. The reaction is dependent upon two rules.

*The First.*—The longer the syphilis virus has acted upon the organism and the more numerous its recurrent manifestations have been, the more frequently will a positive reaction be obtained and the stronger will the antibody content of the serum be.

*The Second.*—The sooner a proper mercury therapy is instituted, the more often it is repeated, and the shorter the interval since the last treatment, the smaller will the antibody content of the serum be and the greater the possibility of a negative reaction.

These points were soon corroborated by numerous other workers in the field, so that at the present day they can be taken as absolute facts. The following chart will explain some of the statements aforementioned.

	First period					Second period							
	Wassermann, Neisser, Bruck & Schuchter	Wassermann and Plaut	Marie and Levaditi	Morgenroth and Sertiz	Schütze	Citron			Bruck and Stern	Fleischmann	Meier	Ledermann	Micheli and Borelli
						1st report	2d report	3d report (with Blaschk)					
	%	%	%	%	%	%	%	%	%	%	%	%	%
Lues I	8	.....	.....	.....	.....	77.5	74	90	48.2	100	68	52.6	....
Lues II with symptoms	26.7	.....	.....	.....	.....			98	79	93	93	100	92.8
without symptoms ("early latent")	14.6	.....	.....	.....	.....			80	20	64	...	75.6	46
Lues III with symptoms	.....	.....	.....	.....	.....			91	57.4	98	100	92.6	88.9
without symptoms ("late latent")	11.3	.....	.....	.....	.....	100	79	57	20.2	42	...	36.8	37.5
Pro- gressive paral- ysis	.....	(80) (spinal fluid)	(75) (spinal fluid)	(100) (spinal fluid)	.....			.....	.....	.....	.....	88	.....
Tabes dorsalis	.....	.....	.....	.....	(66) (spinal fluid)			86.6 (22) (spinal fluid)	.....	.....	.....		.....

As has been repeatedly remarked, *specificity* is the important element in every biological reaction. The reaction, known after the discoverer as the Wassermann Reaction, can also be performed if, instead of the extract from luetic organs, an alcoholic extract of certain normal organs or certain lipid substances is substituted as antigen. Seligmann was likewise able to obtain complement fixation by pure chemical reactions. Consequently, numerous authorities expressed the opinion that the Wassermann Test was non-specific and that it does not at all represent an antigen antibody interaction.



There is no doubt, however, that this exceptional view is incorrect. It is true that the real syphilitic antigen is unknown, but most probably it is neither the pure spirochaetes nor a pure lipid substance. The author has expressed the hypothesis that the antibody producing antigen is a toxolipoid. This explains the fact that pure lipoids cannot stimulate any antibodies, but can react nevertheless with luetic antibodies in vitro.

The accompanying diagram (Fig. 17) explains this hypothesis. In order to answer the objections raised against such a theory, the author has proposed the indifferent term of "Lueseargine" for the luetic antibodies, as long as their biological structure is unknown.

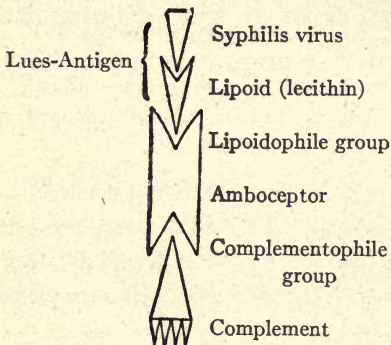


FIG. 17.

Experiments by Citron and Munk prove without any doubt that the luesreagine is a true antibody of an antigen contained only within the *aqueous syphilitic extract*. Aqueous and alcoholic extracts of normal organs do not contain this antigen. Rabbits were immunized with various antigens. Only those injected with watery syphilitic extracts developed antibodies similar to the luesreagines of human syphilis in that they reacted with alcoholic normal extracts. Blumenthal and

Meyer corroborated these findings and further showed that even the alcoholic syphilitic extracts do not contain this antigen.

Since the cultivation of the spirochetes in pure culture has been simplified by Noguchi, extracts of such cultures have been made and employed as antigens for the Wassermann reaction. It was hoped that if such a specific and efficient antigen could be obtained, the basis of a true antigen antibody reaction would be more certain. While fixation occurs with the pallida culture antigen, the results cannot be depended upon for clinical purposes; some cases of undoubted syphilis giving a strongly positive reaction with the syphilitic liver antigen, give absolutely negative results with the culture antigen (Craig and Nichols).

Independent of the question of "biological specificity," the Wassermann reaction must also be considered in the light of "clinical specificity." From this standpoint it fulfills its demands. With only few exceptions, it can be regarded as absolutely specific for lues.

The well established exceptions are, framboësis, trypanosomiasis, leprosy, malaria, scarlet fever, febris recurrens. The reactions obtained here are similar, but not the same as those obtained in syphilis. In leprosy the difference is that the reaction can also be performed with tuberculin as antigen; in scarlet fever the reaction appears only in a small percentage of cases and not with all luetic extracts. Further-

more, it disappears at the latest three months after the infection, usually much sooner. As for trypanosomiasis and malaria convincing data are still too few. In malaria the reaction is always negative if the parasites are absent.

These diseases excluded, a positive Wassermann reaction can be taken as certain proof of the existence of lues. Whether such a test is indicative of a by-gone infection or whether it means that an active process is still going on, has been for a long time a subject of discussion. The author is of the firm opinion *that the demonstration of the "lues reagine" means active lues*. The reasons for this belief are as follows:

1. The almost constant presence of the reaction in all cases of manifest lues excepting primary lesions. During this initial stage it is entirely absent or only faintly positive. It appears, however, later on.

2. The practically assured existence of the reaction with a recurrence of symptoms even though formerly it was negative.

3. The possibility of influencing a positive reaction so that it becomes negative, by the use of mercury. The latter holds true also for those cases which show no symptoms and are therefore incorrectly designated as latent syphilis. It has been proven that such are in reality by no means latent, but have an active process at some point escaping detection, as the aorta. Only cases of a nature which have no symptoms and a negative reaction should be considered as latent syphilis; those, however, with no symptoms, but a positive reaction as belonging to the class of active lues.

4. The evidence that apparently healthy individuals, but with a positive reaction, have infected others, or have suddenly developed tertiary or postlueic manifestations—tabes, paresis, diseases of the aorta, etc.

An objection has frequently been raised, that in spite of existing disease, the reaction has been found negative. If the statistics covering the largest number of cases are studied, it will be seen that such instances are rare. Exceptions are discovered in every biological reaction, especially one which is complicated, and where five different agents come into play; even in the immunization of animals differences will be found in that some produce a highly agglutinating or precipitating, etc., serum, while others will show few or even no antibodies. Individual differences are prevalent to such an extent that exceptions to the rule must be taken for granted. Fortunately, a negative reaction in existing lues is so rare, that for practical purposes its possibility may be overlooked, at least, with reservation.

As a general rule, antibodies persist in an organism for a certain time after infection, when the individual has become perfectly well. Discussion, to the effect that it may be possible for a positive Wassermann reaction to similarly signify a past infection or a state of immunity, has been raised. But it must be said that immunity in syphilis is a condition thus far unproven, and almost unknown. All symptoms previously attributed to such an immunity can more easily be explained in the light of a continuation of the disease. As for the "lues reagine" remaining after the cure of the infection, this phenomenon is undoubtedly possible. The analogy with other diseases seems lost, however, when one considers that the syphilitic reaction is discovered thirty or



forty years after an infection, while antibodies in general persist for weeks, months or at the most for several years, following an infection. Still it may be possible that the syphilis "reagine" is characterized by the difficulty with which it is excreted and by the tendency of the cells when once stimulated to produce antibodies to continue to do so. The influence of mercury, however, demonstrates that this phenomenon is closely allied to similar actions exhibited by the class of bacteria. If a patient whose serum gives a positive reaction is subjected to mercurial treatment, the reaction becomes negative. The mercury has destroyed the stimulant or irritant which has led the cells to the production of antibodies. If this stimulant is excluded, the "lues reagine" disappears from the blood just as bacterial antibodies disappear after the bacteria have been eradicated. Thus there is no basis for attributing to the luetic antibodies any exceptional properties.

The fact, that mercury leads to an alteration in the reaction, Citron's prompted the author to employ the *Wassermann test* as a "Biological guide to the biological mercurial treatment. The aim was not Mercurial Treatment." *only to cause a disappearance of all manifestations, but to obtain a negative reaction.* It soon appeared that a negative reaction once obtained did not necessarily remain so. As soon as a recurrence set in the reaction became positive again; in fact, the reaction also reappeared without a return of symptoms. In the latter case such a return alone was regarded as a fresh manifestation of a reactivation process and an indication for treatment. It became advisable, therefore, to repeat the test at definite intervals and depend upon the return of the reaction for further treatment. This basis of therapy, which at first met with marked opposition, has recently won many followers.

The experiments of Boas in Copenhagen are especially instructive from this point of view.

He examined eighty-two patients with secondary syphilis before and after mercurial therapy. All gave positive reactions before the treatment; after it, seventy-six gave no reaction, six retained the positive reactions; one of the six did not return for observation. Of the remaining five, all had a return of symptoms within one month after cessation of the mercury, while of the seventy-six only three returned with a recurrence. Boas next made observations of sixty-five patients who were in the first three years of their infection, but who gave a negative Wassermann after the treatment. In sixty-two cases, a positive reaction reappeared after one to two months, eight of these having at the same time a recurrence of symptoms; of the remaining fifty-four, nineteen were not treated. They all showed a return of symptoms, but only one and a half months after the appearance of the positive Wassermann. Thus if the scheme of the chronic intermittent mercurial therapy of Neisser and Fournier were followed, these patients would begin to get treatment one and a half months after the active lues had again started, as shown by the positive Wassermann reaction. Of the remaining thirty-five cases all began treatment when the Wassermann test became positive. None of these had any return of symptoms during the following period of observation (three to five months).

The experiments of Boas show distinctly the advantages of the mercurial therapy when based upon the biological reaction instead of upon the

schematic, symptomatic, chronic, intermittent treatment of Fournier and Neisser.

At the present day, when the spirochætes can be so readily found in the primary lesion of syphilis, the biological mercurial treatment should be undertaken in the earliest stage. It is possible even to begin at a time when the serum reaction is still negative, but after the spirochætes have been demonstrated. The most ideal cases are those in which treatment is instituted so early that they never develop a positive Wassermann test.

Naturally the statement made that mercurial treatment should be continued until the reaction becomes negative may be limited by certain contra-indications which may arise in the general condition of the patient. This must always be considered. Especial difficulty to attain a negative reaction is encountered in those cases in which lues has persisted for many years.

It must be kept in mind that the luetic infection does not always present the typical clinical picture ascribed to it in the text-books. The "Lues asymptomatica," that is, the lues apparently presenting no symptoms, is by no means rare. *To-day one must not wait until the syphilitic patient comes to the physician, but it is the duty of the latter to look for the evidence of syphilis among those related to or associated with infected persons.* If one proceeds in such a systematic method it will be found that the mothers of syphilitic children, so frequently regarded as immune, are in reality not so. In such cases, without any clinical evidence of syphilis, the Wassermann reaction is positive in about 56 to 75 per cent.

This question becomes of utmost importance in the prevention of lues. For example the obligatory examination of the serum of wet nurses has shown that of all such applicants at the Dresden Infant Asylum 10 per cent. gave a positive reaction (Rietschels). On further study it was ascertained that 75 per cent. of the children of these apparently healthy women gave luetic manifestations immediately or shortly after birth.

The Wassermann Test also offers a certain guide as to the prognosis of a case. Thus the outlook is unfavorable if in spite of energetic treatment a reaction remains constantly positive. Even with the absence of all external lesions such a condition can be classified as a "lues maligna." As a rule this term is applied to external obvious syphilitic manifestations that remain entirely uninfluenced by mercurial therapy. When it is considered that all general paralysis patients react very strongly positive, the importance of bringing about a negative reaction is sufficiently impressed. Salvarsan is of great help toward this aim.

A rapid disappearance of a positive Wassermann reaction after specific therapy offers a good prognosis. This is the more favorable the longer the negative test persists. Its continuation over several years with no clinical manifestations can be accepted as a cure of the syphilitic infection.



Serum Diag- In close association with the serum diagnosis of syphilis, com-  
nosis of Dis- plement fixation has been employed as a means for the diagno-  
eases Caused sis of conditions caused by the animal parasites and especially  
by Animal by the echinococcus. In the serum of patients suffering  
Parasites. from these infections, substances are found closely allied to  
the "lues reagine." They bind complement with an antigen consisting of  
an extract of the respective worms or hydatid fluid.

Ghedini, Weinberg and Parvu and others have found that in most cases of echino-  
coccus disease, the reaction is positive. If by operation the cyst is only incised, the re-  
action becomes stronger or in few cases appears positive for the first time. After com-  
plete excision of the cyst, the reaction disappears. According to Parvu and Laubry, a  
positive test is found in the spinal fluid only when the echinococcus cysts have invaded  
the brain.

Ghedini described similar findings, caused by the ascaris, ankylostoma,  
etc.

## CHAPTER XIV.

### THE TECHNIQUE OF COMPLEMENT FIXATION.

*Original method of Bordet-Gengou. Wassermann-Bruck's modification. Technique of serum diagnosis for syphilis. Echinococcus disease. Serum diagnosis of other diseases, epidemic meningitis, tuberculosis, gonococcus infections, typhoid fever. Differentiation of proteids according to Neisser-Sachs.*

#### I. The Original Method of Bordet-Gengou.

a. *The antigen* consists of bacteria grown upon agar for twenty-four hours and then suspended in physiological salt solution to make a rather concentrated emulsion.

For typhoid bacteria Bordet and Gengou take 5 c.cm. of salt solution to each culture of bacteria.

For tubercle bacilli 80 mg. of the bacteria are suspended in 1 c.cm. of salt solution.

b. The serum containing the *antibody* is heated for one-half hour at 56° C. to destroy the complement.

c. As complement, the fresh serum of a normal animal or human being is used.

d. *The hemolysin* consists of the inactivated serum of a rabbit that has been immunized against sheep's or goat's erythrocytes, or the serum of a guinea-pig injected with rabbit's red blood cells.

e. The respective red blood corpuscles are washed, to free them of their complement containing serum.

A definite amount of bacterial suspension is mixed with varying amounts of inactivated immune serum and a proportional amount of complement is added. These three ingredients are mixed and allowed to remain at room temperature for four to five hours. During this time the complement is fixed if the antigen and antibody are of a homologous nature. In order to see whether this union has taken place or not, hemolysin and erythrocytes are added in a mixture thus prepared: 2 c.cm. of inactivated hemolysin + twenty drops of washed blood cells are mixed and allowed to remain together for about fifteen minutes so that the erythrocytes are sensitized, *i.e.*, united with the hemolytic amboceptor. Of this mixture each tube receives 0.1 to 0.2 c.cm. If the complement has not become fixed, hemolysis occurs in several minutes. If the complement has become fixed, hemolysis does not occur; since, however, the hemolysin also contains hemagglutinin, the erythrocytes are agglutinated and sink to the bottom of the tubes:

As control tests, Bordet and Gengou considered the following very necessary:

1. Bacterial suspension + inactivated normal serum (instead of immune serum)



+ complement (five hours) + hemolysin + blood. Hemolysis must occur, as the normal serum does not contain enough amboceptors to unite with the bacterial suspension and consequently complement remains unbound.

2. Inactivated immune serum + complement (five hours) + hemolysin + blood. Hemolysis results.

3. Inactivated normal serum + complement (five hours) + hemolysin + blood. Hemolysis.

4. Antigen + inactivated immune serum + hemolysin + blood. No hemolysis, as complement is absent.

5. Antigen + inactivated normal serum (five hours) + hemolysin + blood. No hemolysis, as complement is absent.

The following is the chart of the first complement fixation test as originally performed by Bordet and Gengou in 1901 in which pest antibodies were demonstrated in the serum of an immunized horse.

	Antigen	Antibodies	Complement	Hemolysin and erythrocytes	Results
1	0.4 pest bacilli emulsion.	1.2 inactive pest serum (horse).	0.2 guinea-pig's serum.	2 drops of rabbit's blood sensitized.	0
2	0.4 pest bacilli emulsion.	1.2 inactive normal serum (horse).	0.2 guinea-pig's serum.	2 drops of rabbit's blood sensitized.	Complete hemolysis.
3	.....	1.2 inactive pest serum (horse).	0.2 guinea-pig's serum.	2 drops of rabbit's blood sensitized.	Complete hemolysis.
4	.....	1.2 inactive normal serum (horse).	0.2 guinea-pig's serum.	2 drops of rabbit's blood sensitized.	Complete hemolysis.
5	0.4 pest bacilli emulsion.	1.2 inactive pest serum (horse).	.....	2 drops of rabbit's blood sensitized.	0
6	0.4 pest bacilli emulsion.	1.2 inactive normal serum (horse).	.....	2 drops of rabbit's blood sensitized.	0

Employing this method, Bordet and Gengou found positive results with the following combinations:

1. Pest bacilli + pest horse's serum + guinea-pig complement + guinea-pig hemolysin + rabbit's blood.

2. Anthrax vaccine + guinea-pig immune serum + guinea-pig complement + guinea-pig hemolysin + rabbit's blood.

3. Typhoid bacilli + guinea-pig immune serum + guinea-pig complement + guinea-pig hemolysin + rabbit's blood.

4. Coli bacilli + guinea-pig immune serum + guinea-pig complement + guinea-pig hemolysin + rabbit's blood.

5. Typhoid bacilli + human convalescent serum + human complement + guinea-pig hemolysin + rabbit's blood.

6. Killed tubercle bacilli + guinea-pig immune serum + guinea-pig complement + rabbit's hemolysin + goat's blood or sheep's blood.
7. Whooping cough bacilli + patient's serum + guinea-pig's complement + rabbit hemolysin + goat's or sheep's blood.
8. Meningococci + human convalescent's serum + human complement + guinea-pig's hemolysin + rabbit's blood (Cohen).

Foix and Mallein examined twelve cases of scarlet fever and obtained a positive result in ten cases when the streptococcus grown from a scarlet angina was used as antigen. Antibodies were found on the fourth day. These results were confirmed by Schleissner.

## II. Wassermann-Bruck's Modification.

a. Antigen.—Instead of entire bacteria, only bacterial extracts are employed. These are made in the same manner as the artificial aggressins.

For typhoid bacteria Leuchs advises that the bacterial suspension should first be killed for twenty-four hours at 60° C. and then shaken for two days. In tuberculosis good results are obtained by using Koch's preparation of old and new tuberculin.

The bacterial extracts when very fresh contain a great deal of precipitinogen which diminishes in several days and finally disappears. Its presence does not disturb complement fixation. *The bacterial extracts must be well protected from light and kept in the cold.*

After the extract has stood for some time a sediment forms; under no circumstance should this be disturbed or shaken. The required amount of antigen should be carefully poured off, and not pipetted off. Just as soon as the required amount is obtained, the extract should be returned to the ice-box.

b. The antiserum is inactivated by heating, even if the serum is old and contains very little or no complement.

Old, non-heated serum is often antihemolytic. Temperatures over 60° C. should be strictly guarded against as the amboceptors may be destroyed. Heating for a period longer than one-half hour may make a serum anticomplementary, *i.e.*, bind complement. Sera containing bile at times prevent hemolysis. Chylous sera obtained during the period of digestion and milky sera seen in nursing women usually do not interfere with the complement fixation reaction.

Exudates, transudates, and spinal fluids are treated like sera. Exudates very rich in albumin frequently coagulate during inactivation. In order to avoid this, it is advisable to dilute such fluids with physiological salt solution. Occasionally exudates tend to fix complements spontaneously.



c. Complement is obtained by killing a guinea-pig and using its serum while fresh. The serum preserved in "Frigo" is, according to Sterns, not reliable.

d. Hemolysin is represented by the inactivated serum of a rabbit that has been immunized against sheep's red blood cells.

e. The twice washed sheep's red blood cells are used as erythrocytes.

These five substances are placed in the test-tubes in the following order: antigen, inactivated antiserum, complement; they are thoroughly mixed by shaking and placed in the incubator for one hour in order to hasten their union. After this interval the inactivated hemolysin and the red blood cells are added as indicator. The mixtures are again returned to the incubator to promote hemolysis. As in all biological reactions, the quantitative relationship of these various ingredients determines to a great extent the final result of the complement fixation test. As far as antigen and antibody are concerned, the experiments of Weil and Nakayama must be considered; these are to the effect that only one-half of the maximum dose of each ingredient which does not bind complement is employed. With this point in view, preliminary tests determining the proper dosage of each must be performed.

The amount of complement used is always constant. In Wassermann's laboratory 1 c.cm. of the dilution 1 : 10 represents the quantity chosen. For most tests this quantity is sufficient as it represents about three times the titer of normal guinea-pig's serum. In certain instances it is preferable to work with smaller quantities, as in Marmorek's method of complement fixation with the urine of tuberculous patients. Of the hemolysin the two-fold or three-fold titer dose is taken and of the erythrocytes 1 c.cm. of a 5 per cent. suspension in normal saline solution suffices. For Marmorek's test the hemolysin is employed in just the titer dose, and of the red blood cells only 0.3 c.cm. is taken. Each of the five elements is diluted with saline to make up 1 c.cm. so that at the completion of the test all the tubes contain 5 c.cm. Quite a difference arises if an individual test is performed with a constant quantity of serum and diminishing doses of bacterial extract or reversely. Important tests should be carried out by both methods. The necessary controls are:

1. The double dose of antigen + complement + hemolysin + blood, to prove that the dose of antigen employed in the test is correct (Weil and Nakayama).

2. The double quantity of serum + complement + hemolysin + blood, to prove that the dose of serum employed is correct (Weil and Nakayama).

3. The "system control"; blood + complement + one-half amount of hemolysin, to show that the test was performed with double the hemolytic dose.

4. Blood + salt solution, to prove that the salt solution is isotonic.

In addition, it is advisable to repeat the test with inactivated normal serum substituted for the immune serum and another with a foreign instead of a homologous antigen.

These controls assure beyond doubt the specificity of the reaction.

The accompanying chart represents schematically all that has been discussed.

*Titration of a Meningococcus Serum Obtained from the Horse.**a. Diminishing Quantities of Antigen.*

Antigen	Antibodies	Complement	Hemolysin	Erythrocytes	Results
0.25 c.cm. Meningococcus extract	0.1 c.c. inactive immune serum	0.1 c.cm. guinea-pig's serum	0.002 c.cm. in- active rabbit's (sheep) serum	1 c.cm. 5 % sheep's blood	0 hemolysis
0.1 c.cm.	"	"	"	"	0 hemolysis
0.05 c.cm.	"	"	"	"	0 hemolysis
0.01 c.cm.	"	"	"	"	0 hemolysis
0.005 c.cm.	"	"	"	"	Incomplete hemolysis
0.001 c.cm.	"	"	"	"	Incomplete hemolysis
0.0005 c.cm.	"	"	"	"	Complete hemolysis
1.0 c.cm.	.....	"	"	"	Incomplete hemolysis
0.5 c.cm.	.....	"	"	"	Complete hemolysis
0.25 c.cm.	.....	"	"	"	Complete hemolysis

Antigen	Antibodies	Complement	Hemolysin	Erythrocytes	Result
.....	Inactive immune serum.				
.....	0.4 c.cm.	"	"	"	Incomplete hemolysis
.....	0.2 c.cm.	"	"	"	Complete hemolysis
.....	0.1 c.cm.	"	"	"	Complete hemolysis
.....	.....	"	0.001 c.cm.	"	Complete hemolysis
.....	.....	.....	.....	"	0 hemolysis
Meningococcus extract.	Inactive normal horse's serum.	"	0.002 c.cm.	"	Almost complete hemolysis
0.25 c.cm.	0.1 c.cm.	"	"	"	Complete hemolysis
0.1 c.cm.	0.1 c.cm.	"	"	"	Complete hemolysis
.....	0.2 c.cm.	"	"	"	Complete hemolysis
Staphylococcus extract	Inactive menin- gococcus serum	"	"	"	Complete hemolysis
0.25 c.cm.	0.1 c.cm.	"	"	"	Complete hemolysis
0.1 c.cm.	0.1 c.cm.	"	"	"	Complete hemolysis
0.5 c.cm.	.....	"	"	"	Complete hemolysis

The titer of the meningococcus serum is 0.001 c.cm. of antigen. Since 1.0 c.cm. of antigen binds 0.1 complement and 0.5 c.cm. does not interfere with hemolysis, the maximum dose of antigen which may be used for the trial is 0.25 c.cm.

Inasmuch as 0.4 c.cm. of the inactivated serum binds a part of the complement, and 0.2 does not at all interfere with hemolysis, the maximum dose of serum to be employed is 0.1 c.cm.

The positive reaction must be attributed to the interaction between antigen and antibody, as hardly any complement fixation takes place by using inactivated normal serum with 0.25 c.cm. of antigen. That the reaction is specific is shown by hemolysis occurring when the homologous antigen is substituted by a staphylococcus extract.



*b. Same with Diminishing Amounts of Serum.*

Antigen	Antibodies	Complement	Hemolysin	Erythrocytes	Result
0.25 c.cm. meningococcus extract.	0.1 c.cm. inactive meningococcus serum.	0.1 c.cm. guinea-pig's serum.	0.002 c.cm.	1 c.cm. 5% sheep's blood.	0 hemolysis
"	0.05 c.cm.	"	0.002 c.cm.	"	0 hemolysis
"	0.01 c.cm.	"	0.002 c.cm.	"	0 hemolysis
"	0.005 c.cm.	"	0.002 c.cm.	"	0 hemolysis
"	0.001 c.cm.	"	0.002 c.cm.	"	Almost 0 hemolysis
"	0.0005 c.cm.	"	0.002 c.cm.	"	Incomplete hemolysis
"	0.0001 c.cm.	"	0.002 c.cm.	"	Complete hemolysis
0.5 c.cm. meningococcus extract.	.....	"	0.002 c.cm.	"	Complete hemolysis

Antigen	Antibodies	Complement	Hemolysin	Erythrocytes	Results
.....	0.2 c.cm.	"	0.002 c.cm.	"	Complete hemolysis
.....	.....	"	0.001 c.cm.	"	Complete hemolysis
.....	.....	.....	.....	"	0 hemolysis
0.25 c.cm. meningococcus extract.	Inactive normal horse's serum.	"	0.002 c.cm.	"	Almost complete hemolysis
"	0.1 c.cm.	"	0.002 c.cm.	"	Complete hemolysis
"	0.05 c.cm.	"	0.002 c.cm.	"	Complete hemolysis
.....	0.2 c.cm.	"	0.002 c.cm.	"	Complete hemolysis
0.25 c.cm. staphylococcus extract.	0.1 c.c. inactive meningococcus serum.	"	0.002 c.cm.	"	Complete hemolysis
0.5 c.cm. staphylococcus extract.	.....	"	0.002 c.cm.	"	Complete hemolysis

With 0.25 c.cm. of antigen the titer of this meningococcus serum is 0.0005 c.cm. of serum. One could with this constant quantity of serum, and varying quantities of antigen, titrate the minimum amount of antigen necessary for complement fixation. It would even be preferable for such a test to employ 0.005 c.cm. of the serum, as this amount surely binds no complement. If such a titration is undertaken it will be found that 0.005 c.cm. of serum with 0.05 c.cm. of extract can bind 0.1 c.cm. of complement.

Similarly the antibodies contained in the blood serum or spinal fluid of a patient can be determined by means of complement fixation.

If it is desired to demonstrate the antigen instead of antibody, one proceeds as follows:

*c. Demonstration of meningococcus antigen in the spinal fluid of a patient with a possible meningitis.*

Antigen	Antibodies	Complement	Hemolysin	Blood	Results
0.5 c.cm. active spinal fluid from patient.	0.1 c.cm. inactive horse's meningococcus serum.	0.1 c.cm.	0.002 c.cm.	1 c.cm. 5%	0 hemolysis
0.3 c.cm.	0.1 c.cm.	0.1 c.cm.	0.002 c.cm.	1 c.cm. 5%	Incomplete hemolysis
0.1 c.cm.	0.1 c.cm.	0.1 c.cm.	0.002 c.cm.	1 c.cm. 5%	Complete hemolysis
.....	0.2 c.cm.	0.1 c.cm.	0.002 c.cm.	1 c.cm. 5%	Complete hemolysis
1.0 c.cm.	.....	0.1 c.cm.	0.002 c.cm.	1 c.cm. 5%	Almost complete hemolysis
0.6 c.cm.	.....	0.1 c.cm.	0.002 c.cm.	1 c.cm. 5%	Complete hemolysis
.....	.....	0.1 c.cm.	0.001 c.cm.	1 c.cm. 5%	Complete hemolysis
.....	.....	.....	.....	1 c.cm. 5%	0 hemolysis
0.5 c.cm. active normal spinal fluid.	0.1 c.cm.	0.1 c.cm.	0.002 c.cm.	1 c.cm. 5%	Complete hemolysis
1.0 c.cm.	.....	0.1 c.cm.	0.002 c.cm.	1 c.cm. 5%	Complete hemolysis
0.5 c.cm. of active spinal fluid from patient.	0.1 c.cm. inactive normal horse's serum.	0.1 c.cm.	0.002 c.cm.	1 c.cm. 5%	Almost complete hemolysis
0.3 c.cm.	"	0.1 c.cm.	0.002 c.cm.	1 c.cm. 5%	Complete hemolysis
0.05 c.cm. meningococcus extract.	0.005 c.cm. inactive meningococcus serum.	0.1 c.cm.	0.002 c.cm.	1 c.cm. 5%	0 hemolysis Look at previous test

The spinal fluid contains meningococcus antigen thus proving that the patient is suffering from epidemic cerebrospinal meningitis. Neither the double amount of serum, a double amount of antigen, a mixture of normal spinal fluid with specific serum, nor normal serum with the specific spinal fluid binds complement. Only a mixture of meningococcus extract and specific serum gives complement fixation.

The results obtained by complement fixation depend to a great extent upon the quantitative relationship of the various ingredients. The affinity toward the complement existing between the antigen and amboceptor on the one hand, is balanced by that between the hemolysin + blood on the other. By modifying their quantitative proportions different results may be obtained. If for example the strength of the hemolysin is excessively increased, it is possible that the previously bound complement is again detached and hemolysis ensues. Originally the results were read after the mixtures had remained two hours in the incubator and twenty-four hours in the ice-box. At the present, most authorities agree to read the results at a time when the control tubes are ready; that is when the complement is bound or hemolysis has been completed in those tubes in which these respective phenomena should occur.



Wassermann's modification of the Bordet-Gengou method was first practically employed for the titration of the therapeutic meningococcus serum. One cannot, however, correctly judge the prophylactic or curative value of a serum by its antibody content as they do not run hand in hand (R. Kraus, F. Meyer and Garbat, Citron.)

### III. Serum Diagnosis of Syphilis.

#### *a. Wassermann's Technique.*

The technique of this reaction as carried out in Wassermann's laboratory is practically identical with that just described for the diagnosis of bacterial infections. The preparation of the antigen varies slightly.

The liver obtained from a syphilitic fetus is weighed and cut up into fine pieces. Four times its weight of 1/2 per cent. of carbolic solution in saline is added, the mixture placed in a brown bottle and shaken for twenty-four hours. It is then centrifugalized until the larger liver remnants settle to the bottom and a somewhat turbid fluid remains above. The latter is poured off into a brown bottle and placed in the ice-box. After several days of sedimentation, the fluid assumes a yellowish-brown opalescence and can now be used as a luetic antigen. It should not be exposed to light or heat, should not be shaken, and its contents should not be pipetted off, but carefully poured off without disturbance to the sediment.

By titration of the extract, that dose is determined which does not of itself bind complement. Only such extracts are kept which in the dose of 0.4 c.cm. do not interfere with hemolysis.

Control tests should also be made to ascertain whether the organ extract has any tendency of its own to hemolyze red blood cells without the presence of complement or hemolysin.

Not every luetic extract can serve as antigen for complement fixation. A number of other substances, both normal and pathological, may be extracted from the luetic liver besides that agent necessary for the Wassermann test. These undesired ingredients may interfere with the efficiency of the extract. For this reason a great number of known positive and negative sera should be tested with each new extract, and only if the results are absolutely correct should it be employed as antigen.

In the early work of Wassermann the antigen was described as deteriorating very easily; its activity would either be entirely destroyed or it would become anticomplementary. The author is firmly convinced that these changes are brought about by careless handling of the extract or its exposure to light. If properly taken care of, its activity remains constant.

From practical experience, it has been found that extracts which must be used in amounts less than 0.1 c.cm. are as a general rule unsatisfactory.

Similarly, the luetic sera are most active when doses of 0.2 and 0.1 c.cm. are employed. Amounts greater than 0.2 may result in an unspecific reaction. The most favorable combinations are,

0.2 c.cm. of extract + 0.2 c.cm. serum.

0.1 c.cm. of extract + 0.1 c.cm. serum.

The accompanying table presents the titration of an antigen in detail.

*a. Preliminary Test—Titration of the Antigen.*

Antigen	Complement	Hemolysin	Blood	Result
0.8 c.cm. luetic extract	0.1	Twice the hemolytic dose.	1 c.cm. 5%	Incomplete hemolysis.
0.6 c.cm. luetic extract	0.1	Twice the hemolytic dose.	1 c.cm. 5%	o
0.4 c.cm. luetic extract	0.1	Twice the hemolytic dose.	1 c.cm. 5%	Complete hemolysis.
0.2 c.cm. luetic extract	0.1	Twice the hemolytic dose.	1 c.cm. 5%	Complete hemolysis.
0.8 c.cm. luetic extract	.....	.....	1 c.cm. 5%	Incomplete.
0.6 c.cm. luetic extract	.....	.....	1 c.cm. 5%	o
0.4 c.cm. luetic extract	.....	.....	1 c.cm. 5%	o
0.2 c.cm. luetic extract	.....	.....	1 c.cm. 5%	o

The test proves that 0.4 c.cm. of extract is not able to bind 0.1 c.cm. of complement. That 0.8 c.cm. of lues extract causes only an incomplete hemolysis, while 0.6 c.cm. produces no hemolysis whatever, is explained not by its lessened tendency of binding complement, but by the greater amount of hemotoxin which 0.8 c.cm. possesses.

Keeping in mind the rule of Weil and Nakayama about the summation of antigen, this particular antigen will be employed in the quantities of 0.2 c.cm. and 0.1 c.cm. (0.2 being  $\frac{1}{2}$  of 0.4 c.cm. which is the largest amount that does not of itself fix complement).

*b. Examination and titration of 4 luetic sera by Citron's method (see Plate II).*

The technical details of the test are as follows:

Three test-tubes are assigned for each test and placed in a test-tube rack. The name of the patient is written upon the first of these tubes. Another rack contains one tube for each patient and labelled accordingly. In addition there is an "antigen tube," which is placed in the first rack at the end of all the other tubes; also a "normal extract," a "system," "complement," and "blood control tube," which are placed in the second rack. The amount of syphilitic antigen required for the entire work is calculated as follows. For each test 0.3 of antigen is required; for five cases (including controls) 1.5 c.cm. are needed + 0.4 c.cm. for the antigen control tube = 1.9 c.cm. in all, or 2.0 c.cm. in round numbers. This amount is diluted with normal salt solution in the proportion of 1 : 5 so that 8 c.cm. of saline are added (= 2 : 10); 1 c.cm. of this dilution contains 0.2 antigen and  $\frac{1}{2}$  c.cm. contains 0.1 antigen. The first tube (1, 4, 7, 10, etc., in diagram) of every test therefore receives 1 c.cm., the second



tube 1/2 c.cm., the third tube nothing, the antigen tube (tube 19) 2 c.cm. Physiological salt solution is added to make up 1 c.cm. in each tube; first tube nothing; second tube 1/2 c.cm.; third tube 1 c.cm. of saline.

The normal extract required for the tubes in the second rack is similarly estimated, 0.2 c.cm. is needed for each test, 5 (tests)  $\times$  0.2 = 1.0 + 0.4 for the antigen control tubes = 1.4 or 1.5 c.cm. in round numbers. For purposes of dilution 1 : 5, 6 c.cm. of salt solution are added and 1 c.cm. (= 0.2 of extract) placed into each of the tubes (20 to 26) and 2 c.cm. into the normal extract control test-tube (tube 27). In this series also, salt solution is added to make up equal quantities of 1 c.cm.: first tube nothing, second tube 1/2 c.cm., third tube 1 c.cm.; antigen tube (27), nothing; system (28), complement (29) and blood (30) control tubes each 1 c.cm.

The second ingredient of the test is next added, *i.e.*, the respective serum. This is not diluted but added directly; 0.2 c.cm. into the first tube of each test; 0.1 c.cm. into the second; 0.2 c.cm. into the third tube; also 0.2 into the control series of tubes labelled with the patient's names in the second rack. Salt solution is again added to make up to the equal quantity of 2 c.cm. in each tube, thus: 0.8 c.cm. into first, 0.9 c.cm. into the second, 0.8 c.cm. into the third tube, and 0.8 c.cm. into the control series; nothing into the antigen tubes, 1 c.cm. into system, complement, and blood control tubes.

The addition of complement follows next. Each tube, except the blood tube, receives 0.1 c.cm. of complement. Thus the tubes are counted and if, for example, nineteen tubes are present  $19 \times 0.1$  c.cm. complement is taken, or in round numbers 2 c.cm.

Complement is always diluted 1 : 10, or 2 c.cm. complement + 18 c.cm. saline, so that each tube except the blood tube (30) receives 1 c.cm. of this diluted complement. Tube (30) receives 1 c.cm. of saline instead. All tubes are then carefully shaken and the racks placed in the incubator for one hour.

During this time, the hemolysin and washed red blood cells are properly diluted. The red blood cells are made up in a 5 per cent. suspension of which each tube will receive 1 c.cm. Thus in the present test there are thirty tubes, requiring 30 c.cm. of blood suspension; since 1 c.cm. of washed blood when diluted 1 : 20 will supply twenty tubes, for 30 c.cm. about 1 1/2 c.cm. of blood will be required, or 2 c.cm. will make 40 c.cm. of a 5 per cent. blood suspension.

Luetic extract	Serum.	Comple- ment	Hemoly- sin; 1c. cm.	Sheep's blood	Result of hemolysis	
1. 0.2	0.2 Ser. I. Tubes un- treated; without luetic history.	0.1	1 : 1000	1 c.cm. 5%	No hemol- ysis.	} + + + +
2. 0.1	0.1 As above.	0.1	1 : 1000	1 c.cm. 5%	No hemol- ysis.	
3. ...	0.2 <sup>1</sup> As above.	0.1	1 : 1000	1 c.cm. 5%	Complete hemolysis.	
4. 0.2	0.2 Ser II. Secondary lues untreated.	0.1	1 : 1000	1 c.cm. 5%	No hemol- ysis.	} + + +
5. 0.1	0.1 As above.	0.1	1 : 1000	1 c.cm. 5%	Incomplete hemolysis.	
6. ...	0.2 <sup>1</sup> As above.	0.1	1 : 1000	1 c.cm. 5%	Complete hemolysis.	

Luetic extract	Serum	Complement	Hemolysin; 1c.cm.	Sheep's blood.	Result of hemolysis
7. 0.2	0.2 Ser. III Tabes. Many inunction courses	0.1	1 : 1000	1 c.cm. 5%	No hemolysis.
8. 0.1	0.1 As above.	0.1	1 : 1000	1 c.cm. 5%	Complete hemolysis.
9. ...	0.2 As above.	0.1	1 : 1000	1 c.cm. 5%	Complete hemolysis.
					} ++
10. 0.2	0.2 Ser. IV Gallstones. Lues seventeen years ago. Much treatment. No symptoms for ten years.	0.1	1 : 1000	1 c.cm. 5%	Trace of binding; almost but not quite complete hemolysis.
11. 0.1	0.1 As above.	0.1	1 : 1000	1 c.cm. 5%	Complete hemolysis.
12. ...	0.2 <sup>1</sup> As above.	0.1	1 : 1000	1 c.cm. 5%	Complete hemolysis.
					} ±
13. 0.2	0.2 Negative control serum (carcinoma hepatis).	0.1	1 : 1000	1 c.cm. 5%	Complete hemolysis.
14. ...	0.2 <sup>1</sup> As above.	0.1	1 : 1000	1 c.cm. 5%	Complete hemolysis.
					} -
15. 0.1	0.1 Strongly positive control serum. (Lues maligna).	0.1	1 : 1000	1 c.cm. 5%	No hemolysis.
16. ...	0.2 As above.	0.1	1 : 1000	1 c.cm. 5%	Complete hemolysis.
					} +++++
17. 0.2	0.2 Weakly positive control serum. Primary lesion.	0.1	1 : 1000	1 c.cm. 5%	Incomplete hemolysis.
18. ...	0.2 <sup>1</sup> Weakly positive control serum. Primary lesion.	0.1	1 : 1000	1 c.cm. 5%	Complete hemolysis.
					} +
19. 0.4	.....	0.1	1 : 1000	1 c.cm. 5%	Complete hemolysis.
Normal extract.					
20. 0.2	0.2 Serum I.	0.1	1 : 1000	1 c.cm. 5%	Complete hemolysis.
21. 0.2	0.2 Serum II.	0.1	1 : 1000	1 c.cm. 5%	Incomplete hemolysis.
22. 0.2	0.2 Serum III.	0.1	1 : 1000	1 c.cm. 5%	Complete hemolysis.
23. 0.2	0.2 Serum IV.	0.1	1 : 1000	1 c.cm. 5%	Complete hemolysis.
24. 0.2	0.2 Negative control serum.	0.1	1 : 1000	1 c.cm. 5%	Complete hemolysis.
25. 0.2	0.2 Strongly positive control serum.	0.1	1 : 1000	1 c.cm. 5%	Complete hemolysis.
26. 0.2	0.2 Weakly positive control serum.	0.1	1 : 1000	1 c.cm. 5%	Complete hemolysis.
27. 0.4	.....	0.1	1 : 1000	1 c.cm. 5%	Complete hemolysis.
28. ...	.....	0.1	1 : 1000	1 c.cm. 5%	Complete hemolysis.
29. ...	.....	0.1	.....	1 c.cm. 5%	No hemolysis.
30. ...	.....	.....	.....	1 c.cm. 5%	No hemolysis.



The hemolysin is prepared as follows: its titer for example is 1 : 2000 and it is employed in the dilution of 1 : 1000. Each tube except the blood and complement tubes will receive 1 c.cm. of the diluted hemolysin; 1 c.cm. of the latter if diluted properly would give 1000 c.cm.; 0.1 of the hemolysin which is the smallest amount that can be measured will give 100 c.cm. Every tube except 28 to 30 will receive 1 c.cm. of the hemolysin dilution 1 : 1000. Tubes 29 and 30 will receive none (replaced by saline), tube 28 will receive 1/2 c.cm. of the hemolysin and 1/2 c.cm. of saline.

After an hour's incubation, each tube receives 1 c.cm. of R. B. C. and 1 c.cm. of the hemolysin just mentioned. If it is desired to hasten the results, it is advisable to mix a sufficient equal quantity of R. B. C. and hemolysin solution (30 c.cm. of each) and allow the mixture to remain in the incubator for a short time before the hour's incubation is up. Then instead of adding 1 c.cm. of these ingredients separately, 2 c.cm. of the mixture is added to all except tubes 28 to 30. Tubes 29 and 30 receive 1 c.cm. of blood and 1 c.cm. of saline and tube 28 1 c.cm. of blood, 1/2 c.cm. of hemolysin and 1/2 c.cm. of saline.

#### *Meier's Modification.*

For over a year G. Meier in Wassermann's laboratory has been using a *turbid* suspension of the syphilitic liver instead of the clear antigen as prepared above. After the liver is cut up into fine pieces, placed into four times its weight of one-half per cent. carbolic acid solution in normal saline and shaken for 24 hours, the mixture is filtered

#### I (Qualitative Test)

		A known positive serum	A known negative serum	Salt solution.
Extract X.....	0.2	Complete fixation.	Few non-hemolyzed red cells.	Few non-hemolyzed red cells.
Extract X.....	0.1	Complete fixation.	Hemolysis.	Hemolysis.
Extract X.....	0.05	Complete fixation.	Hemolysis.	Hemolysis.
Extract X.....	0.025	Complete fixation.	Hemolysis.	Hemolysis.
Extract X.....	0.0125	Incomplete hemolysis.	Hemolysis.	Hemolysis.
Known syphilitic extract.....	0.035	Complete fixation.	Hemolysis.	(0.07) Hemolysis.
Salt solution with double the quantity of serum.	.....	Hemolysis.	Hemolysis.	.....

<sup>1</sup> 0.4 c.cm. of luetic serum frequently binds complement of its own accord. Experience has shown that if 0.2 c.cm. does not bind complement and 0.2 c.cm. of serum + 0.2 c.cm. of antigen does bind complement, the unknown serum is surely of luetic origin.

through gauze to remove the very large particles. The filtrate is then ready for use. It is light or dark brown in color, and translucent even in weak dilution. On standing, a brown sediment settles to the bottom; before being used the extract should be thoroughly mixed as in this way it is five to ten times stronger than if the supernatant clear fluid alone is taken.

The efficiency of the extract is proven in two ways: 1. The orientation or qualitative test, to determine whether the new extract is specific for lues. 2. The quantitative test, to determine the exact dosage necessary for complement fixation. (See Table I.)

Thus it is observed that for both of these standard sera (positive and negative) this extract can be employed in two doses 0.05, 0.025.

The next step is to determine which of these two doses is the more efficient for the greatest number of sera. The largest quantity of antigen is always preferred; but it is a known fact that the optimum for one serum is not equally that for another serum. Accordingly, parallel examinations are made of a great number of sera, employing both the new antigen and the old control one, which has already been tested out and

## II (Quantitative Test)

	Rack I with standard extract	Rack II with new extract, dose 0.025	Rack III. Salt solution with double the quantity of serum
1. Serum A. ....	Complete hemolysis.	Complete hemolysis.	Complete hemolysis.
2. Serum B. ....	No hemolysis.	No hemolysis.	Complete hemolysis.
3. Serum C. ....	Incomplete hemolysis.	Complete hemolysis.	Complete hemolysis.
4. Serum D. ....	Complete hemolysis.	Complete hemolysis.	Complete hemolysis.
5. Serum E. ....	No hemolysis.	Moderate hemolysis.	Complete hemolysis.
6. Serum F. ....	No hemolysis.	Good deal of hemolysis.	Complete hemolysis.
7. Serum G. ....	Almost complete hemolysis.	Complete hemolysis.	Complete hemolysis.
8. Serum H. ....	Complete hemolysis.	Complete hemolysis.	Complete hemolysis.
9. Serum I. ....	No hemolysis.	No hemolysis.	Complete hemolysis.
10. Serum K. ....	No hemolysis.	Very slight hemolysis.	Complete hemolysis.
11. Known luetic serum.	No hemolysis.	Very slight hemolysis.	Complete hemolysis.
12. Known normal serum.	Hemolysis.	Hemolysis.	Complete hemolysis.
13. Salt solution with double the quantity of extract.	Hemolysis.	Hemolysis.	Complete hemolysis.



found to give absolutely reliable results; *i.e.*, negative reactions with non-luetic bloods, and a high percentage of positive results (almost 100 per cent.) in cases with florid non-treated lues. If the new extract is found to give a negative or weakly positive test while the standard extract shows a strongly positive reaction, it is evident that the dose of the new antigen must be increased; vice versa, if the new extract gives too many positive results especially with sera of known non-luetic origin, the dose should be diminished. (See Table II.)

From this table it is readily seen that the new extract conforms fully in its results with the standard extract as far as the negative sera and the two strongly positive (2 and 9) sera are concerned. In other cases (5, 6 and 10) the new extract gives only partial inhibition of hemolysis while with the control antigen complete inhibition results. With sera 3 and 7 there is complete hemolysis instead of partial fixation. These data prove that the new antigen is not sufficiently active in the dose of 0.025; consequently another set of control reactions must be undertaken this time employing 0.05 of the antigen. By thus repeatedly altering the dose of the antigen, the results will finally tally approximately with the standard extract (rarely do they do so absolutely) and only then may the former be used for the routine examinations. Once the proper dose of this watery extract is established, it remains constant for a very long period. If the strength of the antigen changes at all, it does so, in contrast to the alcoholic extracts of normal organs, only very gradually, and may then have to be used in smaller or greater dosage as determined by renewed titration.

Several sera from cases with high temperatures or scarlet fever should be included among the tests as in these conditions the tendency toward inhibition of hemolysis in the presence of lipoids is increased.

For the sake of economy, Meier has been working with one-half quantities of all the ingredients; thus

0.5 c.cm. of the diluted antigen.....	instead of 1.0 c.cm.
0.1 c.cm. of the diluted serum.....	instead of 0.2 c.cm.
0.5 c.cm. of the diluted complement.....	instead of 1.0 c.cm.
0.5 c.cm. of the diluted hemolysin.....	instead of 1.0 c.cm.
0.5 c.cm. of the diluted red cell emulsion...	instead of 1.0 c.cm.

The hemolysin is taken in three to four times its minimum hemolytic dose or titer, as the turbid antigen per se inhibits hemolysis more strongly than the clear extracts.

In the hands of an experienced worker, Meier's method undoubtedly yields most reliable results. The author, however, sees no distinct advantage in these modifications. He therefore advises the classical method for the beginner, as it combines simplicity with correctness.

### Citron's Standard for the Strength of a Reaction.

Citron divides the positive complement fixation tests into four grades, as follows:

- |  |   |                           |
|--|---|---------------------------|
| <p>a. Tubes 1 and 2 show complete absence of hemolysis: ++++</p> <p>b. Tube 1 shows complete absence of hemolysis and 2 shows faint hemolysis: +++</p> | } | <p>Strongly positive.</p> |
|--|---|---------------------------|

- |  |   |                  |
|--|---|------------------|
| c. Tube 1 shows complete absence of hemolysis and      | } | Weakly positive. |
| 2 shows complete hemolysis: ++                         |   |                  |
| d. Tube 1 shows partial hemolysis and                  | } | Doubtful.        |
| 2 shows complete hemolysis: +                          |   |                  |
| e. Tube 1 shows doubtful binding and                   | } | Doubtful.        |
| 2 shows complete hemolysis: =                          |   |                  |
| f. Tubes 1 and 2 show complete hemolysis: —, Negative. |   |                  |

When a series of tests is to be performed, it is advisable to include in the reaction three sera previously tested, one strongly positive, another weakly positive and a third, negative, so that the new results can be more readily compared. In this way absolutely reliable and constant values will be obtained.

Every new antigen should be tested for four weeks before its practical value can be assured. During this month, all the tests should be done with both the old and new extract and only if their results are identical should the new extract be employed. The author is in the habit of mixing the new antigen with the old one after the former has proved itself efficient. Occasionally the new antigen varies in strength from the old one. In such a case, if stronger, it must be used in a smaller dose (0.18 and 0.9) or if weaker, must be used in larger dose (0.22 and 0.11). Shaking the antigen should be strictly guarded against.

In order to control the effect of normal liver substances contained in the antigen an extract is prepared from normal fetal liver (normal antigen). When sera from cases of clinically evident lues are to be examined, it is unnecessary to have control tests with the normal fetal extract as antigen. On the other hand, such control tests are absolutely necessary in important differential diagnosis as between lues and carcinoma, and in all diseases of the nervous system. Here a positive reaction can only then be taken as evidence of syphilis if the complement fixation test is positive with the syphilitic antigen and negative with the normal liver antigen. If on the other hand it be positive with both extracts, it does not speak for a luetic infection.

A strongly positive Wassermann reaction indicates the presence of a luetic infection. A weakly positive result can be similarly interpreted if the serum control tube (Tube No. 3) is completely hemolyzed. If, however, the latter still shows some non-hemolyzed red blood cells, the + reaction must be considered as = or a reaction of indefinite nature. Only exceptionally are such doubtful reactions found in perfectly healthy individuals, although they are more often encountered in different infectious diseases (typhoid, measles, scarlet fever) and tumors. A positive diagnosis of lues should never be made upon a = reaction. On the other hand if there is a history of lues, or clinical evidences of its existence, a = reaction is to be interpreted as + and should warrant further specific therapy. As an end result of specific therapy a = reaction is not sufficient.



Not before an absolutely negative reaction has been attained should specific therapy cease.

Several authorities consider only such tests as positive where there is complete absence of hemolysis. This principle is proven as incorrect by their own statistics; a great number of their surely syphilitic cases give a negative reaction.

If the third tube (serum control) does not hemolyze, the test can be considered neither positive nor negative. Very frequently the third tube of very strongly positive cases will hemolyze very much more slowly than negative cases; these tests must therefore remain in the incubator for a longer period than the negative or weakly positive ones and until the serum tube is completely hemolyzed.

### *Other Modifications of Wassermann's Technique.*

On account of the somewhat complex technique of the reactions numerous attempts have been made to simplify the test in one way or another. The greatest difficulty lay in the preparation of a suitable antigen. From the sundry modifications and improvements made in this respect, perhaps the most important was announced simultaneously by Landsteiner, Müller and Pötzl, and Porges and Meier.

They showed that by alcoholic extraction of luetic and even normal organs of human beings and lower animals, substances were obtained which could be used as a substitute for the aqueous syphilitic antigen. The belief therefore arose that the active agents in the luetic extract belong to the class of lipoids, and Porges and Meier endeavored to isolate them from the serum. Thereupon it became evident that lecithin could replace the antigen, but only up to a certain point. Further study by H. Sachs led to the adoption of entire formulæ for artificial antigens.

The new principle disclosed by these discoveries led to many modifications in the preparation of the antigen, the main advantage of which consisted in bringing the reaction into more general use and application. The previous necessity of making an extract from the liver of a luetic fetus somewhat limited this. The Wassermann reaction became in a short period of time much more popular, *although one could not adhere to it with the same idea of specificity as before.*

Other changes in the reaction referred to the serum for examination. H. Sachs demonstrated that the inactivation at 56° C. destroyed a great part of the luetic "reagine." The dispensation of the latter was therefore recommended. It soon became evident, however, that by so doing a great number of normal and non-luetic pathological sera especially from carcinoma cases gave a positive reaction. *It is best therefore that this modification should by all means be discarded.*

As all fresh sera contain complement, the addition of guinea-pig's complement seemed superfluous if the serum for examination is employed in an active form. The following combination was therefore proposed:



1. Luetic extract or one of its substitutes.
2. Active luetic serum (contains luetic "reagine" + complement). One hour in incubator.
3. Inactive hemolysin.
4. Red blood cells.

In view of the above-mentioned objections, especially the too frequent positive results, this modification although advised by various authorities, Stern and others, should not be employed.

Not only the addition of complement, but also of immune hemolysin can be discarded, because every serum normally contains hemolytic antibodies for foreign species of blood. The contraindication for the transfusion of foreign blood depends upon this principle.

Accordingly, some authors advise the following schemes:

- |   |  |
|---|--|
| 1. Luetic extract or its substitute.                  | 1. Luetic extract or its substitute.   |
| 2. Inactive luetic serum ("luesreagine" + hemolysin). | 2. Active luetic serum (luesreagine + complement + normal hemolysin), one hour in incubator. |
| 3. Complement. One hour in incubator.                 | 3. Washed erythrocytes of sheep.   |
| 4. Washed erythrocytes of sheep.                      |  |

The advantage of these modifications is supposed to exist in the omission of the immune hemolysin. The preparation and preservation of this ingredient is, however, technically so simple that this advantage is only theoretical. Bauer believes that this change is preferable to the classical method for the reason that with the latter, the varying amount of normal hemolysin is always added to the constant amount of immune hemolysin, thereby resulting in a different quantity of the hemolysin in each test. Experience has, however, shown that the faint trace of normal hemolysin never influences the result of the test. At times so little normal hemolysin will exist in a patient's serum that it becomes necessary to add some serum of another normal patient. Such manipulations lead to new difficulties so that taken all in all, this innovation offers no advantages and should therefore not be accepted.

Brieger and Renz have recently advised the substitution of potassium chlorate for the immune hemolysin. Had this been correct the biological bases of the Wassermann reaction would have been undermined. Garbat and Munk, however, have shown that in this modification  $KClO_3$  is entirely inert and that the reaction depends upon the normal hemolysin in human serum against sheep's erythrocytes.

Several workers in this field believed that it would be advantageous to use a different species of blood in place of sheep's erythrocytes.

The only suggestion which sounds theoretically correct is that of Noguchi, who employs human erythrocytes and the serum of a rabbit immunized against human red blood cells. In this way he attempts to exclude the heterologous normal hemolysins, as human serum possesses no hemolysins against human blood cells.

- |  |  |
|--|--|
| 1. Syphilis extract or its substitute.                               | 1. Syphilis extract.   |
| 2. Inactive syphilis-serum.  | 2. Active defibrinated syphilitic blood. (Erythrocytes, "reagine," complement), one hour in incubator. |
| 3. Complement from human being or guinea-pig, one hour in incubator. | 3. Immune hemolysin of rabbit (injected with human blood).   |



4. Immune hemolysin of a rabbit against human erythrocytes.
5. Washed human erythrocytes.

From a practical standpoint, however, no distinct advantage is offered by these modifications. In fact, it is the claim of Wassermann and his pupils that by the use of human blood, the error tends toward the opposite direction, *i.e.*, the percentage of positive results obtained are higher than is actually the case.

The number of modifications have become so numerous that almost every one employs his own "method." *There is absolutely no necessity for this*, as an innovation justifies its existence only if it is a distinct improvement, *i.e.*, discloses a new fact or radically simplifies the old.

It is the classical Wassermann reaction performed in the original manner which has taught physicians how valuable a clinical aid it is. Their knowledge has not advanced materially with all the new changes. A single advantage only has been instituted through all this agitation, and that was the discovery that the luetic antigen can be replaced by *the alcoholic extract of guinea-pig's heart*. *In important differential diagnosis, however, even this extract should not be considered as specific as luetic liver antigen.*

For general work, however, its employment may be of service.

The antigen of Landsteiner, Müller and Pötzl is prepared as follows:

The heart of a guinea-pig is washed free of blood, its muscular part finely divided or macerated in a mortar and then extracted with 95 per cent. of alcohol for several hours at 60° C. One gram of the heart substance should be mixed with 5 c.cm. of the alcohol. The material is then passed through filter-paper, the filtrate being kept at room temperature. (The editor prepares the alcoholic extract by simply placing the finely divided guinea-pigs' hearts into 95 per cent. alcohol and allowing them to remain there for four weeks for purposes of extraction. At the end of this period the alcoholic solution is titrated and can be employed as antigen.)

These authors also employ the so-called drop method:

Drop Method. Ten drops of saline and 1 drop of normal guinea-pig's serum as complement are placed in each test-tube. The individual tubes receive the following additional ingredients:

First tube: One drop of the inactivated serum for examination.

Second tube: Same as one + 2 drops of the alcoholic heart extract.

Third tube: One drop of inactivated, surely luetic serum.

Fourth tube: Same as three + 2 drops of alcoholic heart extract.

Fifth tube: One drop of inactive normal serum.

Sixth tube: Same as five + 2 drops of alcoholic heart extract.

Seventh tube: Two drops of extract.

The tubes are well shaken and placed in the incubator for one hour at 37° C. Then 1 drop of a 50 per cent. (!) suspension of washed sheep's erythrocytes and 1 drop of hemolysin (double the minimum hemolytic titer) are added. After one-half hour in the incubator, the results are read.

Bauer's Modification. Bauer entirely excludes the immune hemolysin. His reaction requires the following ingredients:

1. Fresh guinea-pig's complement.
2. Alcoholic organ extract.
3. Five per cent. sheep's red blood corpuscles.
4. and 5. The inactivated serum for examination and an inactive normal control serum.

Four tubes are required for the reaction:

First tube: 0.2 serum, 1.0 c.cm. organ extract in dilution 1:5 and 1 c.cm. complement 1:10.

Second tube: Same as 1, but instead of organ extract, 0.85 per cent. sodium chloride.

Third tube: 0.2 c.cm. normal serum, organ extract and complement as in tube 1.

Fourth tube: Same as third tube, but instead of organ extract 0.85 per cent. saline.

The tubes are placed in the incubator for one-half hour and then 1 c.cm. of a 5 per cent. red blood cell emulsion is added.

After fifteen to forty-five minutes tubes 2, 3, and 4 show hemolysis, while tube 1 shows hemolysis or not, depending upon the absence or presence of syphilis.

Lipemic serum is not suitable for the reaction.

Bauer asserts that this method gives results identical with those obtained by the Wassermann tests. Heinrichs, Bering and others confirm Bauer's findings.

If the alcoholic extract made from luetic or normal human or animal organs is diluted with physiological saline, a milky opalescent solution results. The degree of turbidity of the resulting solution depends upon the rapidity with which the saline for dilution is added. If the first 15 to 20 drops of the latter are added slowly, the resulting solution will be much more turbid than if the saline is added quickly. Sachs first observed this phenomenon and stated that the more marked the turbidity the more active is the power of the antigen to bind complement.

The editor has worked with the guinea-pig's heart extract in thousands of tests and has found it to give perfect results. The amount usually used is 0.2 to 0.1 c.cm. in the first test-tube and 0.1 to 0.05 in the second test-tube as determined by titration. When the antigen is diluted (either 1:5 or 1:10) the first c.cm. of saline should be added drop by drop and shaken, thus producing a distinctly opalescent solution.

The author refrains from describing any other modifications in detail as they have not been verified sufficiently to merit a position in this important field of serum diagnosis. This holds true especially for the recently advised quick and easy short cuts by the use of the various ingredients dried on paper. In order, however, that one may acquaint himself with these modifications, if he so desires, the references to their original publications are here given.

Tschernogubow, Berlin. Klin. Wochenschr., 1908, No. 47, and Deutsche Med. Wochenschr., 1909, No. 15.

Weidanz, Deutsche Med. Wochenschr., 1908, No. 48, Refer.

Noguchi, Journal of Americ. Medic. Associat., 1908, No. 22, u. Münch. Med. Woch., 1909, No. 10.

Hecht, Wien, Klin. Wochenschr., 1908, No. 50, and 1909, No. 10.



Fleming, *Lancet*, 1909, 4474.

Stern, *Zeitschr. f. Immunitätsforschung*, 1909, Bd. I.

Bauer, *Deutsche Med. Wochenschr.*, 1909, No. 10.

#### IV. Serum Diagnosis of Echinococcus Disease.

The technique of this reaction is practically the same as described for the Wassermann test.

As antigen the cystic fluid of the human being or sheep is employed. The latter according to Weinberg is preferable, as human hydatid fluid sometimes reacts with normal serum.

The following is Weinberg's outline for performing the test:

Hydatid fluid from sheep	Inactive serum from patient	Complement	Hemolysin	Blood	Results
0.4	0.5	0.1	2×hemolytic dose.	1 c.cm. 5% sheep's blood.	0 hemolysis
0.4	0.4	0.1	2×hemolytic dose.	1 c.cm. 5% sheep's blood.	0 hemolysis
0.4	0.3	0.1	2×hemolytic dose.	1 c.cm. 5% sheep's blood.	Incomplete hemolysis
0.4	0.2	0.1	2×hemolytic dose.	1 c.cm. 5% sheep's blood.	Complete hemolysis
0.4	.....	0.1	2×hemolytic dose.	1 c.cm. 5% sheep's blood.	Complete hemolysis
.....	0.5	0.1	2×hemolytic dose.	1 c.cm. 5% sheep's blood.	Complete hemolysis
.....	0.4	0.1	2×hemolytic dose.	1 c.cm. 5% sheep's blood.	Complete hemolysis
.....	0.3	0.1	2×hemolytic dose.	1 c.cm. 5% sheep's blood.	Complete hemolysis
.....	0.2	0.1	2×hemolytic dose.	1 c.cm. 5% sheep's blood.	Complete hemolysis

Bauer's modification as employed for the Wassermann test can also be employed here.

#### V. Serum Diagnosis of Other Diseases.

The method of complement fixation can be employed in most diseases where an antigen can be made of the specific etiological agents, and where antibodies against this antigen have been formed in the serum of the infected subjects. The relative importance of the reaction in a given

disease depends upon its specificity and the percentage of positive results.

In epidemic meningitis, this method has been applied by Epidemic Bruck; while specific antibodies are undoubtedly formed, Meningitis. the diagnosis can more readily be made by bacteriological examination of the cerebro-spinal fluid.

In tuberculosis, Koch's old and new tuberculins are used Tuberculosis. as antigen. It is always best to use both of these preparations (0.05-0.2) as occasionally the serum will fix complement with one and not with the other product. The frequency of spontaneously formed antibodies in this disease is not sufficient to make this method of clinical diagnostic importance.

To Müller and Oppenheim (1906) belongs the credit of first Gonococcus applying the complement fixation test to the study of the Infections. gonococcus infections. Meakins, Vannod, Wollstein, Teague and Torrey have all contributed to this subject, but Schwartz and McNeil's careful observations of a larger number of cases have proven the distinct value of this method for the clinic.

In the *preparation of the antigen*, it is of paramount importance to use as many different strains of the gonococcus as possible. The failure to do this probably accounts for the many negative results in the early period of this test. The gonococci are best grown on a salt-free veal agar, neutral in reaction to phenolphthalein, for twenty-four hours; the growths are washed off with distilled water and the emulsion heated in the water bath for two hours at 56° C. It is centrifugalized and passed through a Berkefeld filter. Salt solution is added to the antigen just before it is to be used; it is then made up to 0.9 per cent. strength, by adding one part of 9 per cent. saline solution to nine parts of antigen. The latter can be kept indefinitely if preserved in small quantities in sealed tubes, heated to 56° C. for half an hour on three successive days.

The antigen must be titrated according to the principles outlined under the Wassermann reaction. First, one-half of the maximum dose of antigen which of itself does not bind complement is determined; then these non-fixing quantities are titrated with a known positive human serum or a highly immune rabbit serum; that dose of antigen is selected which binds complement the strongest with the smallest amount of serum. (Parke, Davis & Co. prepares such an antigen.) The antishoop hemolytic system is used.

Instead of using all the ingredients in one-half the quantity employed in the original Wassermann technique (as below), one-tenth of the latter quantity may be used.

Concerning the clinical value of the reaction, Schwartz and McNeil have thus far come to the conclusion that a positive reaction is indicative of a focus of living organisms, present in the body at the time or active



TABLE GIVING EXAMPLE OF TEST.

Antigen	Patient's serum	Complement 10 per cent.	Hemolysin	5 per cent. sheep's cells	Hemolysis
0.1 c.cm.	0.15 c.cm.	0.5 c.cm.	0.5 c.cm.	0.5 c.cm.	0
0.1 c.cm.	0.10 c.cm.	0.5 c.cm.	0.5 c.cm.	0.5 c.cm.	0
0.1 c.cm.	0.05 c.cm.	0.5 c.cm.	0.5 c.cm.	0.5 c.cm.	0
—	0.15 c.cm.	0.5 c.cm.	0.5 c.cm.	0.5 c.cm.	Complete.
—	0.1 c.cm.	0.5 c.cm.	0.5 c.cm.	0.5 c.cm.	Complete.
0.2 c.cm.	—	0.5 c.cm.	0.5 c.cm.	0.5 c.cm.	Complete.
—	—	0.5 c.cm.	0.5 c.cm.	0.5 c.cm.	Complete.
—	—	—	0.5 c.cm.	0.5 c.cm.	0
—	—	0.5 c.cm.	—	0.5 c.cm.	0
0.1 c.cm.	0.1 c.cm. of a positive serum.	0.5 c.cm.	0.5 c.cm.	0.5 c.cm.	0
0.1 c.cm.	0.1 c.cm. of a negative serum.	0.5 c.cm.	0.5 c.cm.	0.5 c.cm.	Complete.

only recently. A negative reaction does not exclude gonococcus infection. If the disease is limited to the anterior urethra, a positive reaction is not obtained; probably the absorption of toxins is insufficient to stimulate the production of antibodies. The positive reaction does not entirely disappear until seven or eight weeks after the patient is apparently cured. If it still persists, one must be suspicious of an active focus somewhere.

The complement fixation test should be of special usefulness in chronic cases where the bacteriological isolation of the gonococci is difficult, in gynecological conditions and in differential diagnosis of joint affections (Schwartz-McNeil, *American Journal of Med. Sciences*, May, 1911, Sept., 1912, Dec., 1912).

That the complement fixation test can also be performed  
 Typhoid in typhoid fever was first proven by Bordet and Gengou.  
 Fever. They used as antigen a suspension of typhoid bacilli in normal saline, and the serum from a convalescent patient. Since then (1901) numerous contributions referring to this subject have been published, but the merit of the test is variously interpreted. (Widal and Lesourd, Ludke, Leuchs, Schöne, etc.) The editor believes that to a great degree the variability in the results can be accounted for by the different antigens employed. Similar to the findings with the gonococcus complement fixation test by Schwartz and McNeil, Garbat has proven that a highly polyvalent typhoid antigen (made from numerous

strains) is absolutely essential. The serum from a typhoid fever patient gave a strong complement fixation test with an antigen made up from his own typhoid bacteria isolated from the blood but did not react with a similar antigen made from seven other different strains. The antigen is prepared like the artificial aggressins of Citron, by growing the bacteria on agar for 24 hours. The growth is washed off in a very small quantity of sterile distilled water; kept at 60°–70° C. for 24 hours (Leuchs); after this the emulsion is shaken thoroughly with glass beads for 24 hours and then centrifugalized until the supernatant fluid is absolutely clear.

The antigen is titrated as usual (see Wassermann reaction, gonococcus fixation test). The antisheep system is used.

On testing the bloods of thirty-six patients in different stages of the infection with such an antigen prepared from twenty-eight different strains, the editor found a positive result in all but one instance. [This patient died before the test could be repeated.] In ten cases two or three examinations were necessary before the reaction became positive. For the sake of comparison, agglutination tests and blood cultures were made at the same time. No distinct relationship between the three could be discovered. In several instances the complement fixation test appeared earlier than the Widal or even before the blood culture became positive. (In one, confirmed by autopsy, as early as the end of the first week.) As a general rule, however, the complement fixatives appear later during the disease, at a time when the bacteria have disappeared from the circulation. Thus, this method becomes of importance as corroborative of the Widal test. The positive reaction becomes stronger during convalescence and persists for several months after.

The exact clinical value of the reaction and its specificity require further statistics. One can assume, however, that almost all typhoid fever patients develop complement fixatives sooner or later, and that these can be detected if repeated examinations with a sufficiently polyvalent antigen are undertaken.

## VI. The Differentiation of Proteids by the Method of Neisser and Sachs.

The technique employed here varies only in a few details from the method advanced later by Wassermann and Bruck for the diagnosis of bacterial infections.

Neisser and Sachs do not employ a constant amount of complement (0.1), but first titrate the complement against the hemolysin in double its minimum hemolytic dose. For the test one and a half to two times the smallest amount of complement is necessary. The hemolysin consists of the serum of a rabbit immunized against ox's blood. This hemolysin acts both for ox's and sheep's erythrocytes.

The amount of antiserum (for example antihuman serum) used for the test, is influenced by two factors.



1. An excess of antiserum can interfere with the fixation of complement.
2. The antiserum if used in large quantities can bind complement of its own accord, without the addition of the human serum. It is therefore best to ascertain by titration the smallest quantities of antiserum which may satisfactorily be employed, as the complement fixation test must be sufficiently delicate to determine 0.0001 c.cm. of the human serum.

Diminishing amounts of antiserum are mixed with 0.0001 c.cm. of human serum and 0.1 c.cm. of complement. A control series is made wherein the human serum is replaced by the same amounts of saline. (The quantity in all tubes should be made uniform by the addition of normal salt solution, but the total amount of fluid in each tube should not exceed 2.3 to 2.5 c.cm.) The tubes are incubated for 1 hour and the hemolytic amboceptor and red blood cells added. After two hours at 37° C. the results are read. The 0.0001 c.cm. of the serum is added in the form of 0.2 c.cm. of a 1:2000 dilution.

TABLE III.

Amounts of antiserum in cubic centimeters.	Series A contains the different quantities antiserum+0.0001 c. cm. human serum (1:2000.02)+ 0.1 guinea-pig's serum. <u>One hour at 37°.</u> +0.001 c.cm. of amboceptor + 1 c.cm. of 5 per cent. ox's blood.	Series B (control) contains anti- serum+0.2 c.cm. physiological saline+0.1 of guinea-pig's serum <u>One hour at 37°.</u> +0.001 c.cm. of amboceptor + 1 c.cm. 5 per cent. of ox's blood.
	Hemolysis	Hemolysis
0.1	Faint trace.	} Complete.
0.075	Faint trace.	
0.05	0	
0.035	0	
0.025	0	
0.015	Trace.	
0.01	Slight.	
0	Complete.	

The antiserum itself as seen in the control series (B) does not exhibit any tendency to interfere with hemolysis, even in the amount of 0.1 c.cm. (larger quantities never come into consideration). On the other hand, series (A) shows that the larger amounts of the antiserum do not bind complement as thoroughly as the medium doses. The zone of complete complement fixation lies between 0.05 and 0.025 c.cm. of the antiserum. It is advisable as a general rule to choose about one and one-half to two times this minimum quantity. Thus from Table III it can be noted that 0.2 c.cm. of a 1:6 dilution could well be adopted as a test dose for com-

plement fixation. If it is required to know how delicate the complement fixation reaction can be with this dose of antiserum, the following experiment (Table IV) is performed.

Diminishing amounts of human serum are mixed with a constant quantity of complement and with this constant dose of antiserum. At the same time a control series of tubes is used, in which the antiserum is substituted by salt solution. After one hour of incubation at  $37^{\circ}$  C. erythrocytes and hemolysin are added.

TABLE IV.

Amounts of human serum in cubic centimeters.	Series A contains human serum +1:6×0.2 c.cm. <sup>1</sup> antiserum, +0.1 c.cm. of guinea-pig's serum.	Series B (control) contains human serum+0.2 c.cm. physiological salt solution+0.1 c.cm. guinea-pig's serum.
	One hour at $37^{\circ}$ . +0.001 c.cm. of amboceptor+1 c.cm. of 5 per cent. ox's blood.	One hour at $37^{\circ}$ . +0.001 c.cm. of amboceptor+1 c.cm. of 5 per cent. ox's blood.
	Hemolysis.	Hemolysis.
0.1	○	} Complete.
0.01	○	
0.001	○	
0.0001	○	
0.00001	Slight.	
0	Complete.	

It is seen from the above table that 0.00001 c.cm. of human serum still suffices to give a partial although incomplete fixation of the complement. The delicacy of the antiserum in this particular instance is not very great. In forensic practice, the reaction is carried out as shown in Table IV, but instead of the human serum, the solution of unknown blood stain in various dilutions is titrated. Control series B should not be omitted, because here, any foreign substance contained in the extract and which might interfere with the reaction can be detected.

<sup>1</sup> 1:6×0.2 c.cm. means 0.2 c.cm. of a 1:6 dilution.



## CHAPTER XV.

### PHAGOCYTOSIS. OPSONINS AND BACTERIOTROPINS.

#### I. Phagocytosis.

By phagocytosis is meant the taking up, or engulfing of foreign substances by certain cells (digesting cells or phagocytes) for the purposes of digestion. As a mode of nutrition, this is well known to exist normally, in the lowest unicellular animals, for instance the amebæ. Intracellular digestion, however, can be traced to organisms higher in the scale of the animal kingdom; even among mammals the function of cell ingestion is found, although limited to a definite group of cells, especially those derived from the mesoderm.

The inspiration for the work on phagocytosis and the greater part of its theoretical considerations have emanated from Metchnikoff and his numerous pupils at the Pasteur Institute at Paris. Metchnikoff divides the phagocytes into two classes, the "sessile or fixed phagocytes," and the "wandering phagocytes." The first is the stationary endothelial lining of blood vessels, and lymph spaces, as well as the large cells of the spleen pulp and lymph glands; the second consists of the white blood cells of the circulation. From another standpoint the phagocytes are divided into "microphages" and "macrophages." The former are practically identical with the neutro- and eosinophile polymorphonuclear leucocytes, while the latter present no distinct group, but include large lymphocytes, myelocytes, giant cells, etc. The cells designated as sessile phagocytes also belong to the class of macrophages. The size of the cell was considered by Metchnikoff as the deciding feature; not all macrophages are mononuclear as generally believed. Thus macrophages appearing in the peritoneal fluid of guinea-pigs frequently possess, like the giant cells of the tubercle, numerous nuclei. According to Metchnikoff it is primarily the microphages to whom the function of bacterial phagocytosis is allotted, while the macrophages serve for the purpose of ingesting dead or moribund tissue structure. Still there are certain pathogenic micro-organisms, tubercle bacilli, lepra bacilli, actinomyces, which are favored in that they also are digested by the selective macrophages. The evidence of phagocytosis is established by mixing either in vitro or vivo the substance for phagocytosis, plus the phagocytes, and noting the changes which ensue; [either in a stained or unstained preparation]. The phagocytes of the guinea-pig's peritoneal cavity are especially well adapted for the study of phagocytosis in vivo. The following experiment of Metchnikoff may serve as a type.

A guinea-pig receives an intraperitoneal injection of goose's blood. Immediately following this, the leucocytes disappear from the peritoneal fluid. This is due partly to a destruction of leucocytes (Phagolysis) and partly because the leucocytes are repulsed and settle upon the peritoneal wall. In one to two hours this so-called negative phase is overcome and there is an increase of the leucocytes, especially of the macrophages in the exudate (Hyperleucocytosis). Now, the leucocytes can be seen sending forth short protoplasmic processes—pseudopodia, by means of which the erythrocytes are drawn into the phagocytes. After a short time the macrophages are filled with the erythrocytes. At first the ingested cells appear normal; gradually, however, they undergo changes, which are clearly visible in the unstained specimen, indicative of a disintegrating process, within the body of the phagocytes.

The same phenomenon as described for goose's erythrocytes can also be observed with bacterial bodies.

In order to exclude the possible bactericidal influences of the serum, it is advisable when one is working with bacteria which are readily destroyed, as cholera vibrios, to previously induce a hyperleucocytosis in the peritoneal cavity. The guinea-pig receives an intraperitoneal injection of 10 to 20 c.cm. of sterile bouillon or aleuronat solution; in about twelve hours hyperleucocytosis takes place, and a capillary pipette inserted into the peritoneal cavity will withdraw a thick and turbid exudate of leucocytes.

If this animal is injected intraperitoneally with bacteria, and a smear of the peritoneal fluid made a short time after the inoculation, the bacteria will be seen lying within the microphages. This important fact has been variously interpreted. Pfeiffer and his pupils claim that the bacteria are first destroyed or their virulence greatly diminished by the bactericidal power of the serum and exudate, and that the phagocytes act only as receptacles for these already destroyed bacteria. Metchnikoff believes that the phagocytes take up the living bacteria and destroy them, thus representing these cells as the most important weapons of the organism in its protection against infection.

"Whenever an organism, that has lost its susceptibility to a particular infection, either on account of a natural born immunity or one artificially attained, comes into conflict with the etiological agent, a struggle arises between the latter and the phagocytes of the threatened individual. The phagocytes appear as victors, since they take up the bacteria into their protoplasmic bodies and digest them, thus forever destroying the evil." (Metchnikoff cited by Levaditi.)

Critically considered, there can be no doubt that the phagocytes by their very nature are capable of dealing with living virulent bacteria. At the same time one must observe that the opsonins and bacteriotropins of the serum, soon to be discussed, in most instances previously modify the living bacteria in a way at present still unknown. That, however, the phagocytes can ingest bacteria or protozoa which are alive and active, has been demonstrated by Metchnikoff's school. Phagocytosis experiments were under-



taken with motile bacteria and spirilla. On microscopical examination it was seen that a phagocyte was in the act of taking up a spirillum, part of which was engulfed by the cell while the remainder was still outside of the cell and continuing its active motility.

Not in all cases does phagocytosis of bacteria lead to destruction of the ingested microbes. More recent experiments also seem to prove that simple phagocytosis of bacteria must not be considered as identical with their death. Thus, the exudate from cases of anthrax in which the bacilli lie within the leucocytes, can still produce fatal anthrax when inoculated into animals.

**Vital Stain-** A more exact understanding of the bio-chemical nature of ing with phagocytic digestion has been offered by the method of vital Neutral Red. staining with neutral red.

Neutral red (used as a 1 per cent. solution in isotonic saline) is a chemical dye which stains only dead cells and not living ones. If live bacteria and phagocytes are mixed and hanging-drop preparations of these are made, and then a drop of the stain be added to different preparations at successively increasing intervals of time, the first slide shows the extracellular living bacteria unstained, while of the intracellular bacteria, a part remains unstained and the other colored red.

The later the mixtures are stained, the more numerous are the intracellular red stained bacteria, showing that the ingested micro-organisms remain alive for a short time, and then die. The intracellular bacteria retain their stain as long as the phagocytes themselves remain alive. Later, when the phagocytes die, the formerly red bacteria lose their stain. Metchnikoff's explanation of the red staining process is that during the act of digestion by the phagocytes an acid ferment is liberated which gives the color reaction with the neutral red.

For many years Metchnikoff's phagocytic theory opposed the conception of Ehrlich and also Pfeiffer in relation to the importance of amboceptor and complement in the mechanism of immunity. It would be out of place here to review the various experiments performed and offered on each side in explanation of its standpoint. Suffice it to say that Metchnikoff denied the existence of free complement within the animal organism. He moreover claimed that the complement was found normally only in the phagocytes and hence called it "cytase," differentiating the two phagocyte groups as "micro- and macrocytase." The "cytase" is liberated when the phagocytes are broken up. The amboceptors are considered as split products of the phagocytes and known by Metchnikoff as "fixators."

## 2. Opsonins.

In recent years the closer agreement which has arisen between the followers of phagocytic and humoral theories was made possible by the fact that Denys and Leclef, Leishmann, Wright and Douglas and others,

demonstrated that phagocytosis occurs in most cases only in the presence of serum. If the phagocytes are thoroughly washed, so that they are entirely serum-free, phagocytosis will not take place, or will do so imperfectly. The belief of some authors that "spontaneous phagocytosis" without serum was altogether impossible was disproved, especially by Löhlein. The manner in which the serum acts, whether it stimulates the digestive activity of the leucocytes or whether it so changes the bacteria that they can more readily be taken up by the phagocytes, has been settled in favor of the latter view through researches, especially of Wright and his followers as well as of Neufeld. The substances within the serum which thus modify the bacteria have been designated by Wright as "opsonins." ("opsono" = I prepare food for.)

Opsonins are demonstrated by mixing bacteria, serum and washed leucocytes, allowing this mixture to remain in the incubator for a short time, and then staining smear preparations. Wright then counts a certain number of leucocytes and the number of bacteria found within these leucocytes. *The relation between the number of ingested bacteria and the counted number of phagocytes is designated as the*

*The Opsonic phagocytic count. Wright compared the phagocytic counts of infected individuals with those of normal persons and found that those of the former were much lower. The relation existing*

*between the two he expressed in the form of a fraction and that is known as the opsonic index.* Thus a smear made from a mixture of equal parts of an emulsion of staphylococci, leucocytes and the patient's serum showed for example 75 cocci to 100 leucocytes; while one made from a mixture of equal parts of the same bacterial emulsion and leucocytes, but a normal individual's serum, demonstrated 150 bacteria to 100 leucocytes. The opsonic index of the patient's serum was therefore one-half (0.5).

According to Wright, the opsonic index expresses the animal's resistance to infection. He believes that a low opsonic index for a given bacterium indicates a susceptibility on the part of the individual for that particular infective agent. Furthermore, the opsonic index he claims can be used as an aid in the diagnosis of infectious diseases, inasmuch as opsonins are specific. Thus the opsonic index in a tuberculous individual is low only for the tubercle bacillus and not for other bacteria.

When an animal is immunized, its opsonic index toward the respective bacterium is considerably increased. The question has been asked whether the immune opsonins formed during this process are identical with the normal opsonins. Wright and a number of the more recent authorities believe that they are different. Neufeld, who discovered these immune opsonins independently of Wright, named them Bacteriotropins, and pointed out that while the normal opsonins are destroyed when heated to 56°, the bacteriotropins remain unharmed. As yet the exact nature



of the immune as well as of the normal opsonins has not been clearly defined. It is still a matter for investigation whether in the case of opsonins one is dealing with entirely new substances or whether they are the old well-known bodies like the agglutinins, complements and amboceptors with a new action.

The fact that the opsonic index is raised by immunization while it is usually found diminished during spontaneous infection in man, led

Wright to believe that good results may be obtained by increasing the opsonic index of the already infected individual by means of immunization. In this way he thought the patient's predisposition to the particular infection would be overcome. Wright's experiments showed that the opsonic index could be increased by injection of extremely small doses of dead bacteria (Wright's vaccines.)

If an individual suffering from an acne or furunculosis, and who has a low opsonic index for the staphylococcus, is injected with a very small number of staphylococci, his opsonic index sinks still more for a short period after the inoculation (negative phase).

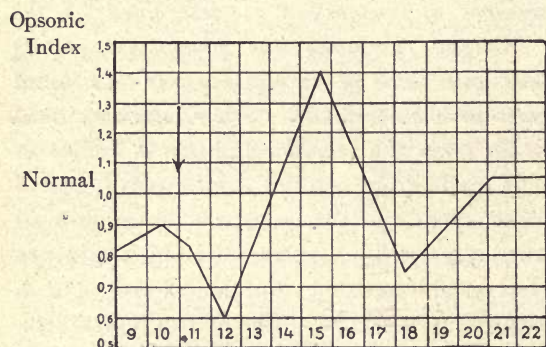


CHART 5.—Curve of the opsonic index following the inoculation of a small dose of staphylococcus vaccine. The arrow indicates the time of injection.

This is explained by the fact that the injected bacteria absorb the existing opsonins. New opsonins are, however, then produced, which immediately make up for the loss occasioned during the negative phase, with the result that after several days there is an increase of the opsonic index (positive phase) which lasts for a short time. Then the index again begins to fall, as the stimulus to the formation of opsonins is transitory. It usually sinks to below the normal level, only to rise again to a point slightly above the normal, where it remains stationary. This irregular curve represents the typical course of the opsonic content of the blood after a vaccine injection; apart from this characteristic picture numerous exceptions exist. Thus by the use of very minute bacterial doses, the negative phase immediately following the injection is entirely absent. Reversely very large doses exhibit a prolonged negative phase.

Wright graphically represents these variations in the opsonic index by charts, an example of which is given here (Chart 5).

In order that the therapeutic effect may persist, it is advisable to repeat the inoculation. A new injection should be given at the height of, or during the positive phase, as an inoculation repeated during a negative phase will result in further depression of the index to a very low level. It is even possible in this way to harm the patient. The

poor results obtained during the first era of tuberculin treatment can, according to Wright, be attributed to the failure of this observation. It is the production of cumulative positive phases that is the aim of vaccine treatment. (Chart 6.)

Wright and his co-workers have noticed that an increase in the opsonic index usually runs parallel with an improvement in the condition of the patient.

Inasmuch as an increase in the opsonic index is occasioned by introducing into the general system even a very small number of bacteria, it seems probable that such spontaneous inoculation will take place during the course of an infectious disease. In fact, a spontaneous rise in the opsonic index is observed during convalescence or after the crisis of an infection. A high index is, however, also noticed at other times; for example, tuber-

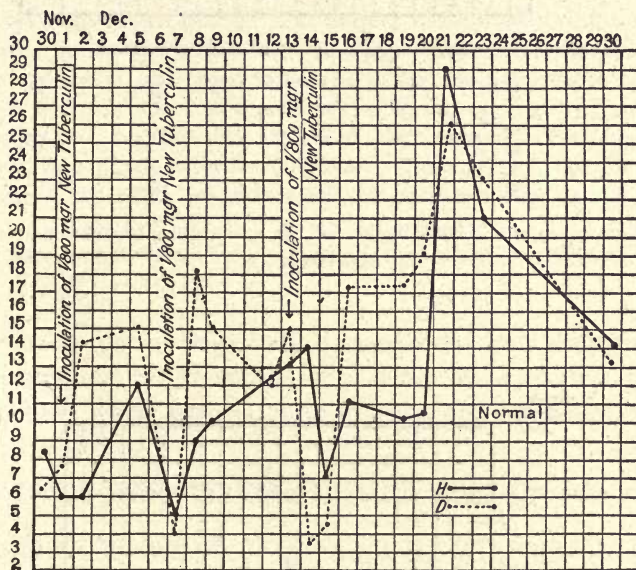


CHART 6.—Opsonic curve during treatment with New Tuberculin.

culous individuals show a higher index than normal persons. Wright explains this by the so-called "auto-inoculation;" for example, after moderate exercise, or work, tuberculin is liberated from the tuberculous focus and in this way acts like a therapeutic injection of tuberculin, *i.e.*, the index will be raised. Therefore, an excessively high opsonic index is of just as great diagnostic value as a low one. Wright furthermore believes that constant irregularities or variations in the height of the opsonic curve serve as plausible evidence for the existence of infection, because under normal circumstances the curve should remain at a level. Not infrequently, however, cases come under observation where in spite of a



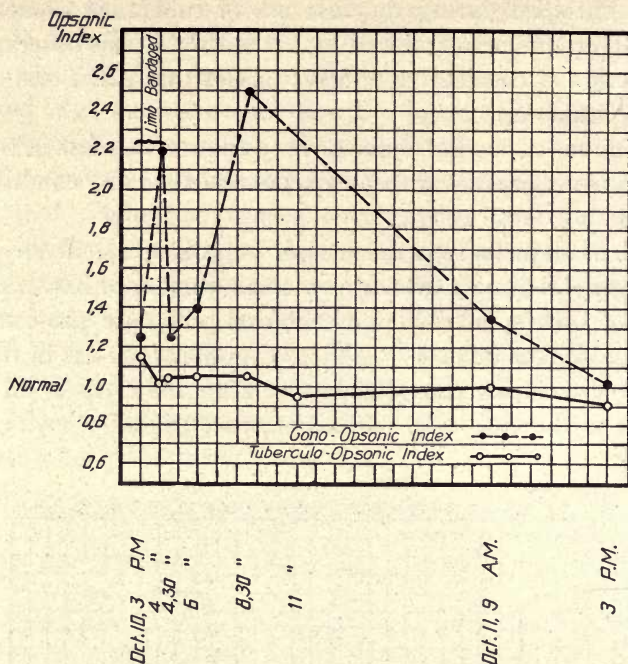


CHART 7.—Increase in the opsonic index for the gonococcus brought about by Bier's hyperemia.

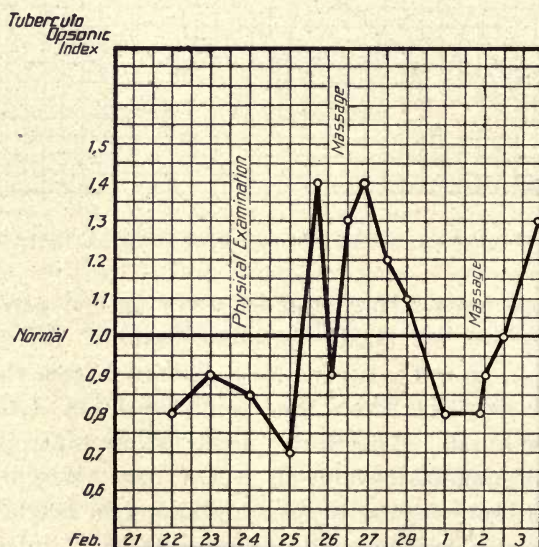


CHART 8.—Tuberculin auto-inoculation following physical examination and massage. (Tuberculous Lymphadenitis.)

distinct evidence of the existence of an infection the opsonic index remains normal. In such instances, for some reason, the bacteria and their products do not reach the general circulation and therefore no occasion is offered for either an elevation or sinking of the opsonic index. Wright and Freeman were able to show that all active and passive motions of an infected joint, as well as any vascular changes which induce a flow of lymph toward the focus of infection, lead to auto-inoculations, which are manifested in a change of the opsonic index. Such artificial production of auto-inoculation can be employed in various forms as a means of diagnosis: thus, in articular rheumatism, massage; in pulmonary tuberculosis, breathing exercises; in laryngeal diseases, loud reading; and in tuberculosis of the lower extremities, active gymnastics will occasion changes in the opsonic curve.

An example is given in Chart 7. The patient was a woman with a swollen wrist joint. In order to decide whether this was a gonorrheal or tuberculous process, the opsonic index was taken and found to be 0.94 to 0.97 for the gonococcus and 1.03 to 1.35 for the tubercle bacillus. As these figures differed very slightly from the normal, the test was repeated, but this time after Bier's hyperemia had been applied and the forearm placed into warm water for one hour. The opsonic index for the tubercle bacillus remained the same, while that for the gonococcus had undergone marked variations.

A similar experiment with a woman having tuberculous lymphadenitis is given in Chart 8.

Wright makes use of these variations of index caused by auto-inoculation in determining the prognosis of a case. An infection is only then considered cured when artificial auto-inoculation is no longer possible.

### **The Technique for the Determination of the Opsonic Index.**

For the determination of the opsonic index are required,

1. Serum of the patient.
2. Serum of the normal individual (as control).
3. Washed blood cells (Leucocytes).
4. Bacterial emulsion.

The blood serum is obtained from the finger tip at the root of the nail. It is most efficacious to first produce a hyperemia of this part by constricting the finger either with a narrow gauze bandage or a small soft rubber tube (the editor has found the latter much more convenient). The prick is then made with a needle or finely drawn out glass tube. The blood flows spontaneously and is collected into one of Wright's capillary tubes (Fig. 19) by approximating the curved end of the latter to the blood (Fig. 18). The straight capillary end of the tube (away from the blood) is then warmed in a small flame and sealed. The tube is laid down flat, and



allowed to cool; by the cooling the blood is sucked back from the unsealed capillary end; this end may also be sealed in the tip of the flame. The blood then coagulates and the serum separates off. The separation of the latter may be hastened by centrifugalization for a short time.

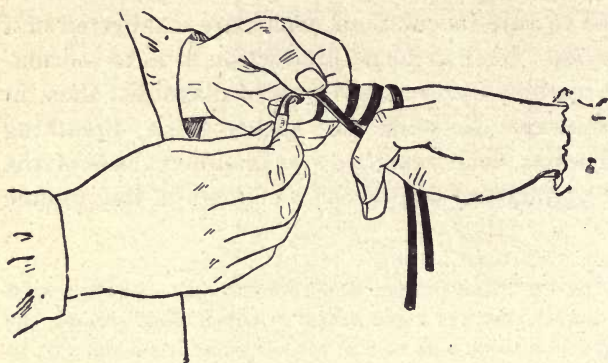


FIG. 18.



FIG. 19.



FIG. 20.

In order to obtain leucocytes, a small test-tube which holds 3 to 4 c.cm. is filled  $\frac{2}{3}$  with a 1.5 per cent. solution of sodium citrate, and about 6 to 7 drops of blood from a healthy individual are collected in this solution (Fig. 21). The tube is inverted several times to thoroughly mix

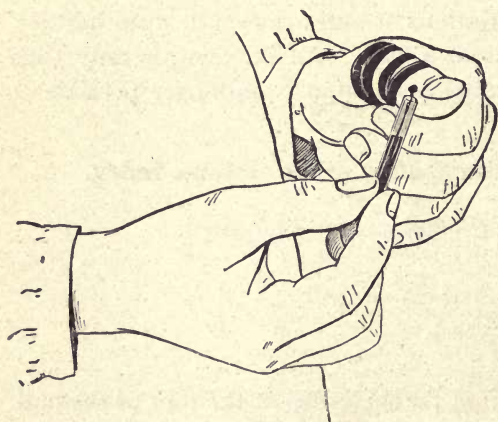


FIG. 21.



FIG. 22.

the blood so that the citrate, by precipitating the calcium salts of the blood, effectively prevents coagulation. The suspension is centrifugalized until the corpuscles are thrown down and a distinct white layer (leucocytes) is seen upon the surface of the red cells (Fig. 22). The clear supernatant citrate solution is pipetted off, care being taken not to disturb the white layer. Some 0.85 per cent. saline is added, mixed and the mixture again

centrifugalized. The washing with normal saline solution is repeated once or twice and as much of the clear liquid as possible is finally removed; the remaining cells are thoroughly mixed and in this form are ready for use.

The bacterial emulsions with the exception of the tubercle bacillus are made from agar cultures; the growths of gram + cocci may be as old as twenty-four hours, while the coliform organisms and the gram - cocci are preferably used only four to ten hours old (the younger the better). A loopful of culture from an agar tube is thoroughly rubbed up with several drops of salt solution in a watch-glass by means of a small glass pestle. The salt solution is best added very gradually, drop by drop, in order to make a more perfect emulsion. This may then be advantageously centrifugalized for a short period, to bring down the large clumps. The supernatant opalescent portion is

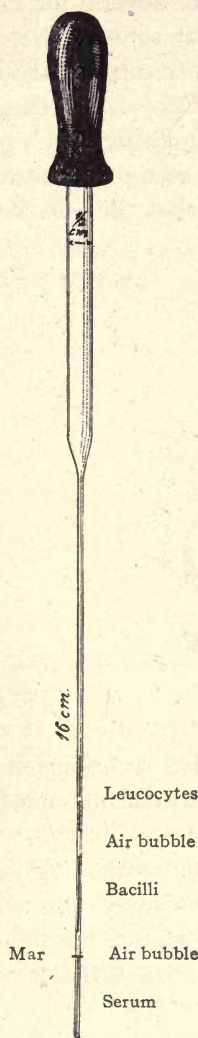


FIG. 23.

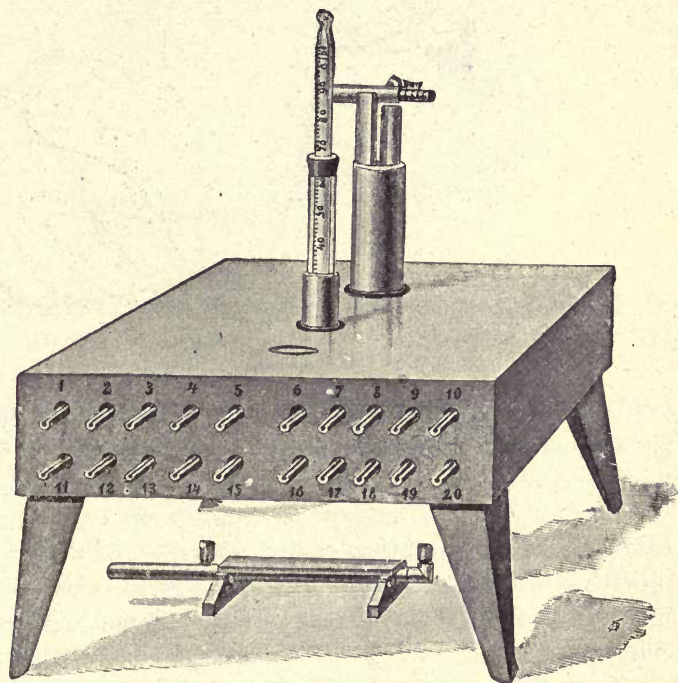


FIG. 24.

taken off for use, thoroughly mixed, and if necessary diluted. Emulsions of coliform organisms are the most easily made. Frequently it is sufficient to rub up with the platinum loop a loopful of such bacteria on the side of a small test-tube containing saline. The proper thickness of the resulting emulsion varies. As a rule, bacillary emulsions are re-



quired to have a thicker appearance to the naked eye than coccal ones. The latter should be only slightly opalescent.

In order to make a satisfactory *tubercle emulsion*, a more elaborate method is necessary. The dead and dried tubercle bacilli are employed for this purpose. A portion of these bacilli is very thoroughly triturated in an agate mortar, or between two slides, or in a grinder devised for this purpose, at first alone and then with 1.5 per cent. salt solution added drop by drop. In this way a paste, and subsequently a comparatively thick emulsion is made. For use, a small portion of the resultant emulsion is centrifugalized until the upper layers are fairly opalescent.

These upper layers are pipetted off, and thoroughly mixed. A smear of this should be made and stained in order to observe that the emulsion

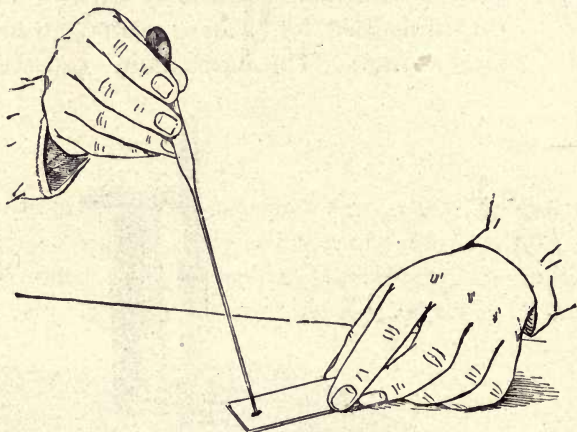


FIG. 25.

is free from clumps and not too thick. Such an emulsion sealed up in a glass tube and sterilized at 60° C. for 1 hour can be kept for about one week.

Streptococci may similarly be rubbed up in a mortar with 0.85 per cent. salt solution and then centrifugalized. As a rule, however, vigorous pipetting into a watch glass with subsequent centrifugalization for a few minutes is sufficient to break up the chains and leave a satisfactory emulsion.

If several specimens of blood are to be examined it is best to do a preliminary phagocytic count in order to test the strength and condition as regards clumping of the emulsion. The phagocytic count for tubercle should be between 1.5 to 2 per cell and for other organisms not less than 3 per cell. According to the preliminary finding further dilution or concentration of the emulsion is necessitated. The pipettes employed for the opsonic index should be about 16 cm. long and made from glass tubing about 5/16 of an inch in diameter. They should all be approxi-

mately of the same caliber and but slightly tapering toward the point. The rubber nipple should tightly fit the piece of tubing or bulb available. For use, the capillary end should be cut square and the pipettes marked with a paraffin pencil about  $\frac{3}{4}$  of an inch from their extremity. The content as far as this mark is the unit of volume in each case.

The rubber nipple is now held between thumb and forefinger and gently compressed, the capillary end introduced into the well-mixed blood cells and the unit volume drawn up by slightly relaxing the pressure on the bulb. Next a tiny bubble is allowed to enter, then an equal volume of the emulsion, followed by another tiny bubble which latter is succeeded by an equal volume of serum. By gentle pressure on the bulb the several volumes are ejected upon a clean glass slide, and thoroughly mixed by alternately sucking the mixture into the pipette and squeezing it out again upon the slide. It is enough to repeat this action three times. Then the

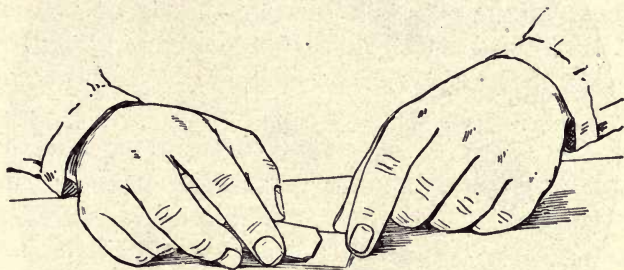


FIG. 26.

mixture is drawn up into the pipette, the end sealed in a small pilot flame, the pipette placed into the opsonizer (Fig. 24) and the time noted. This operation is repeated with each serum.

Coliform organisms and the gram negative cocci should be incubated not longer than six to eight minutes. Tubercle bacilli and other organisms require fifteen minutes, more or less according to the strength of the emulsion.

The pipettes are withdrawn in the same order in which they were placed in the opsonizer. The contents of each are blown out on to a slide and very carefully mixed as before (Fig. 25). The entire quantity is divided between two or three slides and several smears are made, the best one being selected for counting. These slides should previously have been roughened with very fine (00) emery paper and cleaned with a duster, and should rest on their concave surface so that the smear is made on the convex side. (It will be noticed that a slide can be made to rotate if resting on one surface (convex), but does not do so when resting on the concave surface.) The smears are best made by means of a broken slide with a slightly concave edge. This "spreader" (Fig. 26) is made by sharply breaking a glass slide at about its middle, this being



facilitated by scratching the edges of the slide with a glass cutter at the point where it is desired to break it. The editor has broken as many as twenty to thirty slides before a proper spreader was obtained. It pays to do this, because upon the sharpness of the fracture and cleanliness of the spreader depends the edge of the film, and secondarily the ease, rapidity, and accuracy of the count. If the film be well made, it will have a straight edge within which will be found practically all the leucocytes, as they are larger than the red blood cells, and therefore dragged to the end of the film.

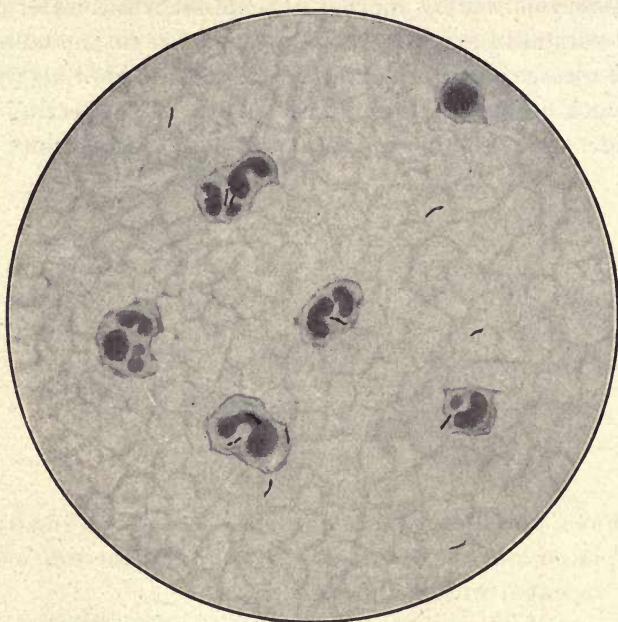


FIG. 27.—Phagocytosis of tubercle bacilli.

The preparations are fixed in a saturated solution of corrosive sublimate for two or three minutes, washed with water, and stained with methylene blue or carbol-thionin ( $\frac{1}{4}$  per cent. thionin, and 1 per cent. carbolic acid). Carbol thionin is preferable. It should be slightly diluted and warmed before being poured upon the slide. Here it is allowed to remain for several minutes, then washed off in water, and the slide dried with filter-paper. The tubercle films are best fixed with formalin vapor, stained with hot carbol or aniline fuchsin, decolorized in 2.5 per cent. of  $\text{H}_2\text{SO}_4$ , treated with 4 per cent. acetic acid to dissolve the erythrocytes and counterstained with  $\frac{1}{2}$  per cent. of methylene blue in  $\frac{1}{2}$  per cent. of sodium carbonate. It is most important that tubercle films be carefully stained because it is desirable to color every bacillus and yet not break up the leucocytes (Fig. 27).

With a  $\frac{1}{12}$  inch oil immersion lens a minimum number of one hundred

polymorphonuclear leucocytes are now examined and the number of microbes they contain enumerated.

Similar calculation is undertaken with the normal control serum. The fraction obtained by dividing the number of bacteria contained in 100 cells on the patient's slide, by the number in 100 cells on the normal slide, gives the opsonic index of the patient's serum.

For example, the normal individual has 284 and the patient 262 bacteria in 100 cells, the fraction which gives the patient's opsonic index would be  $262/284$  or 0.92.

The principle of Wright's technique is simple, but it requires a great deal of practice before it is mastered. Only then are the results reliable. One must adopt the same principles when counting the control slide as when the patient's film is counted. If in the last case, for instance, the cocci situated on the edges of the cells are not included in the count, they should also be excluded in the first case. The absolute count is of no importance. It is the relative proportion which is significant.

As a normal control, it is best to take the average of the phagocytic counts of a series (3 to 4) of normal sera or first equally mix the different sera, and take the phagocytic count of the pool.

Normal sera should not differ from one another in a tubercular opsonic estimation by more than 10 per cent.

### Wright's Vaccine Treatment.

As has been said, the principle of Wright's vaccine treatment depends upon the immunization with small doses of dead bacteria, so-called vaccines, whereby the opsonic index of the individual is raised. This is usually associated clinically, with improvement in the patient's condition.

The effect of the immunization according to Wright depends upon:

1. Individual reaction of the patient.
2. Preparation of the vaccine.
3. Dosage and form of application.

The individual reaction of the patient can be measured by the opsonic index.

As far as the preparation of the vaccine is concerned, Pasteur's contention that a vaccine must necessarily be made up of living cultures has not proved itself correct. Carefully killed cultures suffice in almost all cases. An example of the preparation of Wright's vaccine is here given.

#### *The Preparation of a Staphylococcus Vaccine.*

Agar cultures are grown for twenty-four hours, and about 3 c.cm. of sterile normal saline solution are added to each culture. The growth is washed off in the saline solution by means of a platinum needle or freshly



prepared capillary pipette. The suspension of bacteria is placed in a sterile test-tube, the end of this tube drawn out in the blow-pipe flame and sealed. The drawn-out portion should be about 2 inches in length and as strong as possible. The emulsion is now vigorously shaken for fifteen minutes. The extremity of the drawn-out tube is then cut and a few drops of the emulsion expelled into a clean watch glass, or a small part of the drawn end is cut off so that a portion of the emulsion is still contained within it. The tube is resealed, and then submerged in water kept at 60° C. for one hour. This usually suffices to kill the bacteria.

The small amount placed in the watch glass or in the capillary test-tube serves for the standardization, which is carried out as follows: A pipette with rubber bulb, as prepared for the opsonic-index test, is also used here. A volume of freshly drawn blood of known corpuscular content, best taken from the worker's own finger, and an equal volume of bacterial emulsion are mixed thoroughly with six or seven volumes of 1 1/2 per cent. citrate solution; several even films (which may be fairly thick), are then made by means of the ordinary edge of a slide, and stained with carbol-thionin, Leishmann's or Jenner's stain.

The entire smear is divided up (with a blue skin pencil) into eight equal subdivisions, by one transverse line drawn parallel to the long diameter of the slide at its middle and five vertical lines, one at each edge of the smear, one in the center and one equally distant between the edge and the central line. It is also advantageous to employ an eye piece, the field of which has been divided or made very much smaller by the insertion of a paper screen with a small central opening representing the size of the desired field. Five or six fields are then counted in each of the eight subdivided areas. The number of red blood cells seen in each field are enumerated in one vertical column, the number of organisms in the same field in another column. In this manner an average of the entire slide is obtained.

By means of a simple proportional sum, the number of bacteria per cubic centimeter of emulsion is estimated, *e.g.*, the number of red blood cells counted is 850 and the number of bacteria 1020. The red blood corpuscles used in the standardization are known to number 5,000,000 to a cubic millimeter or 5,000 million to a cubic centimeter; therefore the number of bacteria to a cubic centimeter of the unknown emulsion is expressed as follows.

$$850 : 1020 :: 5,000,000,000 : \text{No. of bacteria per c.cm. of emulsion,} \\ \therefore 6,000,000,000 = \text{the number of bacteria per c.cm. of emulsion.}$$

After the emulsion has been heated for one hour, the tube is opened and a drop is expressed into an agar culture tube which is incubated for twenty-four hours to demonstrate whether the emulsion is sterile or not. At the end of this time, if a growth is observed, the emulsion must be heated again for one hour at 60° C. and its sterility again tested for.

Proper dilution of the emulsion is next undertaken. Small bottles containing 25 c.cm. of 1/2 per cent. carbolic acid in sterile saline are aseptically closed with rubber caps; for example, it is desirable to make up these 25 c.cm. with staphylococcus vaccine so that each cubic centimeter contains 500 million bacteria, then

$$\frac{\begin{array}{l} \text{(desired amt. to each c.cm.)} \\ 500,000,000 \times 5 \text{ No. of c.cm. desired} \end{array}}{6,000,000,000 \text{ (dose of original emulsion per c.cm.)}} =$$

2.08 c.cm.; or approximately 2 c.cm. of the original emulsion must be added to the 25 c.cm. (to be exact 23 c.cm.) to make up the desired dilution.

The rubber cap is finally coated with melted paraffin wax.

For stock vaccines it is best to make up the different vaccines in the following concentrations:

1. Staphylococcus vaccine—prepared from various strains of staphylococcus, aureus, citreus, and albus, in three concentrations: 1000 million, 500 million and 100 million, to the c.cm.

2. Streptococcus vaccine in 20 mil., 10 mil. and 5 mil. concentrations. Since the streptococcus grows very sparingly, cultures of two or three days' growth may have to be employed for the preparation of a vaccine, and even then it may be necessary to use one broth culture instead of sterile salt solution to emulsify the agar cultures. On standardizing such thin vaccines it is frequently necessary to take one volume of blood to two, three, or even more volumes of emulsion and then calculate accordingly.

3. Acne vaccine in 20 mil., 10 mil. and 8 mil.

4. Mixed acne in 20 mil. acne and 500 mil. staphylococcus.

5. Gonococcus vaccine in 50 mil. and 5 mil. Gonococcus vaccines are best employed as autogenous vaccines.

6. Typhoid vaccine in 1000 mil. and 2000 mil. for prophylactic inoculation.

7. Colon vaccine in 25 mil., 10 mil., 5 mil. Vaccines of coliform organisms are very easily emulsified; as a rule they should not be older than twelve hours and not be sterilized for more than three quarters of an hour.

With the exception of the staphylococcus vaccines, it is advisable not to use stock vaccines, but autogenous vaccines, *i.e.*, vaccines made from the specific strain of bacteria causing the infection to be treated. It is very important to isolate the supposed pathogenic organism from the innocuous or less pathogenic bacteria contaminating or complicating the infection.

In tuberculosis Wright employs a dilution of Koch tuberculin (T. R.). Recently he has prepared a tubercle bacillus vaccine in the same way as the other bacterial vaccines.

The initial dosage varies with the different vaccines, but should in general be about 100 to 500 million of staphylococci; one may go as high as 2,500 or even 5,000 millions.



In colon, streptococcus, gonococcus and acne, doses of 1 to 3 million should be used at the beginning and then gradually increased.

In tuberculosis Wright starts with the T. R. in dilution equivalent to about 1/1000 mg. of the dry tuberculin substance and this is increased to about 1/600 mg.

Wright cites two general rules to be observed in the therapy of infectious diseases.

1. In all cases where the normal antibacterial power of the blood has been lowered, immunization is indicated.

2. Whenever the blood possesses strongly active curative powers, an increase of the blood supply to the infected part should be attempted in order that the antibacterial elements of the blood and leucocytes may display their effect. In such cases the production of hyperemia is particularly of help. Similarly, massage and other such therapeutic measures can be useful.

The therapeutic value of auto-inoculation is very slight and should not be encouraged, as in this way the exact dosage cannot be followed.

Wright has employed these vaccines in staphylo-, strepto-, and gonococcus infections, as well as in coli infections, tuberculosis, Malta fever and carcinoma (!) where injections of the bacillus neoformans Doyen were given.

From a critical review of the cases published, which were treated with vaccines by Wright and his fellow workers, one certain conclusion can be reached; namely, that given an infection, inoculations with small doses of the respective dead or extracted homologous bacteria, will result in a therapeutic immunization. Although Koch had advanced the same principle for the treatment of tuberculosis, it is Wright who first recognized the general application of this form of immunity. Furthermore, by means of his opsonic studies, he was able to prove that by the injection of even the minutest doses, for example 1/1,000,000 c.cm. of tuberculin, immune reactions are incited.

In spite of this finding, investigators are still at variance over the question, and two camps exist: one of which believes that the ideal treatment of tuberculosis consists in the repetition of the small doses; the other, that the best results are obtained by gradually increasing the dose of tuberculin until very large doses are administered. Citron has found the latter course more satisfactory.

Since, as is known, tuberculin is one of the harmful agents in tuberculosis infections, it seems more advantageous to get the patient, if possible, into a condition where he is able to neutralize large doses of tuberculin rather than to have him at a stage where even moderate doses suffice to give a reaction.

Other questions of importance in the vaccine therapy are: first, whether any parallelism exists between the increase in opsonic index and improvement in the clinical manifestations; second, whether the opsonic index must necessarily be used as a guide in vaccine treatment.

As to the first, Wright has pointed out numerous cases where exact

study has proved that such parallelism exists. This fact is probably correct in the majority of instances, but it cannot be considered as an infallible rule, inasmuch as the formation of opsonins is only one of a great number of factors in the complicated process of healing; consequently one should not be surprised when in some instances in spite of a rising opsonic index, the patient's clinical condition becomes worse, and reversely when in some cases improvement occurs although the opsonic index does not change.

Accordingly, the opsonic index during the course of treatment becomes secondary in importance to the exact clinical observation of the case. Wright and his school have shown that certain bad effects may follow from the injection when performed during the negative phase. With the use of small doses the negative phase becomes short—only one day or even less; accordingly it is very probable that this state is entirely passed when an injection is repeated on the fifth to eighth day.

The tuberculin therapy at the Kraus clinic is conducted on this principle, without estimation of the opsonic index. And yet, no harmful effects have ever been noted; while general improvement, as increase in weight, diminution in temperature and cessation of cough, are constantly observed. It would be illogical to neglect these clinical data and give preference to the hypothetical action of opsonins as a guide in treatment.

It seems that Wright himself does not insist as strongly as before upon the determination of the opsonic index. One of his assistants, Matthews, has made the statement that in a great number of cases the determination of the opsonic index is entirely out of the question. If the choice between injections without estimation of the index and entire omission of inoculation should arise, therapeutic inoculation without the index is by all means indicated. There is a general tendency at present to omit the opsonic index in the treatment of staphylococcus infections, and this is at times also done in tubercle, gonococcus and streptococcus infections as well as in prophylactic typhoid inoculations.

### Neufeld's Method of Bacteriotropin Estimation.

Neufeld's technique varies from that of Wright in two points:

- (1) He uses serum free of complement.
- (2) He does not count the number of bacteria within the leucocytes; but makes various dilutions of the serum and notes in which dilution the bacteria are still ingested in great numbers, as compared with a normal serum in similar dilution as control.

Neufeld usually obtains the leucocytes by injecting a guinea-pig intraperitoneally 16 to 24 hours previously with 5 to 10 c.cm. of sterile aleuronat solution (1 part aleuronat, 2 parts bouillon). It is best to kill the guinea-pig, and wash out the peritoneal exudate which is full of leucocytes with



40 to 60 c.cm. of sterile salt solution. Occasionally it may be necessary to use 1 1/2 per cent. sodium citrate solution instead, in order to prevent coagulation. The leucocytes must be washed free of any serum. They should not however be centrifugalized too rapidly, as this tends to clump them.

If rabbit's leucocytes are preferred, 50 to 100 c.cm. of 3 per cent. to 10 per cent. peptone bouillon should be injected intraperitoneally. For mice 1 c.cm. aleuronat bouillon is sufficient. Human leucocytes are obtained from abscesses or from the blood (Wright).

The serum is inactivated by heating it at 50° to 60° C. for one-half hour. This may be omitted for old or carbolized sera as they are usually free of complement. Also tuberculous sera should not be heated as their bacteriotropins are very susceptible to heat.

The various serum dilutions (1 : 10, 1 : 100, 1 : 1000, etc.) are prepared as usual, but small quantities suffice since only 0.5 to 1.0 c.cm. of each dilution is necessary.

The bacteria are best employed in the form of a 16 to 24 hours homogeneous broth culture. If agar cultures are used, three loopfuls are rubbed up in 1 c.cm. of salt solution.

For meningococci Neufeld advises agar cultures. Tubercle bacilli can either be triturated in an agate mortar or bought in the form of tuberculin residuum, T. R. (Hoechst Farbwerke).

Equal parts (0.02 to 0.1 c.cm.) of each of the three ingredients (serum, bacteria, leucocytes) are transferred by means of a capillary pipette, to small tubes with flattened bottoms about 4 to 5 cm. long and 1 cm. wide. Double the quantity of leucocytes may be needed if they are in a weak suspension.

The tubes are closed by small corks or non-absorbent cotton, gently but thoroughly shaken and placed in the incubator for definite periods of time, depending upon the micro-organism, from one-fourth to four hours. The supernatant fluid is then pipetted off, cover-glass preparations made of the sediment, fixed by heat and stained by 1 per cent. methylene-blue solution.

A great number of fields are examined microscopically and note made of the weakest dilution which still favors phagocytosis. This is the bacteriotropic titer of the serum.

The necessary controls are: (1) Tube containing normal serum + bacteria + leucocytes. Phagocytosis of less virulent bacteria frequently occurs even with normal serum. (2) Tube containing bacteria + leucocytes, without any serum; a virulent bacteria are sometimes taken up by leucocytes even without serum. This is never the case with virulent organisms.

If phagocytosis is entirely absent, one should not conclude that bacteriotropins are not present. Errors in technique are possible:

(I) Leucocytes may have been injured; this is especially prone to occur if one has worked with heterologous leucocytes; control examinations with homologous leucocytes (from same animal as the serum) should result in phagocytosis.

(II) The serum concentration may be too high. The use of sensitized bacteria will obviate this. Bacteria are first mixed with the serum for a short time, the mixtures centrifugalized, and the serum pipetted off leaving a sediment of sensitized bacteria.

(III) The time during which the tubes were in the incubator may have been too short or too long. Most micro-organisms require one-half to two hours; pneumococci usually need four hours; cholera vibrios 20 to 30 minutes, as they undergo intracellular digestion very readily.

Neufeld's technique is considered by many simpler than Wright's method. In the determination of normal opsonins, however, concentrated or only slightly diluted sera are employed, thus encountering the difficulties mentioned above (II and III). Homologous leucocytes and sensitized bacteria will remove this interference.

The editor has found it puzzling at times to decide upon the dilution in which bacteriotropins still exist. One may be helped in this decision by the presence or absence of a great number of extracellular organisms.



## CHAPTER XVI.

### IMMUNITY AND SERUM REACTIONS IN REFERENCE TO MALIGNANT TUMORS. EXPERIMENTAL TRANSPLANTATION OF TUMORS. IMMUNITY TOWARD TUMORS. SERUM REACTIONS. MEIOSTAGMINE REACTION.

The etiology of malignant tumors is still unsolved. It has been definitely proven, however, that tumors or at least some of them can be transplanted. Naturally such experiments with human cancer are still limited and inconclusive, as attempts to transplant growths from one person to another are entirely out of question. On the other hand, the inoculation of human cancer into lower animals has been successful in the hands of only few reliable workers as Dagonet and C. Lewin.

These failures are the less surprising when one considers that malignant tumors obtained from the lower animals, as rats, and transplanted into other animals of the same species, not infrequently cease to grow in their new surroundings. Successful implantations of spontaneous animal tumors vary very greatly, from 4.1 per cent. (Bashford) to 40 to 50 per cent. (Jensen). The percentage becomes still lower or even entirely negative if the transplantation is made upon animals not of the same but of very closely related species, for example, when the gray house mouse is employed instead of the white mouse.

Once, however, the tumor continues to proliferate in its new host, it becomes more easily transplantable. Ehrlich has shown that the virulence of a tumor increases the more frequently it is successfully transplanted. Thus a growth can be obtained which may give 90 to 100 per cent. of positive grafts. During this long-continued process, the histological structure of the tumor may change. Not only may a carcinoma be transformed into an adenoma, but even into a sarcoma, as was first observed by Ehrlich and Apolant, and later on by others (Liepmann, Bashford, C. Lewin).

That an immunity toward carcinoma may possibly exist has been considered.

This has been based upon the facts that some animals resist all attempts at transplantation ("natural immunity"), that in others the tumor grows only to a limited degree, and that in a few, comparatively large tumors recede and disappear spontaneously ("acquired immunity"). The last class now remains refractory to all tumors, even the most virulent, thus proving a non-specific immunity ("pan immunity"). Ehrlich attempted to produce an immunity against a highly virulent mouse carcinoma by using as vaccine the hemorrhagic mouse tumor which only rarely allows of transplantation.

Further experiments proved that a certain degree of resistance in an animal may be attained by the injection of various cells, be they embryonal tissue elements or simply red blood cells.

Naturally all the known bacteriological methods for the destruction of bacteria have been applied to tumor cells; for example the addition of disinfectants, or the application of heat. Passive immunization with the serum from animals in whom the tumors disappeared spontaneously, or from rabbits treated for a long time with increasing quantities of an emulsion of carcinoma cells, also gave only doubtful results even though at times the experiments seemed encouraging. While it seems probable that the growth of neoplasms is attended by processes of immunity, one cannot directly compare this with the bacterial or proteid form of immunity and expect the same antibodies as given by the latter. In fact, all attempts at a serum diagnosis for carcinoma or sarcoma by the methods of precipitation or complement fixation have failed to withstand careful criticism. The reactions to be discussed will, however, tend to show that the sera of carcinoma patients possess certain characteristics which may play an important rôle in the future development of this problem.

"Brieger's cachexia reaction" has been reviewed in the chapter on anti-ferments. This test demonstrates that in a certain number of carcinoma patients the serum contains greater amounts of antitrypsin than normal sera. Similarly, many carcinoma sera have a stronger than normal hemolysin for the erythrocytes of the same or different animal species (iso- and heterolysins). Here too the results are inconstant and the sera of many noncarcinoma patients, especially tuberculous, give similar reactions. The same may be said of the test based upon the hemolysis of the carcinoma patients' red blood cells by cobra venom.

### The Freund Kaminer Reaction.

Freund and Kaminer found that if normal serum is mixed with an emulsion of carcinoma cells and allowed to remain at 40° C. for 24 hours, the latter are broken up and dissolved. This does not occur if the serum is derived from a carcinoma patient. The destruction of cells is determined by counting them in the Zeiss Thoma blood chamber both before and after the 24 hours' incubation; or one may take the clearing up of the turbid cell emulsion as an evidence of the cell destruction.

The technique of the reaction is as follows:

Carcinoma tissue rich in cellular elements and not degenerated is excised as soon after death as possible, cut up into small pieces and placed into five times as much of a 1 per cent. solution of sodium diphosphate. The whole is pressed through gauze and allowed to stand until the cells have sunk to the bottom, after which they are washed in 0.6 per cent. salt solution. They are again allowed to settle and then mixed with



1 per cent. solution of sodium fluoride. The sodium fluoride solution is first neutralized (alizarin as indicator) until the violet color is reduced to its minimum. The cell emulsion may be thus preserved for several weeks.

In the test, the carcinoma extract is diluted with 0.6 per cent. sodium chloride solution until it becomes opalescent. To 3 c.cm. of this emulsion are added 10 drops of the patient's serum preferably fresh and active. They are allowed to remain at 40° C. for 24 hours and the test is positive if the turbidity persists. Control tubes must be made of the serum alone, carcinoma extract without serum, and carcinoma extract with normal serum. The last should become clear.

Recently Freund and Kaminer proposed the following more delicate modification. The supernatant fluid from the emulsion of carcinoma cells is mixed with acetic acid (5 c.cm. of acid to 100 c.cm. of fluid) heated in the water bath for one-quarter of an hour at 80° C., filtered, and after cooling neutralized with sodium bicarbonate against litmus. Then it should again be heated and filtered. Boiling or heating over the free flame is to be avoided. An extract can also be made by heating the tumor itself (preserved in alcohol) in 0.25 per cent. of acetic acid, then filtering and neutralizing. Serum of cancerous individuals added to this extract produces a cloudiness; non-cancerous serum and extract remains clear. Freund and Kaminer advise that both the cell-counting method and the turbidity reaction should be applied to each serum, one acting as a control upon the other.

Ranzi and Admiradzibi, Kraus and Graff have corroborated Freund and Kaminer's findings. Rosenberg working under Citron's guidance has found that although in the main the reaction is obtained as above quoted, some carcinoma sera give a negative reaction and some non-carcinoma patients (tuberculosis, pregnancy) give a positive result.

### The Meistagmine Reaction.

Weichard showed that by bringing together antigen and antibodies in certain dilutions, the rapidity of diffusion is increased (epiphanin reaction). M. Ascoli further demonstrated that the union between a specific antigen and its specific serum is associated with appreciable lowering in the surface tension, so that the number of drops to a definite quantity of fluid is distinctly increased (Meistagmine reaction). This term is of Greek derivation, "*μείων*," smaller, "*στάγμα*," drops. Traube's "Stalagmometer"<sup>1</sup> measures the number of drops.

The meistagmine reaction has been tested by Izar and Vigano in typhoid, paratyphoid and lues and found to possess a certain degree of specificity. In tuberculosis, the test is positive only in active cases. Ascoli and Izar claim to get reliable results with their method also in carcinoma. Their technique is as follows:

<sup>1</sup> The stalagmometer of Traube is merely a very finely and elaborately graduated pipette with a central bulbous reservoir. The dropping end of the instrument ends in a flattened ground base thus insuring uniformity in the size of the drops. The instrument is so graduated that a fraction of a drop can be estimated.

1. *Extract*: a malignant tumor which has not undergone degeneration (or a sheep's pancreas) is cut into fine slices or small pieces. It is spread out on glass plates and dried in a temperature of  $37^{\circ}\text{C}$ . The dried residue is powdered and extracted with methyl alcohol in the proportion 1 : 4 for 24 hours at  $50^{\circ}\text{C}$ . The extract is filtered through a hard filter, at first hot and then cold. It should be protected from the light, but should not be placed in the ice-box. The dilutions cannot be preserved, but must be made fresh every time, by measuring out the requisite amount of extract with a dry pipette, placing it in a dry test-tube or flask, and adding the required quantity of distilled water and shaking thoroughly.

The extract is first titrated: dilutions 1 : 50, 1 : 75, 1 : 100, 1 : 125, etc., are made, and to 1 c.cm. of each of these, 9 c.cm. of a 1 : 20 normal serum diluted with saline are added. The mixtures are placed for two hours at  $37^{\circ}\text{C}$ . The number of drops to each quantity is estimated. A control tube containing the same quantity of normal serum plus 1 c.cm. of distilled water is also counted. That dilution is chosen as the working dose which differs from the control tube by 3 to 5 fractions of a drop.

The test is performed as follows: To 9 c.cm. of each serum diluted with saline 1 : 20 is added 1 c.cm. of distilled water and the number of drops falling from the stalagmometer is counted before and after incubating for two hours at  $37^{\circ}\text{C}$ . or one hour at  $50^{\circ}\text{C}$ . This is used as a control. To 9 c.cm. of each serum diluted 1 : 20 with normal saline is added 1 c.cm. of antigen emulsion of the desired strength, e.g., dilution 1 : 125. The number of drops in this mixture is counted only after incubation. As further controls it is advisable to have several known normal and tumor sera. The reaction is positive if the number of drops contained in the mixture of tumor extract and tumor serum is greater by at least  $1\frac{1}{2}$  drops than in the controls after the incubation.

9 c.cm. serum 1 / 20	Number of drops per c.cm. with 1 c.cm. tumor extract 1 : 125 (after incubation)	Number of drops per c.cm. with 1 c.cm. of distilled water (after incubation)
Normal serum.....	58+4 <sup>1</sup>	58+2
Normal serum.....	59	58+7
Carcinoma serum.....	61+6	58+8
Carcinoma serum.....	60+4	58+5

The antigens spoil very easily. Recently, chemicals in the form of lipoids are being substituted for the specific antigen. This simplifies matters greatly. In looking over the numerous publications by Ascoli and his

<sup>1</sup>58+4 means 58 drops + 4 fractions of a drop.



followers, one meets with constant variations in the technique of the reaction. Thus it is very difficult to arrive at an exact description or a conclusive opinion. Many non-cancerous sera give a positive meiostagmine reaction. The test has not yet been adopted as a clinical aid. A review of the early literature of this subject is given by Bernstein and Simons, *Amer. Jour. Med. Sciences*, Dec., 1911.

## CHAPTER XVII.

### ANAPHYLAXIS.

*Introduction. Proteid Hypersusceptibility. Anaphylaxis. Passive Anaphylaxis. Anaphylatoxin. Serum Disease. Hay Fever.*

The explanation for many phenomena discussed in the former chapters was based upon the observation that by overcoming an infectious disease the organism undergoes some transformation whereby it becomes refractory or less susceptible to the same infection. This changed state, demonstrated neither by chemical nor microscopical methods, was termed "Immunity." The most constant evidence of immunity is the presence of antibodies. This, however, should by no means imply, as is so often done in literature, that the demonstration of antibodies and the existence of immunity are identical.

On the contrary, attention has already been called to the possibility of paradoxical reactions. Animals or individuals in whom a grade of immunity is expected, show instead a certain susceptibility toward the particular antigen. This altered reaction of the organism was termed "Hypersusceptibility" and "Anaphylaxis" (to distinguish it from prophylaxis-immunity). The "paradoxical reaction" was discussed under diphtheria where it was mentioned that horses immunized for a long period of time would suddenly become severely ill after the injection of small doses of toxin. The tuberculin reaction was another instance of such hypersensitiveness. Infection by the tubercle bacillus not only made the body highly sensitive toward the bacteria and their derivatives (tuberculin), but a severe and characteristic reaction always took place as the expression of this increased sensitiveness.

In recent years, experimental studies have proven that this peculiar phenomenon is not based upon entirely new laws, but that anaphylaxis has as its governing influences principles that are closely related to those influencing the process of immunity. It is still to be determined whether the hypersusceptibility observed with various antigens like the pure toxins, the tuberculins, and the proteids all follow the same biological mechanism. The author is of the opinion that just as there are various forms of immunity so also must the existence of various forms of anaphylaxis be presumed. At least a cellular and humoral variety seem distinctly plausible; the tuberculin reaction is an example of the former, the proteid anaphylaxis of the latter.



The parenteral introduction of a foreign proteid into an organism results in the formation of antibodies against this Proteid. Anaphylaxis. proteid: precipitins, cytolytins, complement fixation bodies, etc. At first it was assumed that a "proteid immunity" had taken place. Such conclusion proved erroneous.

Richet and Portiers (1902) in working with actino-congestin (a substance extracted from the tentacles of Actinia) demonstrated that dogs injected with sublethal doses of this toxin would die acutely if the injection were repeated after an interval of three weeks. In the light of more recently established facts, the explanation of Richet's work was complicated because actino-congestin consists of two components, a true toxin against which an immunity can be stimulated, united with a proteid element which like all proteids produces anaphylaxis.

It was therefore of extreme fundamental importance when Arthus and Theobald Smith showed that proteid substances, of themselves apparently non-toxic, can constantly produce the phenomena associated with hypersensitiveness.

### The Arthus Phenomenon.

If a rabbit is injected subcutaneously with horse's serum at intervals of six days, a soft infiltrate which remains for two to three days appears at the site of injection after the fourth inoculation, a harder infiltration which continues for a longer period of time after the fifth inoculation, and gangrene after the sixth or seventh. A rabbit repeatedly treated intravenously with horse's serum may die with severe general symptoms several minutes after one of the later inoculations.

### The Theobald Smith Phenomenon.

Theobald Smith observed that guinea-pigs injected with neutral mixtures of diphtheria toxin and horse's antitoxic serum would be killed if after an interval of several weeks they were given a subcutaneous injection of normal horse's serum.

Otto and others showed that both of these phenomena, above described, are identical in their principle; thus, that of Arthus can be likewise induced after a single injection of horse's serum if the first dose is small, and if the interval between the first and second inoculation is sufficiently long (about three weeks or more).

*Further study proved that anaphylaxis may be incited by repeated parenteral introduction of practically any foreign proteid.*

The first inoculation prepares the animal in such a manner that after a definite incubation period the second injection of the same serum will

bring on characteristic acute symptoms which may terminate fatally. The picture of hypersensitiveness or "serum sickness" in man, as described by v. Pirquet and Schick, is classical. Other investigators who deserve merit for work in this field are Arthus, Otto, Rosenau, Anderson, Kraus, Doerr, Besredka, Weichardt, Wolff-Eisner, Friedemann, Friedberger, H. Pfeiffer, Schittenhelm.

It is through their efforts that the close relationship between hypersensitiveness and immunity is more clearly understood. Like the state of immunity, anaphylaxis is either spontaneous or acquired. It is also specific, that is a guinea-pig made sensitive toward horse's serum will react only when again treated with horse's serum but not when receiving rabbit's or human serum.

A classical anaphylaxis experiment may be carried out as follows. A guinea-pig receives a subcutaneous injection of 0.001 to 0.01 c.cm. of horse serum and after three weeks an intravenous injection of 3 to 5 c.cm. is repeated. This quantity, which under normal conditions has no influence upon the animal, will now produce very alarming symptoms or even death in a couple of minutes.

There are several factors upon which the occurrence of the anaphylactic phenomena strictly depends.

(a) The first, preparatory, or sensitizing dose must enter the system in some way other than through the gastro-intestinal tract. Only exceptionally does hypersensitiveness arise if the antigens are given per os.

(b) The quantity of antigen is of the utmost importance. The smallest amounts of proteid suffice, *e.g.*, 0.000001 c.cm. serum. As a rule 0.001 to 0.01 c.cm. are employed.

(c) An incubation period (the preanaphylactic state) is always necessary. This period varies in the widest degree, but depends primarily upon the amount of antigen first injected.

With guinea-pigs, it is never less than seven days. If 0.01 to 0.1 c.cm. serum is injected, symptoms may usually be stimulated after ten days. Very large doses as well as very minute ones increase the length of the preanaphylactic state very markedly (as long as three months).

(d) The susceptibility of the various animal species differs greatly. The most suitable is the guinea-pig, the rabbit far less so.

(e) The actual anaphylactic shock is dependent upon the quantity of the second dose injected and the mode of injection. The intravenous path of a massive dose is the most reliable; then comes the intraperitoneal and then the subcutaneous method. While with the intravenous procedure the final result is usually death, this is almost never so with the subcutaneous injection. Here the anaphylaxis expresses itself in a more marked local inflammation, edema and eventually necrosis.



An animal that recovers from the second injection becomes immune to further administration of the same proteid. This non-susceptible condition has been wrongly termed antianaphylaxis. It is much more reasonable to speak of "anaphylaxis," since the absence of hypersensitiveness is not due to the neutralization of the bodies necessary for bringing about the hypersusceptibility. The anaphylactic state sets in as early as two hours after the anaphylactic outburst.

In order to prevent the shock of anaphylaxis, it has been suggested by Besredka and Steinhardt to give the second injection during the period of incubation, *i.e.*, about the eighth day, or give a very minute dose of serum at the regular time of the second inoculation and give the larger dose in 24 hours.

Just as an immunity may be transmitted by injecting the serum obtained from an immune animal, so also can the tendency to hypersusceptibility be transmitted by introducing into a normal animal the serum from a sensitized one, *i.e.*, one that has been injected with a foreign proteid. This is best demonstrated by injecting the anaphylactic serum subcutaneously, followed in 24 hours by the intravenous inoculation of the respective antigen.

A fully satisfactory explanation for all the phenomena of anaphylaxis has not as yet been advanced. Certain it is that a number of underlying factors exist which bring anaphylaxis and immunity into close relationship.

Since the term immunization usually implies a beneficial process, while anaphylaxis in most instances represents a situation of an injurious nature, v. Pirquet recommended the term "allergie" to designate the reactive changes which an organism generally exhibits after infection or injection of an antigen. The allergic phenomena are divided into those associated with diminished susceptibility, *i.e.*, prophylaxis, and those with increased sensitiveness, *i.e.*, anaphylaxis.

Besredka adheres to the view that the anaphylactic syndrome expresses an insult to the central nervous system. He was able to show that susceptible guinea-pigs, when etherized, will bear the second inoculation of the serum perfectly well. V. Pirquet and Schick consider the precipitin action as the basis for the anaphylactic phenomena. In the main, however, there are two theories, a cellular and a humoral one. The former suggests that the hypersusceptibility is due to the stimulation of new specific receptors which remain sessile, *i.e.*, attached to the body cells instead of being thrown off into the blood stream. When more antigen is injected, these cells, due to their greater affinity, absorb more of the toxic substance of the antigen than they do normally, and thus the anaphylactic shock is incited.

The humoral theory represents the main activity within the serum. This hypothesis was adopted by Wolff-Eisner, a pupil of Pfeiffer, and was based upon Pfeiffer's endotoxin principle. It takes for granted that all antigens, cells and proteids, contain

within their bodies a toxic substance that does not form antibodies when liberated in animals. The first injection of antigen produces bacteriolytic or cytolytic antibodies possessing the power of liberating the endotoxic poisons from the proteid molecule. When the second injection is given, these bacteriolytic antibodies at once cause a rapid liberation of the intracellular toxic fraction, and injury to the animal results. Wolff-Eisner's theory can apply only in a certain number of instances, because the essential factor of cytotoxicity is not always present; very many bacteria, especially tubercle bacilli, are not thus broken up.

This view has, however, been the fundamental thought for the later very important work of Friedemann and Friedberger. By these supporters of the humoral theory the main influence is placed upon the union which takes place *in vivo* between the antigen + amboceptor + complement. As a result, anaphylaxis may be incited in one of two ways. The mere absorption of the complement may bring about the anaphylactic shock. This diminution in the complement content of the serum is always demonstrable. Or, as is more probable, the union of the above three elements causes a destruction of the antigen and a liberation of a toxic agent. This theory is strengthened by the demonstration that not only *in vivo* but also *in vitro* can such a toxic substance be obtained by mixing antigen + amboceptor + complement.

Anaphylatoxin. In 1902 Weichardt immunized rabbits with a proteid derived from syncytial cells. By mixing the fresh immune serum with an emulsion of the syncytial cells and filtering, he derived a very toxic fluid which was named syncytiotoxin. In 1909 Friedemann and Citron simultaneously isolated *in vitro* toxic elements which could bring about anaphylaxis. This was confirmed by Friedberger several months later. Friedemann got his poisonous agent by mixing erythrocytes, inactive hemolytic serum and complement.

Three c.cm. sheep's blood + 1 c.cm. inactive hemolytic serum are allowed to remain at room temperature for one-half hour, then centrifugalized. The sensitized erythrocytes are mixed with 4 c.cm. fresh rabbit's serum (complement) and placed into the water bath at 37.5° C. for exactly five minutes; then cooled down by ice and centrifugalized. The red blood cells remain almost all unhemolyzed. The supernatant fluid is only slightly reddened and when injected intravenously into a rabbit (1310 gms.) causes general weakness, diarrhea and death on the following day.

It is to be observed that the anaphylatoxic substance was formed even though no hemolysis took place.

Citron accomplished the same end by a mixture of tuberculin, serum from a tuberculous individual who spontaneously produced antituberculin amboceptors, and normal guinea-pig's serum (complement).

0.1 to 0.2 c.cm. T. + 0.2 to 0.4 c.cm. antituberculin + 0.3 to 0.5 c.cm. complement are mixed and incubated for one hour. The entire quantity is injected intraperitoneally into tuberculous guinea-pigs; the animals die in four to five hours. Tuberculous animals receiving 0.1 to 0.2 c.cm. T. alone or 0.1 to 0.2 c.cm. T. + 0.2 to 0.4 c.cm. antituberculin, remain alive.



Friedberger demonstrated that a toxic product which he named "Anaphylatoxin" can be obtained *in vitro* from every proteid, be it animal or bacterial in nature. His technique is almost identical with that of Friedemann described above.

Accordingly, Friedberger proposed a theory explaining the course and nature of all infectious diseases upon the basis of such a common anaphylatoxin, independent of the character and virulence of the bacteria. While this hypothesis takes into consideration the older belief that disease is due to a struggle between the infectious agent and the invaded organism, it does so in a somewhat broader sense. From Friedberger's standpoint it is the unspecific anaphylatoxin which is the important factor; its variable quantity, which stands in direct association with the variable proportions of antigen, amboceptor and complement, accounts for the various pictures of the different bacterial infections.

This explanation hardly suffices for all the characteristic symptoms of the infectious diseases. It does, however, account for one, the fever.

That some relationship exists between hypersusceptibility and the thermal center was first observed by H. Pfeiffer and Mita. The anaphylactic shock is always associated with a fall in temperature, and at times this may be its only manifestation.

A different effect upon the temperature results according to the different quantity of anaphylatoxin injected (Schittenhelm and Weichardt, Friedberger and Mita). Well-defined limits have been established.

Aside from the effect upon the temperature, other characteristic phenomena of the anaphylactic intoxication are the incoagulability of the blood and the marked distention of the lungs as found by post mortem examination.

The symptoms of active anaphylaxis differ in the various animals. In man very exact studies of the results of the repeated injections of foreign serum have been made by V. Pirquet and Schick.

The evidences of serum sickness are numerous. Those which are present most frequently are fever, skin eruptions, swelling of the joints, glandular enlargement and edema.

These symptoms may follow even the very first injection of serum. As a rule they develop after an incubation period of eight to ten days. Slight reddening at the point of injection accompanied by moderate swelling of the regional lymph glands appear as prodromal manifestations.

The general condition of the patient is generally only very little disturbed in spite of the frequently associated high fever. Still there are instances, especially after the introduction of large amounts of serum, where the symptoms continue for about four to five weeks and then lead to severe disturbances.

The associated skin eruptions are usually of the type of an urticaria; although Hartung describes rashes simulating scarlet fever and measles.

V. Pirquet and Schick consider the following as the most positive symptoms of serum sickness:

1. The occurrence of the exanthema seven to fourteen days after injection.

2. First appearance of the rash around the point of injection.

3. Regional enlargement of the lymph glands.

4. Complete absence of any changes in the mucous membrane.

Measles is excluded by the absence of Koplik spots, coryza, and conjunctivitis.

In scarlet fever the following symptoms help to exclude serum sickness:

1. Initial vomiting.

2. Occurrence of angina before or at the same time as the exanthema.

3. High fever.

4. The simultaneous existence of the infection among others in the hospital or neighborhood.

If the serum disease does not arise after the first, but after a later injection, it is characterized by the absence of, or very marked diminution in, the length of the period of incubation, and in addition by increased severity of the symptoms.

In the dog these phenomena have been carefully observed by Anaphylaxis Biedl and Kraus and Arthus: 3 to 5 c.cm. of horse serum is in the Dog. administered for the first injection and after three weeks 10

c.cm. are given intravenously. In about one-half a minute the dog becomes restless, begins to vomit, and has involuntary evacuation of urine and feces. This is followed by a period of excessive prostration during which the dog lies with his limbs outstretched and almost motionless as if paralyzed. After several hours the animal either begins to recover or dies.

Biedle and Kraus noticed that about 15 to 30 seconds after the intravenous reinjection, the arterial blood pressure begins to sink rapidly. This has been ascribed to the wide dilatation of the peripheral blood vessels due to a paralysis of the peripheral vasomotor system. In addition there is a leucopenia and a diminution in the coagulation power of the blood, symptoms which have been observed also in man. The animal par excellence for anaphylaxis experiments is the guinea-pig. The rabbit is the next choice.

If guinea-pigs receive as their second inoculation a large dose of serum in Anaphylaxis travenously, they die acutely (Th. Smith); (animals that have not been in the sensitized bear this same quantity of serum without any disturbance Guinea-pig whatsoever). The blood pressure first rises, then sinks rapidly. At and Rabbit. postmortem the lungs are firmly inflated (Gay and Southardt).

Death is probably caused by respiratory paralysis, Auer and Lewis having described a tetanic spasm of the bronchial muscles.



If for the second injection only a small dose of serum is administered, or it is introduced intraperitoneally instead of intravenously, acute death does not result and the picture is similar to that given by the anaphylactic dog (Pfeiffer and Mita). The guinea-pigs become very restless, move about continuously and are very easily frightened. Their hair becomes raised, and isolated clonic muscular contractions may be observed. Continuous hiccough sets in and evacuation of urine and feces, first formed and later fluid occurs. The abdominal muscles become spastically rigid and at times a severe pruritus of the skin probably exists. Following this transient period of excitation the animals enter into a stage of depression. They stagger about or fall to one side and remain for hours as if paralyzed; breathing is slow and superficial. In the midst of these symptoms the temperature falls abruptly, often as many as  $7^{\circ}$  to  $13^{\circ}$  C. Death is due to a paralysis of the peripheral blood-vessels and takes place usually in one to two hours, sometimes four to eight hours, being always preceded by Cheyne-Stokes respiration.

The symptoms of the anaphylactic shock simulate very closely those occasioned by peptone poisoning, as seen after intravenous injection of Witte's peptone, for example. Biedl and Kraus thus considered that anaphylaxis was a manifestation of peptone poisoning. They believed that through immunization with proteids, antibodies of a ferment nature were stimulated, and that these split up the proteids into peptones. According to these authors, animals exposed to peptone poisoning become hypersensitive, and vice versa, animals that recover from an anaphylactic attack withstand poisoning by peptones. The production of anaphylatoxin *in vitro*, and the demonstration by Pfeiffer that peptone is formed in the test-tube during this process, further added to the support of their theory.

This hypothesis cannot as yet be definitely accepted. In Toxopeptid. the first place Manwaring was unable to confirm the experiment that recovery of an animal from an anaphylactic shock renders it refractory toward future peptone poisoning. Furthermore, M. Wassermann and Keysser mixed kaolin + inactive immune serum + complement and obtained a poison which caused the same disturbances as anaphylatoxin. Since kaolin is no proteid and cannot be split up, the anaphylactic symptoms in this instance cannot be due to a peptone as a split product of the antigen.

M. Wassermann and Keysser have a different conception of the nature of anaphylaxis. The antigen serves in a physical chemical capacity to fix the amboceptor. By the addition of complement the amboceptor is broken up with the formation of "Toxo-peptids," and it is these which stimulate the anaphylactic phenomena. Passive anaphylaxis is explained by the passive transmission of the specific amboceptors; the antigen itself plays no chemical rôle; it is not split up. The difference between anaphylaxis and immunity lies in that in the former the complement under cover of the antigen digests the amboceptor alone, while in the state of immunity the strength and number of amboceptors are very much greater and the activity of the complement extends not only to the amboceptor but also to the secondarily attached antigen.

It would be impractical to discuss all the theories enlisted for anaphylaxis. Exact facts are still insufficient. Experimental work is constantly disclosing new ideas.

More detailed reviews may be found in the following articles:

H. Pfeiffer: Problem of Proteid Anaphylaxis. G. Fischer, Jena, 1910.

A. Schittenhelm: Anaphylaxis from the standpoint of pathological physiology and the clinic. Jahresbericht über die Ergebnisse der Immunitätsforschung. Enke, Stuttgart, 1910.

J. Citron: Critical review of the problem of anaphylaxis. Fol. serologica, 1911, Bd. vii, H. 3.

Friedberger: Anaphylaxis. Fortschritte der deutschen Klinik. Bd. ii. Urban u. Schwarzenberg, Berlin-Wien, 1911.

H. Dold: Bacterial Anaphylatoxin and its Importance in Infections. G. Fischer, Jena, 1912.

Besides bacterial infections, recent teaching places urticaria, Hay-fever, eclampsia, bronchial asthma and hay-fever into the class of diseases with an anaphylactic basis. This is especially applicable to hay fever, formerly considered a pure intoxication, the pollen toxin having been described by Dunbar as the etiological factor. In Germany the disease seems to come chiefly from pollen of the grasses and grains (rye pollen being most active); whereas in America, apparently, the most important pollen springs from the ambrosia (rag weed), solidago (golden rod), and other members of the family of the *compositæ*.

The toxin is isolated by mixing for ten hours the ground pollen with 5 per cent. NaCl solution and 0.5 per cent. phenol at 37° C. Then, in the form of a proteid it is precipitated by the addition of eight to ten volumes of 96 per cent. alcohol and the resultant white precipitate dissolved in physiological salt solution.

Susceptibility to the pollen toxin is limited only to certain individuals. Some are influenced by the rye pollen only, others by the golden rod alone, while a third class is affected by all. The cause for this peculiar idiosyncrasy is unknown. The majority of observers are, however, now agreed that one is dealing here with a reaction of hypersusceptibility, as was first pointed out by Weichardt and Wolff-Eisner. Only by means of antibodies does this non-toxic pollen proteid become a poison.

All those who suffer from hay-fever develop a marked conjunctivitis whenever even the slightest amount of pollen proteid (1/1000 mg.) is dropped into the conjunctival sac. In addition, all the symptoms of hay-fever or asthma may be incited. Similar effects are in evidence when subcutaneous injections are resorted to.

For purposes of immunization horses are most suitable, but only those which after an injection of pollen extracts manifest a local and general re-



action. This is found in one-third of the animals. Their serum thus rendered immune is capable of neutralizing all effects of the pollen toxin.

As regards the standardization of this serum, it is effected by mixing the *dosis minima certe efficax* of the toxin with various dilutions of the serum and instilling the mixture into the conjunctival sac of individuals with a tendency for hay-fever. That amount of the serum which suffices to neutralize the toxic action is taken as the unit of measure. Sera of at least thirty times the unit strength are selected for therapeutic application.

The immune serum is manufactured in fluid and powder form, and is placed on the market under the name of "Pollantin." Its use is mainly local, by spraying a small quantity of the pollen powder upon the nasal mucosa several times daily and by placing several granules into the conjunctival sac with a camel's-hair brush. The serum can also be employed as a prophylactic.

If the eyes are especially reddened, it is best to deposit some fluid serum into the conjunctival sac every day. Prausnitz advises the injection of 1 to 2 c.cm. of the serum subcutaneously when asthmatic attacks occur, when the above local treatment has failed.

In America a special Pollantin is made against the frequent form of hay-fever known as "autumn catarrh" by immunization with the pollen of the golden rod and rag weed.

The pollantin therapy and prophylaxis has been quite satisfactory, inasmuch as two-thirds of the patients remain either entirely free from attacks or are so greatly benefited that their general duties are not interfered with.

The specificity of anaphylaxis has been proposed for diagnostic aid. As yet the methods have not been perfected sufficiently to be of clinical value.

## CHAPTER XVIII.

### PASSIVE IMMUNIZATION (SERUM THERAPY). BACTERIOLYTIC SERA. SPECIAL SERUM THERAPY.

In the former chapters it was proven that during *active immunization* of an animal specific protective bodies were formed which circulate in the blood and can by means of the serum be transferred to another organism. It was further found that such bodies exert this protection against fatal intoxication or infection in various ways; thus, as antitoxins and anti-aggressins they neutralize toxic poisons and aggressins; as bacteriolysins they bring about lysis of the bacteria; while as bacteriotropins they prepare the bacteria for phagocytosis. The defensive qualities of such a transferred serum are evident not only if the infection is incited at the same time as, or a short period after the serum is given, but in numerous instances curative effects are observed if the serum is given even after infection has already taken place.

Of all sera, those with *antitoxic properties* have met the greatest success in therapeutic application. They have already been referred to in their respective chapters.

*The efficiency of the pure bacteriolytic sera on the other hand has been disappointing.* One reason given for this lack of curative action is, *the inability of bacteriolytic serum to neutralize the endotoxins.*

Pfeiffer's experiment revealed that if the number of bacteria exceeded a certain limit, then in spite of bacteriolysis, death of the animal takes place. This was explained by the existence of endotoxins. By bacteriolysis the endotoxins normally confined within the bacteria are liberated and thus get a chance to show their toxicity.

The aim, therefore, was to produce antiendotoxic sera. The accomplishment of this was prevented by the erroneous view of Wolff-Eisner who claimed that it was impossible to immunize against endotoxin.

Numerous methods have been advocated for the liberation of these endotoxins: maceration of bacteria, exposure to very low temperature, admixture with chemical substances which would dissolve the outer capsule, ferment digestion, growth upon certain culture media, etc. At the present day, there is absolutely no doubt that the bacterial bodies contain poisonous substances against which it is difficult and to a certain degree impossible to attain an immunity.

Whether one should adhere to the old idea and apply to these the term endotoxin, or include them in the class of true toxins with the only difference that they are not secreted but contained within the bacterial body and therefore more difficult to isolate, is purely a question of theoretical importance.

Another cause for the therapeutic failure of bacteriolytic serum, as ad-



vocated by Bail and his school, is the lack of its antiaggressin action. This applies only to the cases in which the bacteriolytic serum was produced by immunization with dead bacteria.

When live bacteria are used, this objection is not to be considered, as according to the experiments of Wassermann and Citron, "aggressin" is nothing more than the immunizing substance of the living bacteria. As far as the structure of the antiaggressins is concerned, the author was able to show that like the bacteriolysins, they are amboceptors which bind complement.

Artificial aqueous extracts of living bacteria, belonging to the class of half parasites made according to the method of Wassermann and Citron, contain the endotoxin as well as the aggressin. Such artificial aggressins, therefore, represent ideal antigens. The sera produced by their injection contain but few bacteriolytic bodies and a very large number of amboceptors, easily demonstrable by the Bordet-Gengou reaction.

Wassermann explains the lack of therapeutic efficiency on the part of the bacteriolytic sera by the absence of complement of the organisms, as well as by the inability of human complement invariably to fit animal amboceptors. As is known, amboceptors increase during immunization while the complement content remains the same. But since amboceptors without complement remain inactive, even a very strong serum may only be slightly effective, depending upon the amount of existing complement. If too many amboceptors are injected, the serum may become entirely powerless due to a phenomenon similar to Neisser and Wechsberg's complement deviation. Wassermann advises therefore the addition of complement to a serum before its injection, in order to activate it. This suggestion has not been widely adopted in practice.

It is for a similar reason, that the classical experiment of bacteriolysis is so beautifully demonstrable in the guinea-pig's peritoneal cavity, an area relatively poor in cells, while this phenomenon is incomplete and replaced by phagocytosis when occurring in the blood, inner organs, and subcutaneous connective tissue. It is in this connection that Metchnikoff and his followers see the main reason for the failure of the therapeutic activity of bacteriolytic sera.

An additional impediment is offered by the wide differences which exist among the numerous strains of the same bacterium. This may be so marked that an immune serum produced with one strain will offer no protection against a different strain of the same bacterium. It is now overcome to a certain extent by immunization with as many different strains of the same bacterium as possible (polyvalent sera).

Cultures grown upon artificial media for a very long time adapt themselves to their new surroundings and frequently lose some of their biological characteristics, *e.g.*, virulence. If the culture is then inoculated

into an animal, the virulence is increased usually only for that animal species, but may remain the same or even lowered for man. Many authors, therefore, employ for the production of immune sera only virulent strains of bacteria freshly isolated from a human being.

In spite of all the above considerations, the fact still remains that most immune sera excepting those of the cholera, typhoid, and paratyphoid bacteria, show no bacteriolytic tendencies even under the most favorable circumstances; but by means of their amboceptors they fix free complement and with the aid of bacteriotropins, stimulate phagocytosis.

Whether complement fixation is at all to be considered as a protective phenomenon, cannot with the evidence existing at present be definitely decided.

Conditions are much more favorable as far as the bacteriotropins are concerned. Active phagocytosis is always an expression of good resistance. It is not necessary for the leucocytes to digest the bacteria; it is amply sufficient if a protective wall of these cells is formed (Ribbert, Citron, Gruber); moreover they can neutralize the bacterial poisons. In this connection it must always be borne in mind that phagocytosis by no means necessitates the death of bacteria.

Granting, however, that all the above requirements have been fulfilled and a suitable serum has actually been produced, will such a serum always be effective, or are there any other causes which may interfere with its good results? In order to answer this, the infectious diseases must be divided into acute and chronic. With the first class, success is quite assured as long as it is possible to bring sufficient amounts of the active serum substances into direct contact with the bacteria. In meningeal infections, subdural injections may have to be adopted. It is difficult, however, in cases of this nature to judge definitely whether the serum therapy was really the effective agent, inasmuch as diseases like erysipelas, meningitis, pneumonia, etc., are self limited, lasting for a period of time and then subsiding of their own accord.

With the chronic infections, on the other hand (especially tuberculosis), serum therapy has a new difficulty to overcome. As a result of the long course of the disease, it is naturally impossible by means of a single injection to introduce sufficient curative bodies, as can be done in diphtheria, for example. It is necessary, therefore, frequently to repeat the injections. Under such conditions the human organism produces antibodies against the foreign proteid (anaphylaxis), perhaps even against the curative substances in the serum (antiamboceptors). In both instances the desired effect of the serum is lost. A further impediment lies in the possibility that bacteria remaining in a system for a long period immunize themselves, and thus resist the action of the antibodies directed against them. Such bacterial strains are known as "serum fast."



## Special Serum Therapy.

1. *Meningococcus Serum*.—Numerous investigators have attempted the production of an immune serum for man, among these Jochmann, the Berlin Institute for infectious diseases, Ruppel, Kraus, Flexner and Jobling, and others. The sera of Jochmann (Merck) and Ruppel (Höchst) are produced by immunization of horses with meningococci which are at first employed in dead, and later in live form. The other sera mentioned are obtained by immunization with bacterial extracts or bacterial extracts plus full bacteria, and therefore contain agglutinins, precipitins, bacteriotropins, amboceptors and antiendotoxins. It is difficult to test the efficiency of these sera in animals, as the meningococci vary greatly in their virulence toward them. Jochmann and Ruppel assert that they have been successful in growing cultures extremely virulent for animals which they employed for the titration of the therapeutic value of the serum. In the institute for infectious diseases, the method of complement fixation is taken as the index of the therapeutic value of the serum. This procedure is very unreliable. The protection of the serum in mice against the meningococcus endotoxin as well as the demonstration of the bacteriotropic action of the serum is far more significant.

In man, the immune serum is injected subdurally, after a quantity of spinal fluid has been withdrawn to relieve the pressure. In adults 20 to 40 c.cm. and in children 10 to 20 c.cm. are daily injected until there is clinical improvement or a fatal prognosis becomes inevitable. It is advisable to precede the serum inoculation by a morphine injection, and to elevate the pelvis for eight to twelve hours after the inoculation. *The earlier the serum therapy is instituted, the more favorable are its results.* Subcutaneous applications of the serum or employment of a serum more than three months old is absolutely of no use.

Both in the United States and in foreign countries the value of the serum as a therapeutic agent seems fairly established. In Germany, the serum is obtained gratis at the institute for infectious diseases at Berlin. The serum in Switzerland is distributed by the serum institute of Bern (Kolle). In the United States, the Rockefeller Institute of New York first conducted its dispensation, but now it is under the supervision of the New York Board of Health.

Numerous statistics can be cited exemplifying the good results of the serum. The following figures given by Levy describing the experiences in the Essen epidemic are especially instructive:

From the first of January until the first of November, 1907, the total number of epidemic meningitis cases which occurred in Essen were:

55 cases with 29 deaths = 52.72% mortality,

of these, treatment was given outside of the barracks to

15 cases with 12 deaths = 80% mortality,

inside the barracks were treated

40 cases with 17 deaths = 42.5% mortality,

of these

14 cases were not treated with serum with 11 deaths as a result  
= 78.6% mortality,

those treated with serum were

23 cases with 5 deaths = 21.7% mortality,  
of these, those which were treated only incompletely (subcutaneously) and with insufficient doses, numbered

6 cases with 3 deaths as the outcome = 50% mortality,  
systematic subdural treatment with large doses.

17 cases with 2 (1) deaths = 11.8 (6.3)% mortality.

The figures in parentheses represent the moribund cases coming under treatment and the percentage which would result if these were not included in the calculation.

The experiences with the serum of Flexner and Jobling are similarly encouraging. In a report of 400 cases (1909) the mortality is reported as lowered from 80 per cent. to 20 per cent.

In a more recent report (1913), 1,294 patients treated with serum show the following results: recovered 894; died 400 (30.9 per cent.). Flexner's tables which are here reproduced are instructive.

#### MORTALITY ACCORDING TO THE PERIOD OF INJECTION OF THE SERUM.

Period of Injection	No. of cases	Recovered	Died	Per cent. recovered	Per cent. died
1st to 3d day.....	199	163	36	81.9	18.1
4th to 7th day.....	346	252	94	72.8	27.2
Later than 7th day. .	666	423	243	63.5	36.5
Totals.....	1,211	838	373	69.2	30.8

#### MORTALITY ACCORDING TO AGE.

Age	No. of cases	Recovered	Died	Per cent. recovered	Per cent. died
Under 1 year.....	129	65	64	50.4	49.6
1 to 2 years.....	87	60	27	69.0	31.0
2 to 5 years.....	194	139	55	71.6	28.4
5 to 10 years.....	218	185	33	84.9	15.1
10 to 20 years.....	360	254	106	70.6	29.4
Over 20 years.....	288	180	108	62.5	37.5
Age not given.....	18	11	7	61.1	38.9
Totals.....	1,294	894	400	69.1	30.9



## MORTALITY ACCORDING TO AGE AND PERIOD OF INJECTION.

Age	Injected 1st to 3d day				Injected 4th to 7th day				Injected later than 7th day			
	No. of cases	Recovered	Died	Per cent. died	No. of cases	Recovered	Died	Per cent. died	No. of cases	Recovered	Died	Per cent. died
Under 2 years...	13	12	1	7.7	37	28	9	24.3	159	81	78	49.1
2 to 5 years...	30	24	6	20.0	66	49	17	25.8	93	63	30	37.3
5 to 10 years....	55	49	6	10.9	69	61	8	11.6	77	61	16	20.8
10 to 20 years....	67	58	9	13.4	106	73	33	31.1	171	115	56	32.7
Over 20 years....	34	20	14	41.2	65	39	26	40.0	164	101	63	38.4
Totals.....	199	163	36	18.1	343	250	93	27.1	664	421	243	36.6

2. *Streptococcus Immune Sera*.—The rôle of the streptococcus in some diseases, for example, scarlet fever, is imperfectly understood. Moreover it has only been indefinitely established whether there are various groups or only one kind of streptococcus; even the significance of their virulence or hemolysin formation is not clear. These difficulties account for the great number of methods advocated for the production of an immune streptococcus serum. The oldest serum is that of Marmorek. It was produced by immunization with a strain made highly virulent by passage through animals. The other sera on the market are:

a. *Serum of Aronson* (Schering).—This is a polyvalent serum produced by immunization of horses with cultures pathogenic for man; some strains having previously been passed through animals, others not. The strength of the serum is tested in mice infected with the latter strains.

b. *Serum of Meyer-Ruppel* (Höchst Farbwerke).—Horses are first immunized with a strain of streptococcus whose virulence has been raised by passage through horses and mice; each horse is then injected with a different strain of human streptococcus. When the serum of each animal is of such a strength that doses of 0.01 to 0.0005 c.cm. protect mice infected with its own particular strain, the sera of the different horses are mixed. Thus a polyvalent serum is obtained.

c. *Serum of Menzer* (Merck) is monovalent and produced by immunization with a culture which is pathogenic for man and not passed through animals.

d. *Serum of Moser* is polyvalent, produced by injections of streptococci from scarlet fever. The sera of Menzer and Moser are not tested by injections of white mice. The others are. One cannot strictly rely upon this method of serum titration for its employment in man. The virulence of streptococci against mice and human beings bears no definite relation. A serum may be perfectly efficient in mice both for prophylactic and therapeutic purposes, and be entirely inactive in man; also vice versa. The action of the serum should be in the main of bacteriotropic nature.

Antistreptococcus serum has been tried in scarlet fever, puerperal sepsis, erysipelas, and articular rheumatism.

Complement fixation experiments (Foix and Mallein, Schleissner)

have shown that the streptococci of scarlet fever can be definitely separated from the other varieties of these bacteria. This work could not be confirmed.

In this disease favorable results have been observed by the use of Moser's serum.

Escherich states that of 112 scarlet fever cases injected, those receiving the serum on the first and second days of their illness all recovered, while of those injected later on, there was a high percentage of mortality. Other authorities have seen no, or only very slight effect from the serum treatment.

Two hundred cubic centimeters of Moser's serum must be given subcutaneously.

The treatment of puerperal fever has been favorably influenced by Aronson's and Meyer-Ruppel's serum, of which 50 c.cm. are injected on several successive days.

Menzer's serum is said to serve its purpose best in acute and chronic rheumatism as well as in tuberculous mixed infections.

In erysipelas all the well known sera have been tried. On account of the very variable course of the disease it is difficult to judge the exact value of the serum. In fact, thus far one cannot with certainty depend upon any serum treatment of a streptococcus infection, but the serious nature of such infections makes every possible therapeutic measure strongly justifiable.

3. The pneumococcic sera most frequently used are those of Pane, Pneumococ- Römer and Merck. Pane immunizes donkeys with highly virulent pneumococci and uses the serum for the treatment of pneumonia. Several Italian investigators record favorable results.

Römer prepares a polyvalent serum by injecting horses with different strains of pneumococcus obtained directly from man; the strength of the serum is tested in mice. The serum is mainly employed both for the protection and cure of *ulcus cornea serpens*.

The result according to Römer depends upon the very variable virulence of the pneumococci. The severity of the infection in man is said to run parallel with the virulence in mice. Römer, therefore, ascertains in every case of *ulcus serpens* whether his serum has any protective bodies for that particular strain of pneumococcus, and tests the virulence of the same.

The serum can be injected intravenously and subcutaneously, and in pneumococcus meningitis, subdurally. It is manufactured by the Höchst Farbwerke, in vials of 10 and 20 c.cm.

A similar serum is manufactured by Merck. It is obtained from horses and standardized at the Institute for experimental therapy in Frankfort so that 0.01 c.cm. injected subcutaneously protects a mouse inoculated intraperitoneally 24 hours later with 10 to 100 times the lethal dose of a living pneumococcus culture. This is known as a normal serum and 1 c.cm. contains one immunity unit (I. E.). The serum on the market contains 20 to 40 units per c.cm.

In pneumonia 200 to 400 units are given subcutaneously and repeated in three to four days, if the fever does not subside. As a prophylactic inoculation, 200 to 400 units are given to old people where a "hypostatic" pneumonia is feared. In *ulcus serpens* of the cornea 200 to 400 units are employed and if no improvement sets in, the dose is repeated upon the third day. In addition, several drops of the serum are



instilled into the conjunctival sac every two hours. As a prophylactic dose in this disease, 100 units suffice.

Merck also prepares a vaccine of dead pneumococci in doses of 1 c.cm. which further aid in the treatment of pneumococci infections. One cubic centimeter of such dead pneumococci can be administered for the prophylaxis of *ulcus serpens*.

4. *Pest Sera*.—A large number of pest sera are in use.

a. The Paris serum (Yersin) produced at Pasteur Institute by immunization of horses with dead and later on living bacilli.

b. The Bern serum of Tavel employs the same principles.

c. *Lustig's Serum*.—For this serum, horses are immunized with the pest-nucleoproteids. Pest cultures are broken up by 1 per cent. of potassium hydroxide and from this, by the addition of acetic acid, the nucleoproteid is precipitated and then suspended in salt solution to serve as antigen.

d. Serum of Terni-Bandi is prepared by the immunization of donkeys and sheep with natural pest aggressins.

e. Serum of Markl is supposedly an antitoxic serum prepared by immunization with filtrates of old pest bouillon cultures.

All the above sera contain agglutinins, precipitins, bacteriotropins and amboceptors; the serum of Terni-Bandi contains aggressin amboceptors, that of Markl, anti-endotoxins.

The sera are tested for their anti-infectious properties in animals such as guinea-pigs, rats, mice. Markl also estimates the toxin neutralization power of his serum.

The Paris serum comes either in dry form or in bottles containing 20 c.cm. without any preservatives. Ten to 20 c.cm. should suffice as a prophylactic injection, although Martini advises 100 c.cm. at least. The period of protection is short, averaging about fourteen days.

Prophylactic injection is advisable in those instances where an immediate protection is necessary, like the inoculation of physicians and nurses attending pest patients. Under all other circumstances either active immunization or the simultaneous method of Shiga should receive the preference.

For the treatment of pest infections, Calmette and Salimbeni advise intravenous administration of 20 c.cm. and two subcutaneous injections of 40 c.cm. each—all to be given on the first day; on the second day two similar subcutaneous injections; and if the case is of a severe nature, the dose may be doubled. The results are variable.

From comparative studies, it seems that Lustig's serum is somewhat weaker than the Paris serum. The sera of Terni-Bandi and Markl have not been sufficiently employed, so that opinion is reserved.

5. *Tuberculosis Sera*.—The best known and most studied are those of Maragliano and Jarmorek.

a. Serum of Maragliano is prepared by Maragliano's institute in Genoa from horses which are immunized for about six months with the soluble substances of tubercle bacilli. The favorable action of the serum is reported on, especially by Italian authorities.

b. *Serum of Marmorek* is prepared in the laboratory of Marmorek, at Paris-Neuilly, by the immunization of horses with the so-called "primitive" tubercle bacilli, *i.e.*, young tubercle bacilli whose acid-fast character is still very slight or entirely absent. When the horses have attained a high grade of immunity, they receive injections of various strains of pure cultures of streptococci obtained from the sputum of tuberculous patients. The serum of these animals is, therefore, antituberculous and at the same time polyvalent antistreptococcic (a double serum), serving against the mixed infections.

This serum is administered daily, either subcutaneously 5 to 10 c.cm. or per rectum

20 c.cm. The latter form is more advisable for the sake of preventing anaphylaxis. Citron has found the serum entirely harmless, the bad effects described by some being probably due to the idiosyncrasy of patients against foreign sera. The most favorable results have been claimed in localized bone and joint tuberculosis and in the incipient stages of pulmonary tuberculosis. Especial consideration of the serum should be given in those patients who evince persistent temperature or the very severe but not hopeless cases, where the tuberculin therapy cannot be undertaken. In some of these instances very encouraging results have been noted.

Occasionally the author started with the serum treatment, and then combined with it the tuberculin administration and finally left the serum entirely.

6. *Anthrax Sera*.—Sclavo, Deutsh, Sobernheim and others have produced immune sera by the immunization of donkeys, sheep and horses. These have been mainly employed in veterinary practice.

In man the serum has been tried only by Sclavo. He injects 30 to 40 c.cm. subcutaneously for several successive days; in severe infections 10 c.cm. are administered intravenously. Two cases described by Bandi received 150 c.cm. intravenously.

7. *Typhoid Immune Sera*.—The ordinary bacteriolytic sera (Tavel) have not met with the desired success in the therapy of typhoid fever. Attempts have, therefore, been made to produce antiendotoxic sera. Chantemesse treats horses for several years with bouillon filtrates; Besredka injects first dead and then living typhoid bacteria from agar cultures, Mac Fadyen breaks up the bacteria at very low temperatures and thus liberates the endotoxin for purposes of immunization. Kraus and von Stenitzer use bouillon filtrates and aqueous bacterial extracts as is likewise done by Meyer-Bergell and Aronson. Garbat and Meyer employ sensitized typhoid bacilli, *i.e.*, bacteria united with their bacteriolytic amboceptors.

Chantemesse injects several drops of his serum subcutaneously. Its effect lasts ten days. Only occasionally is a second inoculation necessary; if so, it must be much smaller. His results have been good and have mainly depended upon an increase in the opsonic index.

Meyer and Bergell as well as Kraus give 20 to 50 c.cm. subcutaneously.

8. *Cholera Serum*.—Similar attempts for the production of a cholera antiendotoxic serum have been made. Kraus has succeeded in obtaining an antitoxin against some El-Tor vibrios which have all the characteristics of true cholera vibrios.

The experiments with Kraus' serum, and Kolle's serum (Bern Institute), at present being conducted in Russia, seem to be favorable.

The serum therapy of infectious diseases is still in its prime. The contradictory results of many authors are to be attributed not only to the variable efficiency of the sera, but also to the method, the time, and the dose chosen for administration.

The same serum in the hands of different physicians may yield opposite results. These subjective sources of error must be overcome or minimized by making a complete and thorough study of the effects which a certain serum may have and actually does have; here all the clinical and laboratory guides must be made use of. Employed in this manner, serum therapy will even at the present stage lead to beneficial results.

Wright's motto at the beginning of his book on vaccines "The physician of the future will be an immunizator," can justly be reversed to read, "the immunizator of the future will be the physician in the true sense."



## CHAPTER XIX.

### CHEMOTHERAPY.

DEFINITION. METHOD. ATOXYL. SALVARSAN. CHEMOTHERAPY OF MALIGNANT TUMORS. CONCLUSION.

Serum therapy proved the fundamental fact that it is possible by the injection of specific sera selectively to destroy or counteract the poisonous effects of certain micro-organisms without in any way injuring the infected host. That similar results are attainable by chemical means is demonstrated by the empirical use of quinine in malaria. For a long time this chemical specific stood in a class by itself. In recent years, the progress made in the study of infections by the protozoa, especially the trypanosomes and spirochetes, and the possibility of transmitting these diseases to the lower animals, stimulated a renewed effort in quest of chemical substances analogous to quinine. Paul Ehrlich led the way in this new direction and termed this study "Chemotherapy" in contradistinction to "Pharmacotherapy."

Only those agents can be employed chemotherapeutically in which the "organatropé" and "parasitatropé" relation is favorable, that is, primarily when the curative dose is only a very small fraction of the toxic dose (Ehrlich). In infectious diseases it is the causative parasite that is aimed at, in malignant tumors the tumor cells are the objects for destruction. In both, however, it is absolutely essential that the *normal tissues of the body remain entirely uninjured*.

Thus far the most favorable results of chemotherapy have been accomplished in trypanosome and spirochete infections.

Researches under Ehrlich's direction clearly defined that there are three different types of substances which can destroy the trypanosomes:

- (a) The group of basic silk dyes (Fuchsin).
- (b) The group of cotton dyes (Benzopurpurin series) of which trypan red and trypan blue have proved most efficient.
- (c) The group of arsenical products (atoxyl and its derivatives).

The basis for differentiating these three classes is seen from the following experiments. If a mouse suffering from trypanosomiasis receives an injection of an active fuchsin preparation, the trypanosomes disappear from the blood. They remain away permanently ("Sterilisatio magna") provided that the injected dose is large enough. The animal is thus cured by a single injection. If the dose is not sufficient, many

of the parasites are destroyed, but some remain alive. This number may be so small that for several days the blood may seem sterile, but then the remaining trypanosomes multiply and a relapse occurs. The same fuchsin preparation is again injected and now, one may attain a complete cure if the dose this time is sufficiently large; but if not a relapse sets in.

Thus the treatment is continued. If, however, the relapses are numerous and the same curative agent is repeatedly employed, a state is reached in which the fuchsin no longer has any beneficial influence whatever. The trypanosomes have become immunized or acclimated to fuchsin, in other words, have become "fuchsin-fast."

To prove that it is the trypanosomes that have acquired a new characteristic and not that the organism of the mouse had become altered in its susceptibility, one may infect for the first time another mouse with this "fuchsin-fast" strain, and here again this dye will have no effect at all. The artificially attained resistance toward fuchsin remains as a permanent characteristic of this particular strain of trypanosomes, even though transplanted from mouse to mouse for years. It disappears, however, if the parasites are allowed to increase by a sexual cycle of development which is possible for example in the rat louse *Hematopinus spinolosus*.

This fuchsin-fast character has not at the same time altered the susceptibility of the trypanosomes toward any of the trypanocidal agents of the second or third group as trypan red or atoxyl. Just as guinea-pigs immunized against cholera may succumb to infection by the typhoid bacillus, so also may trypanosomes, which have become inert to fuchsin, be attacked by trypan red or atoxyl. *Furthermore the acquired resistance of the protozoa to the action of these various chemicals is just as specific as is the immunity of an animal toward bacteria.*

Group reactions play a rôle in chemotherapy just as they do in immunity; the individual vaccinated against cow pox becomes immune toward small-pox; the strain of trypanosomes which has become resistant toward trypan red can no longer be affected by trypan blue.

Ehrlich's conception that it is necessary for substances to be taken up by the parasites before they can be acted upon destructively is concisely expressed by him: "*Corpora non agunt nisi fixata.*" In terms of the side chain theory, the protoplasm groups which have a specific affinity for the chemical radicles are known as chemoceptors. Accordingly, the trypanosomes may be said to possess three distinctly different chemoceptors, one directed against fuchsin, one against benzopurpurin and another against atoxyl.

While the early chemotherapeutic studies with the aniline dyes were mainly of theoretical interest, the arsenic derivatives soon claimed their position of practical importance.

In 1902 Laveran and Mesnil discovered that arsenous acid could



destroy the trypanosomes in the blood of infected mice; but at the same time it acted as a severe general poison. Atoxyl was then introduced by Ferd. Blumenthal and was found to be less poisonous and just as efficient. Extensive experiments were undertaken with this drug and its effects both in infected mice and human beings (sleeping sickness) carefully studied. Most of this work was carried out in the infected districts of Africa by Robert Koch and his co-workers. While the trypanocidal action of atoxyl was distinct, its effect was not permanent and relapses occurred. Uhlenhuth tried this arsenic compound in animal syphilis, basing his application upon the apparently close relationship existing between the spirochete and trypanosome as pointed out by Schaudinn. The results were very encouraging, so that for a time atoxyl was employed in the treatment of human syphilis with beneficial effect upon its clinical manifestations. Its use had to be abandoned, however, on account of its very severe associated toxic actions, of which optic atrophy was the most important.

Up to that time the chemical composition of atoxyl had been represented as meta-amino-phenylarsenic acid. Ehrlich and Bertheim found that this was incorrect and that in reality it was p-amino-phenylarsenic acid, called by Ehrlich arsanilic acid. It was thus made possible by systematic substitution to attain specifically active but less poisonous products.

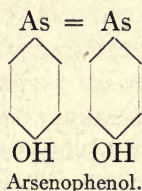
For a long time too it was unexplained why the trypanocidal action of atoxyl was so very marked in vivo and yet no effect could be noticed in vitro. Ehrlich presumed that when atoxyl enters the system it undergoes the chemical change of reduction; from a quinquivalent arsenic combination a trivalent one resulted; and it is this reduction product to which atoxyl owed its activity. He proved this hypothesis by reducing atoxyl in the test-tube; he thus obtained the phenylarsenious acid which even in the dilution of 1 to 1,000,000 was capable of destroying the trypanosomes, whereas atoxyl in the dilution of 1 to 100 was inactive. "The arsenoceptor cannot unite with quinquivalent arsenic products, but requires ones with trivalent arsenic; this lack of saturation of the arsenic molecule is essential for the chemical union with the arsenic receptor" (Ehrlich).

The numerous trivalent arsenic combinations are by no means equally potent. Many are entirely unable to destroy the protozoa, others are slightly active, while only few show a distinct specific action. Belonging to the last class is arsenophenyglycin which contains an acetic-acid radical. This compound is peculiar in that it can attack trypanosomes which have become resistant to atoxyl. If, however, these "atoxyl-fast" trypanosomes are treated for a long time with arsenophenyglycin, they became resistant also to the latter compound. According to Ehrlich's explanation for this

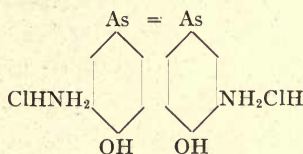
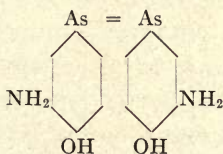
Arseno-  
phenyl-  
glycin.

phenomenon, the trypanosomes possess in addition a chemoceptor with an affinity toward acetic acid, and therefore any strain which has become resistant toward arsenophenylglycin has become not only arsenic fast but has become immune to all combinations which like arsenophenylglycin contain an acetic acid radicle.

The action of an arsenical compound depends upon its being anchored not only by means of one of its groups but by many groups, "Just as a butterfly that is to be mounted must be fixed by more than one attachment. First a needle must be passed through its body and then successive fixation through the animal's wings is necessary." In this sense Ehrlich speaks of primary and secondary haptophores. In arsenophenylglycin the primary anchoring group is the receptor of the acetic acid radical which delivers the arsenic element to the cell. Whereas for the trypanosomes it is this acid group which plays the main rôle, for the spirochete, it is the hydroxyl group, placed in the para-position of the trivalent arsenic molecule (arsenophenol).



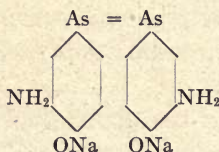
By the introduction of different elements or combinations into the above formula (as iodine or an amido group) the toxic action of the compound can be further diminished, whereas the spirillocidal property is greatly enhanced. In this manner, Ehrlich and Bertheim developed the preparation 606 (salvarsan) which is the dihydrochloride of dioxydiamidoarsenobenzol.



Dioxydiamidoarsenobenzol is a canary yellow powder undergoing oxidation very readily so that it is kept in small vacuum tubes. It is insoluble in water but soluble if sodium hydroxide is added. The hydrochloric acid salt, salvarsan, is soluble in water giving a clear yellowish solution. It is acid in reaction and absorbed by the system with difficulty (Hata). On the addition of sodium hydrate first a monochloride and then a neutral salt in the form of a flocculent precipitate results. More of the hydroxide redissolves, the precipitate giving a clear solution of the alkali salt; its reaction is strongly alkaline. It makes no difference whether the dioxydiamidoarsenobenzol is dissolved in sodium hydrate or whether the hydrochloric acid salt is made alkaline



by adding the hydroxide: in both instances the sodium salt of dioxidyamidoarsenobenzol is formed.



*The toxicity* of salvarsan is comparatively mild. The following table of Hata shows this in detail.

Animal	Mode of application	Dose tolerated
Mouse.....	Subcutaneous.....	1 : 300 pro 20 g.
Mouse.....	Intravenous.....	1 : 350 pro 20 g.
Rat.....	Subcutaneous.....	0.2 pro 1 kg.
Hen.....	Intramuscular.....	0.25 pro 1 kg.
Hen.....	Intravenous.....	0.08 pro 1 kg.
Rabbit.....	Intravenous.....	0.1 pro 1 kg.
Rabbit.....	Subcutaneous.....	0.15 pro 1 kg.

Salvarsan has no effect upon spirochetes in the test-tube. In vivo, however, Hata found a single injection of a relatively small dose sufficient to effect a complete cure in mice infected by the spirillum of relapsing fever. The results are seen from the following table.

#### CURATIVE EFFECT OF DIOXYDIAMIDOARSENOBENZOL.

Complete cure obtained after.

Dose	1 Injection	2 Injections	3 Injections
1 : 600	100%	.....	.....
1 : 700	100%	.....	.....
1 : 800	100%	.....	.....
1 : 1000	75%	100%	100%
1 : 1500	18%	75%	100%
1 : 2000	16%	66%	100%
1 : 3000	0%	0%	33%

Salvarsan is even more active in hen spirillosis; here Hata found 0.0035 gm. pro kilogram to be a curative dose.

Syphilis in rabbits requires 0.01 to 0.015 gm. pro kilogram by intravenous injection for curative action.

After these very favorable results were repeatedly obtained in the lower animals, salvarsan was used in man. A very large number of cases has now been treated and important and definite conclusions have been reached.

1. *The toxicity of salvarsan for the human being either normal or infected is very slight, provided that the solution be properly prepared and injected intravenously. The therapeutic doses are non-poisonous.*

Isolated examples of idiosyncrasy against salvarsan may be met with, but this is possible with any of the more powerful drugs. Such idiosyncrasy is always more liable after repeated administration (hypersusceptibility).

In animals it has been found that a coexisting bacterial infection may greatly raise the poisonous action of salvarsan. One should therefore guard against injecting a syphilitic who is in addition suffering from an acute infection as an influenza.

2. *Salvarsan acts as a curative agent in all varieties of spirochete invasion.* The most favorable reports have been obtained in *Febris recurrens* (Iversen, Bitter and Dreyer) and *Framboesia tropica* (Koch and Flu). These diseases may be completely cured by a single injection of a sufficiently large dose. This realizes Ehrlich's ideal: "*Therapia magna Sterilisans.*"

*Its use in syphilis* is also of undoubted value; it is one of the Syphilis. most efficient antisymphilitic agents, and as a rule, influences favorably the different manifestations of the various stages of lues. Its action is all the more marked in those individuals in whom mercurials seem to be ineffective (lues maligna) or in those having an idiosyncrasy to mercury. One should remember, however, that the radical cure of syphilis and the disappearance of all the visible clinical manifestations are two entirely different considerations. They are by no means identical, as has been definitely shown by the complement fixation test for syphilis. If a positive Wassermann reaction is to be taken as an evidence of active lues requiring treatment, then it must be granted that some stages of luetic infections cannot be entirely eradicated by salvarsan alone, even by the intravenous application. A complete cure by one or several salvarsan injections can be looked for only in the first or early secondary stage of the specific infection, especially if the Wassermann reaction has not yet become positive or is only weakly so. Even here ultimate cure is more certain if instead of salvarsan alone, mercurial treatment is added (gray oil, calomel, inunctions). In the other stages there seems to be no question any longer but that one must depend upon this combined therapy for lasting and ideal results. Especially does this pertain to those cases of lues asymptomatica where the only evidence of a still existing old infection is a positive Wassermann test. To make the reaction negative in these cases is no easy task. A single intravenous injection of salvarsan (0.3 to 0.6 gm.) never brings this about; neither is it accomplished if a few injections of sublimate or mercury salicylate are added. In the early era of the new therapy, the author obtained much better results in such conditions by oft repeated intramuscular



injections in large doses 0.6 to 1.0 gm. per injection than by the intravenous method. He had to abandon this, however, on account of the danger from the large quantities of arsenic thus administered.

In order to change such positive reactions it is sometimes necessary to give as many as four or five intravenous salvarsan injections (0.6 gm.) within a period of three to four weeks, and at the same time undertake mercurial inunctions or injections; then one or two additional salvarsan injections (0.6 gm.) and finally large doses of iodides extended over a long period of time. After a rest of two months the entire procedure may have to be repeated. Naturally this plan of treatment can be varied in many ways. It is not advisable, however, to continue with salvarsan too long. It is borne well, but its effect ceases. Whether the spirochetes acquire a resistance toward arsenic has not been definitely proven, although animal experiments of Ehrlich and Hata favor this view.

The value of salvarsan in para-syphilitic diseases has to a degree been misjudged on account of the early sensational communications of Alt who claimed to have cured general paralysis. Corroborative evidence by others has been lacking. The treatment of cerebrospinal lues on the other hand is frequently attended by remarkably good results, and when one considers how closely this affection may simulate tabes, tabo-paralysis, or paresis, it is appreciated with what care the diagnosis of true paralysis should be made. Here, while occasionally a certain symptom complex may be improved there is not sufficient evidence to claim a cure by even this latest most remarkable antisyphilitic agent.

Salvarsan has been of service also in other than spirochete infections. Most trustworthy are the results in tertian malaria (Iversen and Werner) furunculosis orientalis (Nicolle and Flu) and bilharziosis (Johannides). In anthrax and scarlet fever the general experience as yet allows of no definite conclusion. How great a help salvarsan as an arsenic compound will be in the blood diseases is still to be determined.

The effects upon certain animal diseases, African glanders (lymph-angitis epizootica), and the spirillosis of fowls, have been very encouraging.

3. Harmful results from salvarsan have been described by various investigators. As a general rule no deleterious effects are observed with the intravenous injections of properly prepared solutions. The intramuscular and subcutaneous injections were frequently attended by severe local reactions which at times led to skin and muscle necroses. Also toxic exanthemata simulating scarlet fever were met with. Blindness, such as has been ascribed to atoxyl or arsacetin, has not been reported. On the other hand, fever, vomiting and diarrhea often occur but in most instances are probably due to bacterial contamination of the injected solution. Still, the sudden breaking up of the large number of spirochetes

and the consequent liberation of their toxins can bring about similar phenomena. Usually all these symptoms pass off promptly.

Somewhat less transient are affections of different nerves, which have been observed in cases of recent syphilis a short time after injections of salvarsan. At first there was some doubt whether these injuries were due to the action of the new remedy or to the original specific infection. In most instances, they have proved to be of luetic origin and have disappeared under further specific treatment. Thus one dealt not with a "neurotropic" action of the salvarsan, but with the so-called "neuro rezidive" or neuro-recurrences of syphilis. Benario has demonstrated by the following careful statistics that such "rezidive" appear almost as frequently in the early stages of the infection when treated by mercury alone as when salvarsan is employed.

"NEURO REZIDIVE" WITH SALVARSAN. 194 CASES.

	Right	Left	Both sides	Unknown	Total	Percentage
Optic.....	18	11	27	8	64	29.1
Oculomotor.....	5	9	2	3	19	8.6
Trochlear.....	4	.....	.....	1	5	2.3
Trigeminus.....	2	3	.....	1	6	2.7
Abducens.....	6	2	2	3	13	5.9
Facial.....	16	16	4	.....	36	16.3
Acoustic.....	19	27	27	4	77	35.0
	70	68	62	20	220	

"NEURO REZIDIVE" WITH MERCURY. SAME CLASS OF CASES AND DURING SAME INTERVAL. 122 CASES.

	Right	Left	Both sides	Unknown	Total	Percentage
Optic.....	3	3	19	8	33	25.1
Oculomotor.....	6	3	0	6	15	11.5
Trochlear.....	.....	.....	.....	.....	.....	.....
Trigeminus.....	.....	.....	.....	1	1	0.7
Abducens.....	2	.....	.....	1	3	2.3
Facial.....	14	7	0	9	30	23.0
Acoustic.....	12	9	13	13	47	30.8
Olfactory.....	1	.....	.....	.....	1	0.7
Hypoglossal.....	1	.....	.....	.....	1	0.7

These paralyses are due to isolated spirochetes lying inaccessible in the nerve fibers. Thus, probably, they escape the action of the injected



antisyphilitic agents and remain unmolested only to multiply and cause a focal disease of the nerve.

A few reports of severer complications such as meningomyelitis, or death, due to salvarsan may be found in the literature. Upon careful review these instances reveal some responsible factor other than salvarsan alone. In this connection a possible independent bacterial infection existing at the time of the salvarsan injection (as a grippe or cold) is especially to be emphasized.

4. As contra-indications for the use of this medicament, Ehrlich mentions more serious disturbances of the cardio-vascular system, more advanced degeneration of the central nervous system, fetid bronchitis as well as cachexias, unless these be a direct consequence of syphilis. Each case, however, should be considered by itself and not as belonging to one of the above wide groups; thus while a mild valvular defect is a cardiac disturbance it does not per se necessarily contra-indicate treatment; only severe decompensation should be the inhibitory factor. The same may be said of aneurysms and renal disturbances. In incipient tabes, in early paralysis and epilepsy of syphilitic origin a prospect of success can be held out, provided the treatment is commenced immediately after the very earliest appearance of symptoms. The doses of salvarsan employed here should be smaller, because the susceptibility is greater. In syphilis of the liver the author has observed rather poor results. Others report the contrary. Luetic infections of the eye, even beginning optic atrophy, offer no contra-indication. In general, tuberculosis is no danger signal.

5. The mode of application of salvarsan is of paramount importance. It may be administered intravenously, intramuscularly or subcutaneously.

It can be prepared in an alkaline or acid solution, as a neutral suspension or in an oil emulsion. *At present the method of choice is the intravenous injection of an alkaline solution.*

The *advantages* of this procedure are the absence of any pain, complete resorption of the entire quantity injected, very rapid action, almost no by-effects. The only *disadvantage* is that the salvarsan is excreted very rapidly so that in about four days it has almost entirely left the system.

### The Technique of the Intravenous Salvarsan Injection.

(a) *Preparation of the Alkaline Solution.*—About 30 to 40 c.cm. of 0.85 per cent. sterile salt solution are measured in a narrow-necked, graduated glass-stoppered sterile cylinder of 300 c.cm. capacity, containing about 50 sterile glass beads. (In the official instructions accompanying the salvarsan, it is advised to use plain distilled water instead. In the author's experience it has not been found necessary to make matters somewhat complicated by using distilled water here and further on salt solution. He



uses either 0.85 per cent. salt solution or distilled water throughout the preparation with excellent results. The 0.5 per cent. saline solution as recommended by many instead of the 0.85 per cent. is also a superfluous refinement. It is however absolutely necessary that the *sodium chloride* is *chemically pure* and the *water* used for making up the saline solution be *freshly distilled*. Then too in the sterilization of the salt solution the latter should not simply be brought to the boiling point, but be kept at this, preferably in the moist heat sterilizer, for a prolonged period of time.) The salvarsan, *e.g.*, 0.6 gm. is added. Upon vigorous shaking the substance goes into solution. It is imperative that the salvarsan be completely dissolved and that no gelatinous drop-like particles remain. Fifteen per cent. caustic soda solution is now added in accordance with the following table:

0.2 salvarsan (0.456 gm.)	requires 0.38 c.cm. 15% Sod. Hydrate = 8 drops.
0.3 salvarsan (0.654 gm.)	requires 0.54 c.cm. 15% Sod. Hydrate = 12 drops.
0.4 salvarsan (0.872 gm.)	requires 0.76 c.cm. 15% Sod. Hydrate = 15-16 drops.
0.5 salvarsan (1.09 gm.)	requires 0.95 c.cm. 15% Sod. Hydrate = 19-20 drops.
0.6 salvarsan (1.307 gm.)	requires 1.14 c.cm. 15% Sod. Hydrate = 23-24 drops.

(Citron uses a little more of the alkali; for every 0.1 salvarsan, approximately 0.2 c.cm. sodium hydrate solution is taken.)

A yellow precipitate at once begins to form and gradually increases until a fine suspension ("neutral suspension") results; finally, when sufficient of the alkali has been added the precipitate redissolves giving a perfectly clear solution. Its reaction is strongly alkaline and would if injected in this concentrated quantity be sufficient to destroy the erythrocytes and injure the vessel wall. Warm salt solution is therefore added up to 250 or 300 c.cm. If less than 0.6 gm. salvarsan and 1.2 c.cm. sodium hydrate have been employed, a proportionately smaller quantity of saline is required. Should this solution not be quite clear or become slightly turbid after a few minutes, a few more drops of caustic soda solution should be added, a drop at a time and waiting two or three minutes after each drop to see if this quantity suffices to clear the solution. Thus prepared, it should be used at once and not allowed to deteriorate by standing, as the oxidation products of salvarsan are highly toxic.

(b) *The Technique of the Intravenous Infusion.*—The technique of the intravenous injection of salvarsan and the instruments employed vary with almost every physician who administers it. Each one has his "own method and instrument." This is hardly necessary. The author's technique is exceedingly simple and fully satisfactory.

The instrument consists of a long narrow round graduated funnel of about 300 c.cm. capacity; to it is attached a rubber tube interrupted near its lower end by one or two small pieces of glass tubing so as to



allow of inspection of the fluid before it enters the vein; to the rubber tubing is connected the Strauss canula or Schreiber curved needle. The whole is sterilized by boiling and then washed by sterile salt solution flowing through it.



FIG. 28.—First stage in the intravenous injection of salvarsan. Needle being inserted into distended vein and saline solution flowing into the vessel.

The steps in the actual injection are as follows:

1. The patient's forearm near the median vein is disinfected (alcohol, iodine, etc.).
2. An assistant compresses the upper part of the arm of the patient

to make the veins more prominent; this can be further increased by having the patient make a fist.

3. The glass funnel filled with about 20 to 30 c.cm. of normal saline solution is supported at about the level of the patient's head. The air is forced out of the tubing.



FIG. 29.—Second stage in the intravenous injection of salvarsan. Funnel lowered, blood flowing from vein into rubber tubing and can be seen in the glass-connection tube.

4. The operator inserts the needle, from which the warm saline is flowing, into one of the veins of the elbow (Fig. 28); care should be exercised that the needle does not penetrate the posterior wall of the vein.



5. As soon as the puncture has been made the nurse lowers the funnel (Fig. 29); if conditions are satisfactory the saline in the glass tubing near the needle will become blood tinged.



FIG. 30.—Third stage in the intravenous injection of salvarsan. Salvarsan solution being added to the saline solution in the funnel.

6. All pressure upon the upper arm is quickly released, the funnel is again raised and the salt solution flows into the vein. *This should not be painful and no infiltration should form.*

7. Only then is the salvarsan solution added to the saline (Fig. 30) and the funnel kept at the same high level. (It is best to have the

prepared salvarsan solution transferred from the original cylinder with the glass beads into another empty one, so that when the solution is now poured off into the funnel no glass beads can fall into the latter.)

8. The infusion is stopped when the funnel empties, thus not making use of the fluid that remains in the rubber tubing; if also this quantity is desired more salt solution can be added to the funnel.

9. On completion, the needle is removed by a quick motion.

The infusion should be absolutely painless. If it is not, or if an infiltration beneath the skin forms, the needle should at once be removed and the entire procedure repeated using another vein.

The dose for *adults* is 0.6 gm., for children 0.2 to 0.3 gm., and for infants 0.02 to 0.05 to 0.1 gm. Cachectic or very weak individuals should also receive smaller doses.

There are numerous modifications in the technique; one may fill the glass funnel with the prepared salvarsan solution right at the beginning but have the rubber tube clamped. The apparatus is fixed high up to a stand. The needle is detached from the rubber tube and inserted into the vein; when a distinct stream of blood escapes, showing that the needle is within the vein, the clamp is removed and the rubber tubing with its escaping salvarsan solution attached to the needle. Or, when the needle is in the vessel one may force some salt solution with a syringe through the needle into the vein and notice whether any infiltration occurs, before the salvarsan solution is allowed to flow. Or, a three directioned stop-cock may be employed with this object.

There is also apparatus with double funnels, one for salt solution the other for the salvarsan; the two are joined below by their rubber tubings into a single outlet; this allows of the infusion of saline first, to discover whether the vein has been properly punctured.

### The Intramuscular and Subcutaneous Injection.

These methods of administration have to a great degree been discontinued on account of the accompanying severe local reaction and pain. It is also uncertain how much of the salvarsan thus injected is actually absorbed. Simple aqueous (acid) solutions of salvarsan, 10 per cent., alkaline solutions with 5 c.cm. of fluid, neutral emulsions, and oily suspensions have been employed.

To prepare the neutral emulsion, the salvarsan powder, *e.g.*, 0.5 gm. is placed in a sterile porcelain dish and triturated carefully with 3 to 4 c.cm. of salt solution. The caustic-soda solution is added drop by drop (7 to 9) until on testing with litmus paper the reaction is exactly neutral. If need be a drop of dilute hydrochloric acid may be used. Salt solution or freshly distilled water is added up to 8 c.cm.



The oily suspension may be made by simply triturating the salvarsan in fatty oils 1 : 10; Ol. Amygdal. dulc, Ol. Sesami, Ol. Olivæ, Paraffin, Iodipin.

The *subcutaneous injection* is made between the shoulder-blades at the sides of the vertebral column and in a downward direction. It is best however to discard this method entirely.

The *intramuscular injection* is made in the upper outer quadrant of the gluteal region; it should be given deeply and very slowly, so as not to injure the muscles. The sciatic nerve must be carefully avoided. This method is chosen when the intravenous application is impossible (small veins, excessive fat) or when the injections cannot be repeated and a prolonged action is essential. A favorite plan is to give one to two intravenous injections followed by one to two intramuscular ones.

Owing to the difficulty with which the salvarsan solution is prepared, and its marked irritating local effect, Ehrlich produced a modification of this substance eliminating these two characteristics. This new chemical agent, *neosalvarsan*, is a

condensation product of formaldehyde sulphonylate of sodium ( $\text{CH}_2\text{-(OH)O.SO.Na}$ ) and salvarsan. Its composition is dioxydiamidoarsenobenzol-monomethane sulphinate of sodium. It consists of a yellowish powder of peculiar odor and dissolves very easily in water, with a completely neutral reaction resulting.

The average single dose of neosalvarsan is half again that of salvarsan as it contains only 66 per cent. salvarsan. On account of its non-irritating properties, it is very readily administered by intramuscular injection. Subcutaneous injection must be avoided, owing to the danger of infiltration.

The preparation of the new solution is exceedingly simple. The powder dissolves in freshly distilled water with hardly any shaking. A 0.4 per cent. saline solution may be used instead of the plain water, provided it is made from chemically pure sodium chloride and freshly distilled water. As with salvarsan, solutions of neosalvarsan must be injected immediately after their preparation. The temperature of the injected fluid should not rise above  $20\text{--}22^\circ\text{C}$ . ( $68\text{ to }71.6^\circ\text{F}$ ). Warming the liquid must be avoided.

For intravenous injection 25 c.cm. of distilled water or saline are required for each 0.15 gram neosalvarsan; but if desired it may be administered in much more concentrated solution as it is by the intramuscular method; for example, 0.6–1.5 gm. in 10–15 c.cm. of fluid. This quantity can be injected by means of a 15 c.cm. glass syringe, thus eliminating the use of large or complicated apparatus. Especially in children is this of advantage.

The injections of neosalvarsan are without a doubt better borne than a corresponding dose of salvarsan. On this account the new remedy

may be given four or five times at one or two day intervals. Naturally such treatment will depend entirely upon the stage of the disease, its clinical manifestations and the physical condition of the patient.

As for the comparative value of salvarsan and neosalvarsan, statistics are still too few. Thus far it seems that while the immediate effect of neosalvarsan upon active luetic lesions is similar to that of salvarsan, its ultimate effect upon the Wassermann reaction and the final eradication of the syphilitic infection is not as striking.

While chemotherapy has already made great progress in protozoon infections, its importance in bacterial diseases is still very slight. The drawback lies in the injury to the normal tissues which occurs simultaneously with the attack upon the pathogenic bacteria by the chemical agents. Most favorable results in this field have been obtained in typhoid fever with Xylene, and in pneumococcus infections with quinine derivatives (Morgenroth). The latter experiments especially offer a promising prospect for successful therapy in the human being.

Far more interesting and remarkable are the experiences of A. von Wassermann and his co-workers M. Wassermann and Keysser with the chemotherapy in malignant tumors. In a mixture of tumor cells with sodium tellurate and sodium selenate, it was noticed that a certain affinity existed between the cells and this chemical, in that the metal was found precipitated within the cell around the nucleus. This was confirmed in living mice by injecting solutions of these salts directly into the tumors. Thereupon it was shown that softening and liquefaction of the tumors occurred. Instead of the hard masses, a cyst-like structure resulted which usually ruptured and evacuated a brownish semifluid material. This was sterile and had a strong odor of selenium and tellurium. The cyst finally healed.

The next problem consisted in obtaining selenium and tellurium compounds which could be injected into the circulation and thence reach selectively all the carcinoma cells. v. Wassermann placed great stress upon the "building of rails which would reach the tumor and by which the selenium could travel." After painstaking experiments involving hundreds of preparations he finally utilized eosin with its exceptional diffusing power as the rails by which to run the selenium into the system and send it straight to the cancer cells. Such a chemical agent employed to transport another substance to particular cells or organs Wassermann named "Cytotrochin."

"If three consecutive daily intravenous injections of the eosin selenium compound are given in 2.5 gm. doses for 15 gms. mice, a distinct softening and elasticity of the tumor are noticed on the fourth day; on the fifth day a fourth injection of the same dose is given, after which there is no longer



the feeling of a solid tumor but rather that of a fluctuating cyst in which small movable tumor particles can be discovered. After the fifth injection on the seventh day, this soft mass becomes smaller, the capsule becomes lax, and the configuration of a circumscribed tumor can no longer be distinguished, but only a long edematous cord can be felt. Usually as a result of the sixth injection, in favorable cases, the absorption and diminution proceed so that one gets the feeling of an empty sac. In case no intercurrent disease occurs, the animal is cured in about 10 days, with a disappearance of all remnants of the tumor."

The intravenous injection of mice requires experience. The substances are injected into the caudal vein. The mouse is put into a special retainer and the cover of the trap is closed, leaving the tail alone outside. In Mice. The mouse is fixed firmly by grasping the tip of the tail between the left thumb and index finger and holding the tail fully extended. The caudal vein, which is already somewhat congested by the pressure of the cover of the retainer upon the root of the tail, is made more prominent by gentle exposure to heat in the form of a small electric bulb repeatedly passed over and close to the skin. After a little while the superficial epithelial cells become injured by the heat, so that they can be removed by gentle scraping with a sharp scalpel. This exposes the vein. The successive inoculations should begin at the tip of the tail and gradually approach the root. A very fine needle is essential. If the tip of the tail becomes necrotic, it should be simply cut off.

The exceedingly instructive observations of v. Wassermann Conclusion. are at present mainly of scientific character. It would be erroneous at once to extend such application to human therapy. Still, one cannot fail to see in this work the promise of greater things in the future. The important principle that it is possible to have chemical substances pass from the blood vessels and specifically attack tumor cells has been definitely established; and thus it seems merely a question of time before the right step is taken toward the solution of one of the greatest of human problems.

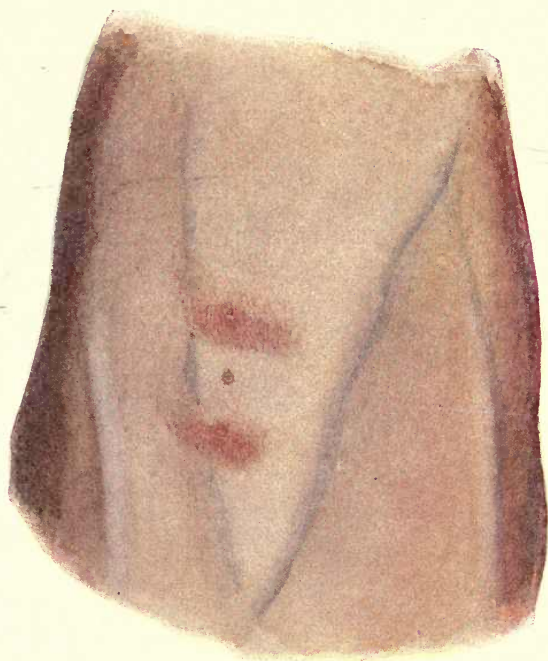


Fig. 1. Positive v. Pirquet Reaction  
(Original drawing)

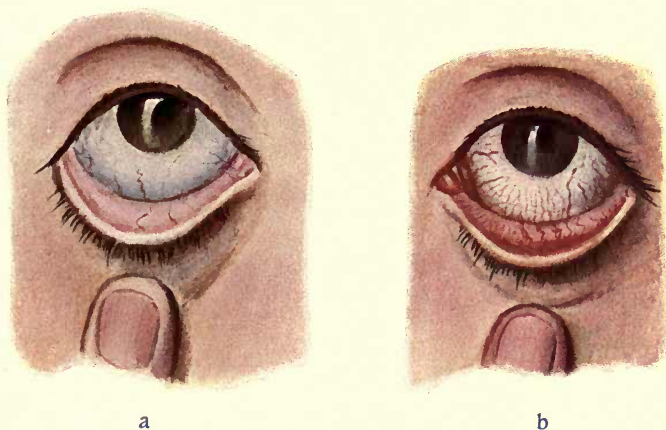


Fig. 2. Ophthalmo-reaction  
(Original drawing)

a) Control eye

b) Reaction of 1<sup>0</sup>–2<sup>0</sup> grade





## INDEX OF SUBJECTS

### A

- Abrin, 95
- Acne Vaccine, 211
- Actinocongestin, 222
- Actinomyces, 196
- African glands, 246
- Agglutination reaction, differentiation of bacterial species by, 106
  - group agglutination, 110
  - partial agglutination, 110
  - in cholera infections, 113
  - in dysentery, 114
  - in epidemic cerebrospinal meningitis, 114
  - in glands, 115
  - in Malta fever, 114
  - in paratyphoid fever, 113
  - in pest, 114
  - in tuberculosis, 114, 115
  - in typhoid fever, 106, 113
- Agglutinating sera, production of, 109
  - preservation of, 109
- Agglutination tests, technique of, 106-109
- Agglutinins, against red blood cells, 115; see "hemagglutinins."
  - bacterial, 105-115
    - action of, 105
    - biological structure of, 106
    - definition of, 105
    - diagnostic value of, 106
    - group agglutinins, 110
    - partial agglutinins, 111
    - preservation of, 109
    - production of in sera of animals, 109
    - specificity of, 105-106
- Agglutininogen, 110, 112
- Agglutinoids, 112
- Agglutinophore group, 112, 160
- Aggressins, 36-44, 61, 232
  - aqueous, 39
  - artificial, 38, 232
  - natural, 35-38
  - serous, 39
- Albumin differentiation, 124-130, 155, 193-195; See under "precipitin," "proteid"
- Aleuronat solution for injection to produce peritoneal exudates, 138, 197
- Alexin, 133, 152
- Allergie, 224; see "Anaphylaxis"
- Amboceptors, 133, 152, 153, 232, 233
- Ananaphylaxis, 224
- Anaphylaxis, 201-230
  - Arthus phenomenon in, 222
  - classical picture in guinea pig, 223
  - due to proteids, 222
    - to actino congestion, 222
  - factors governing occurrence, 223
  - forms of, 221
- Anaphylaxis, in dog, 227
  - in guinea pig, 227-228
  - in hay fever, 229
  - in man, 223, 226-227
  - in rabbit, 227-228
  - in tuberculosis, 161, 162, 221
  - passive, 224
  - prevention of, methods for, 224
  - relation to peptone poisoning, 228
  - serum sickness, 223, 226, 227
  - Theobald Smith phenomenon, 222-223
  - theories of, 224-225, 228
- Anaphylatoxin, 225-226
  - in tuberculosis, 161, 221
- Anemia, pernicious, 100
- Animal sepsis, varieties, 22
- Ankylostoma, 170
- Anthrax, 26, 153, 172, 246
- Antiaggressin, 43, 232
- Antiamboceptors, 143, 233
- Antianaphylaxis, 224
- Antibodies, 3, 4, 5, 31, 167, 171, 221
- Antibody production, local, 64, 95
- Anticomplements, 155
- Antiendotoxin, 224, 225, 231
- Antiferments, 101-104
- Antigen, 21, 110, 156, 157, 158, 171, 173, 174.
  - See under "complement fixation in syphilis."
- Antigenophile group, 154
- Antihemotoxin, 92-94
- Antikutine, 163
- Antileucocyte ferment, 102
- Antilysin, 92-94
- Antiserum, 173; see under "serum therapy in—"
- Antistaphylolysin, 92-94
- Antitoxin unit, 80
- Antitoxins, 4, 77-101, 160. see under "diphtheria"
- Antitrypsin, 102-104
- Antituberculin, 46, 155, 156, 157, 158, 160, 162
  - as guide to prognosis, 163
- Aortitis, 167
- Arachnolysin, 96
- Arsenous acid, 241
- Arsanilic acid, 242
- Arsenophenylglycin, 242, 243
- Arthus phenomenon, 222
- Ascarides, 170
- Atoxyl, 240-242
- "Atoxyl fast," 242
- Autoinoculation, 201
  - in tuberculosis, 201-203
- Autumn catarrh, 229-230

### B

- Bacilli emulsion (Koch), 63
- Bacillus neoformans, Doyen, 212



- Bacterial destruction by heat, 32  
 Bacterial emulsion in opsonic determination, 205, 206  
 Bacterial extract, 36-44, 173, 234  
 Bacterial filter, 16, 18  
 Bacterial immunization, 31  
 Bacterial precipitin, 121-124; see under "precipitin"  
 Bactericidal plate method, 141-143  
 Bacteriolysin, 34, 231  
 Bacteriolysis, 131-143, 232  
   demonstration by Pfeiffer's method, 131, 134-140  
   by bactericidal plate method, 141-143  
   in vitro, 132  
   explanation of, 133  
   necessity of complement for, 132, 133  
   phenomenon of, 131, 133  
   relation to complement fixation, 153  
   specificity of, 131, 133, 134  
 Bacteriotropins, 34, 197, 231, 233  
   differentiation from opsonins, 199  
   Neufeld's method of estimation, 213-215  
 Bail's classification of bacteria, 33  
 Basedow's disease, 102  
 Basic silk dyes, 240  
 Bee posion, 96, 100  
 Béranek's tuberculin, 53  
 Bilharziosis, 246  
 Biological mercurial therapy, 168-169  
 Blood cells, washing of, 98, 145, 147  
 Blood pressure, fall in diphtheria, 84  
 Blood relationship, 125-126  
 Blood removal, 13  
   by wet cups, 13  
   from vein, 13  
 Blood transfusions, 116  
   tests in, 117, 118  
 Bothriocephalus latus, 100  
 Botulism toxin, 88, 89  
 Bovine tuberculin, 71  
 Bovovaccine, 30, 71  
 Brieger's cachexia (carcinoma) reaction, 102
- C
- Cachexia reaction, 102  
 Canula for obtaining blood, 13  
 Capillary pipettes, 136, 203, 204  
 Carcinoma, 206-220  
   chemotherapy of, 255-256  
   immunity toward, 216  
   serum diagnosis of, 102, 206-220  
   transplantation of, 216  
 Casein, 103  
 Castellani's test, 111, 112  
 Centrifuge rules, 14  
 Cerebro-spinal lues, salvarsan therapy, 246  
 Chamberland filter, 17  
 Chemotherapy, 240-256  
   group reaction, 241  
   in bilharziosis, 246  
   in furunculosis orientalis, 246  
   in malaria, 246  
   in malignant tumors, 255, 256  
   in pneumonia, 255  
   in syphilis, 245-255  
   in trypanosomiasis, 240, 241  
   in typhoid fever, 255  
   principles of, 240, 241  
 Chicken cholera, 26, 36, 41  
 Cholera extract, 44  
   serum, 105, 106-108, 131, 132, 143  
   vibrios, 32, 44, 85, 105-115, 134, 135, 138, 140, 143  
 Cholesterin, 89, 99  
 Classification of bacteria, 23  
 Cobra hemolysis, 97-100  
   immunity, 99, 100  
   toxin, 96, 97-100  
 Coccidiosis, 22  
 Coli bacilli, 2, 44, 107, 172, 205, 207, 211  
 Colubrids poison, 96  
 Complement, 171, 174, 200, 231-232  
   definition, 132  
   importance of, in bacteriolysis, 132-133  
   multiplicity of, 152  
   preparation of, 148, 174  
   structure of, 134  
   titration of, 150, 151  
 Complement deviation (Neisser-Wechsberg), 143, 232  
 Complement fixation or binding, 152-170, 171-190  
   antigen in, 171, 173  
   antisera in, 171, 173  
   controls, 174, 175  
   definition, 152  
   demonstration of respective antigens by, 155, 158, 177  
   antibodies by, 156, 174-176  
   of antituberculin, 156, 157, 158, 160  
   method for serum diagnosis, 153  
   principles of, 152, 153, 154, 158  
   reaction with exudates and transudates, 173  
   relation of, to bacteriolysis, 153, 155  
   to precipitation, 155  
   specificity of, 153, 154  
   technique of, method of Bordet Gengou, 171-190  
   theories for phenomenon, 156  
 Complement fixation in diseases caused by  
   animal parasites, 170, 190  
   in echinococcus disease, 170, 190  
   in epidemic meningitis, 191  
   in gonococcus infections, 191-192  
   in proteid differentiation, 193-195  
   in scarlet fever, 236-237  
 Complement Fixation in syphilis, 163-170, 178-190  
   antigens in, 165  
   nature of, 166  
   lipoid extracts, 186, 187, 188  
   guinea pig's heart, 187, 188, 189  
   extract of spirochetes, 166  
   syphilitic liver, 166, 182, 185  
   preparation of, 178, 179  
   titration of, 178, 179, 185  
   development of reaction, 163  
   diseases of nervous system, 164  
   effect of mercurial treatment, 164, 167, 168  
   of salvarsan therapy, 164  
   general principles of, 152, 153, 154, 158  
   guide to prognosis, 169  
   in lues asymptomatica, 165, 167  
   in lues maligna, 169  
   in mothers of syphilitic children, 159  
   in wet nurses, 169

- Complement Fixation, other diseases giving  
 the reaction, 166, 185  
 modifications in technique, 182-190  
   Bauer's method, 189  
   Citron's method, 179, 184, 185, 186  
   Noguchi's method, 187  
   Meier's method, 182-184  
 positive reaction means "active" lues,  
   167, 185, 245  
   results in the various stages, 165, 167  
   rules for its occurrence, 164  
   technique, 179-182  
   with cerebro-spinal fluid, 164  
 Complement fixation in tuberculosis, 155-  
   163, 191  
   antigen in, 171, 173  
   demonstration of antituberculin, 155,  
   156, 157  
   theory, 156  
   with urine, 163, 174  
 Complement fixation in typhoid fever, 172,  
   192-193  
   principle of, 152, 153, 154  
   preparation of antigen, 171, 173, 192  
 Complementoids, 147  
 Complementophile group, 133, 160  
 Conjunctiva reaction, 52, 54, 57, 58  
   in hay fever, 229  
 Control tests, value, 5, 6, 174  
 Cow-pox, 25  
 Croton, 96  
 Cutaneous reaction in tuberculosis, 49, 50, 55,  
   58  
 Cytase, 152, 198  
 Cytolysin, 151  
 Cytophile group, 133, 154  
 Cytotoxin, 151  
 Cytotrochin, 225

## D

- Death of bacteria, 32  
 Desiccator for drying serum, 15  
 Deviation of complement, 143  
 Dilutions, 18  
   preparation of, 18, 19, 20  
 Diphtheria, 74-84  
 Diphtheria serum or antitoxin, 77-84  
   preparation of, 77, 78  
   prophylactic application, 84  
   standardization, 79-82  
   therapeutic application, 82-84  
 Diphtheria toxin, action in animals, 75-76  
   estimating strength of, 76  
   obtaining of, 74-75  
 Donath Landsteiner's test, 100-101  
 Dysentery antitoxin, 90-92  
   bacilli, 32, 44, 90-92, 114, 143  
   serum, 90-92, 114, 143

## E

- Echinococcus, 170, 130  
 Ehrlich's experiment, 100  
   side chain theory, 112, 158-160  
 Endocarditis maligna, 113  
 Endotoxin, 85, 138, 224, 225  
 Eosin selenium, 255, 256  
 Ergophore group, 112, 147  
 Erysipelas, 236

- Erythrocytes; see "blood cells"  
 Exudate, 36, 37, 136, 173  
 Extracts of bacteria, 36-44  
   production of, 138  
   removal of, 12, 136  
   staining of, 136

## F

- Fatty acids, 100  
 Febris recurrens, 166  
 Ferments, 101-104  
   biological structure of, 160  
 Fever in tuberculin reaction, 161  
 Ficker's diagnosticum, 107  
 Filters, bacterial, 16  
 Filtration, 16  
 Focal reaction, 46, 64  
 Food-stuff substitution, 125, 127, 128, 129  
 Forensic serum differentiation, 125, 129  
 Fornet's ring test, 123  
 Fowl plague, 36  
 Frambesia, 166  
 Friedberger's position, 11, 136  
 "Frigo" for preserving serum, 16, 148  
 Fuchsin, 240  
 "Fuchsin fast," 241  
 Functionating radicle of cell (biological), 158,  
   159  
 Furunculosis, 200  
   orientalis, 246

## G

- Glanders, 115  
   diagnosis with mallein, 58  
 Glycogen, 155  
 Gonococcus vaccine, 211  
 Group agglutination, 110-112  
 Group reactions, 122  
 Guinea pig sepsis, 21

## H

- Hamburger's local reaction, 47  
 Haptine, 160  
 Haptophore group, 112, 147, 160  
 Hay fever, 229-230  
   diagnosis of, 229  
   serum therapy, 229-230  
 Helminthiasis, 170  
 Hemagglutinins, 115-119  
   in transfusions, 116  
 Hemoglobinuria, 100-101  
 Hemolysis due to cobra toxin, 97-100  
 Hemolysins, 140-151, 171  
   characteristics of hemolysins, 144  
   estimation of strength, 146-149  
   immune hemolysins, 144-151  
   normal hemolysins, 144  
   phenomena of hemolysis, 144  
   preservation of, 146  
   production of by immunization, 144-145  
 Hemolytic system, 147-149  
 Hemorrhagin, 97  
 Hemotoxin, 86, 87, 96-100  
 Hen spirillosis, 244  
 Hog cholera, 4, 31, 113, 135  
 Hydrophobia, 26-30  
 Hypersusceptibility; see "anaphylaxis"



## I

Immune bodies, 133  
after typhoid prophylactic injections,

34

Immune hemolysin, 144-151

Immunity, 3

definition, 221

absolute and relative, 4

active, 3

acquired, 216

antiaggressin, 40

antitoxic, 100

attained, 4

chicken cholera, 41

conception of, 3

diphtheria, 77-84

against hog-cholera, 4, 113

against pure parasites, 41

in lues, 167, 168

against snake poison, 100

swine pest, 37, 40, 41, 42

bactericidal, 3, 40

cellular, 3

continued, 4

general, 4

"histogene," 3

local, 4, 63, 95

natural, 4

"pan immunity," 216

to tumors, 216

partial, 63, 110

passive, 4, 231, 239

tissue, 3

transitory, 4

tuberculin, 162

Immunization, active principle of, 21, 77

technique, 22, 77

with aggressin, 40-45

with dead virus, 31-34

with erythrocytes, 145

with living virus, 24-31

with toxins, 77-83

Inactivation, 132, 174

Incubation period, 27, 75, 78, 86, 223

in anaphylaxis, 223

Injection, technique, 9-12, 37

intracardial, 10, 83

intracerebral, 86

intramuscular, 83

intraneural, 88

intraperitoneal, 83

intraspinal, 234-236

intravenous, 9, 10

subcutaneous, 12, 83

subdural, 88

Intracutaneous tuberculin reaction, 51

Isoagglutinins or Isohemagglutinins, 116-118

Isoprecipitins, 126

Isohemolysins, 116-118

## J

Jennerian immunization, 26

Jequirity seed, 95

## K

Kaolin, 228

Kidney tuberculosis, 67

Killing of bacteria, 31, 32

Klausner's reaction, 124

Kolles' flasks, 38

## L

Laboratory equipment, 7-9

Law of multiple proportions, 85

Lecithin, 89, 97, 98, 99

tuberculin, 73

hemotoxins, 97-100

Leprosy, 166

treatment with nastin, 72

Leucoantifermantin, 102

Leucocidin, 151

Leucocytes, 4, 36

obtaining of, 182, 213, 214; see also under

"opsonic index," "bacteriotropin"

Lilliputian filter, 17

Limes death, 80, 81

zero, 80, 81

Lipoids; see "lecithin," 90, and "cholesterin"

Loeffler's serum plates, 102

Loop standard, 8, 19

Lues; see "complement fixation in syphilis"

asymptomatica, 165, 245

Lupus, 67

Lyssa, 26-30, 60

## M

Macrocytase, 152, 198

Macrophage, 196, 197

Malaria, 113, 166

salvarsan in, 246

Mallein, 57, 58

Malta fever, 114, 212

Measles, 41, 124, 185

Meiostagmine reaction, 218-220

Mendelian law for agglutinins, 116

Meningitis, epidemic, 114

serum diagnosis, 175-177

serum therapy, 232-236

Meningococci, 32, 172

Meningococcic serum, 113, 234, 236

titration of, 175-177

Mercurial therapy (biological) 168, 169

combined with salvarsan, 245, 246

Metchnikoff's resistance test, 138, 196-197

Microcytase, 152, 198

Mouse typhoid bacilli, 31, 70, 113

Much and Holzmann reaction, 99

Multipartial sera, 111, 112

## N

Nastin, 72

Negative phase, 78, 200

Neosalvarsan, 254-255

Nephrotoxin, 151

Neurorecurrences, 247-248 ("neurorezidive")

Neurotoxin, 86, 87, 97, 99, 151

Neutral red for vital stain, 198

New tuberculin, 62-63, 69-71

Nonagglutinable strains of bacteria, 112, 113

Nonbinding doses, 156-158, 174

Normal bacteriolysins, 138

Normal curative serum, 79

Normal hemolysin, 144

Normal loop, 10, 19, 20  
Normal toxins, 79

O

Obtaining blood, 12-14  
Ointment reaction, Moro, 50  
Oleic acid, 98-99  
Ophthalmic reaction, 52-54, 160  
Opsonic index, 199-209, 212, 213  
    definition, 199  
    diagnostic importance of, 201  
    increase by immunization, 200-201  
    relation between index and clinical condition of patient, 201, 212, 213  
    technique for determination of, 203-209  
    variations by auto-inoculation, 201, 202  
Opsonins, 198-213  
Opsonizer, 205  
Organotrope, 240  
Original tuberculin, old, 61  
Ozena bacilli, 123

P

Paradoxical reaction, 240  
Paralysis, progressive, 164, 165, 167  
Parasites, 23, 41, 232  
    half, 23, 41, 232  
    total, 23, 41  
Parasitotrope, 240  
Parasyphilitic infections, 246  
Paratyphoid bacilli, 30, 110-113, 140, 143  
Paroxysmal hemoglobinuria, 100, 101  
Partial agglutinins, 110  
Partial aggressins, 62  
Partial immunization, 63, 110  
Pathogenicity, 23  
Peptone poisoning in anaphylaxis, 228  
Peritoneal exudate, 136  
    method for removal, 12, 136  
Pernicious anemia, 100  
Pest, 114, 153, 172, 238  
    sera, 114, 131, 238  
Pfeiffer's phenomenon, 131  
    technique and practical application, 134-140  
    in cholera, 139  
Phagocytosis, 139, 196-215  
    cells engaged in phagocytosis, 196-197  
    definition, 196  
    demonstration of, 197, 198  
    object of phagocytosis, 196-197  
    during artificial immunity, 196-213  
    during natural immunity, 4  
Phagolysis, 197  
Phrynolysin, 96  
Phytotoxin, 95-96  
Pipettes for removal of peritoneal exudates, 136  
v. Pirquet's reaction, 48-50, 55  
Pneumococci, 32, 199, 237-238  
Pneumococcic sera, 119, 237-238  
Pneumonia, 102  
    chemotherapy of, 255  
    bacilli, 123  
Pollantin, 230  
Pollen poison, 229-230  
Polyvalent sera, 111, 232  
Porges reaction, 123

Positive phase, 78, 200  
Preservation of serum, 15  
    in "Frigo," 16  
Precipitation, 120-130  
Precipitate in precipitin reactions, 130  
Precipitins, 120-130  
    biological structure of, 120, 160  
    phenomenon of precipitation, 120  
    bacterial, 121-124  
    clinical value of in typhoid, 122, 123  
    in syphilis, 123, 124  
    Fornet's ring test, 123  
    group reactions of, 122  
    Klausner's reaction, 124  
    Porges' reaction, 123  
    precipitating antisera for, production of, 121  
    precipitinogens, 121  
proteid, 124-130  
    action of, 124, 125  
    determining nature of meat, by, 125, 127, 128, 129  
    differentiation of proteids by Uhlenhuth's method, 127-128  
    distinguishing blood of animal species, 125, 126  
    production of precipitating antisera, 126, 127  
Prognostic employment of autoinoculation, 201  
    of the lues reaction, 167, 169, 185  
    of the tuberculin reaction, 58-59  
Prophylactic inoculations in cholera, 35  
    hog cholera, 31  
    hydrophobia, 28  
    small-pox, 27  
    typhoid, 33-35, 44  
Prophylaxis in diphtheria, 84  
    in dysentery, 90-92  
    against hay fever, 230  
    in pest, 238  
    in tetanus, 88  
    in ulcer corneæ, 237  
Proteid differentiation by complement fixation, 193-195  
    by precipitins, 124-130  
Proteid anaphylaxis, 221-230; see "anaphylaxis"  
Pseudo-agglutination, 108  
Psycho-reaction, 99  
Puerperal sepsis, 237  
Pukal filter, 17

R

Rabbit sepsis, 21  
Rabies, 26-30  
    simultaneous treatment of, 30  
    vaccination against, 28-30  
Rat trypanosomiasis, 22, 240, 241  
"Reagine," 166, etc.  
Receptors, 133, 147, 151, 158-160, 231, 232-233  
Reichel filter, 17  
Relapsing fever in mice, salvarsan therapy, 244  
Resistance test of Metchnikoff, 138  
Rheumatic fever, 236  
Rhinoscleroma bacilli, 123  
Ricin, 95  
Ring test, 123



## S

- Saprophytes, 23
- Salvarsan, 243-255  
     chemical formula and properties, 243-244  
     therapy in hen spirillosis, 244  
     therapy in relapsing fever, 244  
     therapy in syphilis of man, 245-255  
     therapy in other diseases, 245-246
- Salvarsan therapy in human syphilis, 245-255  
     action, 245, 246  
     combined with mercurial therapy, 245, 246  
     contraindications, 248  
     harmful effects, 246, 248  
     mode of application, 248-255  
         intramuscular, 253-254  
         intravenous, 248, 249-253  
         subcutaneous, 253-254  
     neuro-recurrences in, 247  
     value in parasymphilitic diseases, 246
- Scarlet fever, 124, 166, 173, 185, 246
- Scorpion poison, 96, 100
- Scrofulous reaction to tuberculin, 50
- Seiden peptone, 155, 228
- Selenium, 255-256
- Sensitized bacteria, 70, 143  
     advantages of, 70
- Sensitized tuberculin, 70-71
- Sepsis, 113, 221, 237  
     of guinea pigs, 22  
     of rabbits, 21, 23
- Serum, color, 15  
     bacteriolytic, 231  
     obtaining of, 12, 13, 79  
     preservation of, 15, 16, 79  
     value in therapy, 231-232
- Serum diagnosis; see under individual infectious diseases and under "Complement fixation"
- of malignant tumors, 216  
     antitrypsin test, 122-124, 217  
     Freund-Kaminer reaction, 217-218  
     Meiostagmine reaction, 218-220  
     of meningitis, 175-177  
     of syphilis, 123, 124. see under "complement fixation"
- "Serum fast," 233
- Serum sickness, 83, 223, 226-227
- Serum therapy, 231-239  
     in anthrax, 239  
     in cholera, 239  
     in diphtheria, 83-84  
     in erysipelas, 236, 237  
     in hay fever, 229-230  
     in meningitis, 234-236  
     in pest, 238  
     in pneumonia, 237  
     in puerperal fever, 237  
     in rheumatism, 236  
     in scarlet fever, 236, 237  
     in sepsis, 237  
     in streptococcal infections, 236-237  
     in tetanus, 88  
     in tuberculosis, 162, 233, 238  
     in typhoid fever, 239  
     in ulcus serpens, 237
- Sessile receptors, 160, 161
- Side chain theory, 158-160

- "Simultaneous method," 30, 31, 77, 238  
     definition, 31
- Small-pox, 25
- Smith's phenomenon, 222-223
- Snake poison, 96-100  
     serum, 100
- Specificity, 1, 5, 54, 105, 110, 125, 134  
     of Koch subcutaneous reaction, 56  
     of ophthalmic reaction, 57  
     of v. Pirquet reaction, 55  
     of tuberculin reaction, 54
- Spermatoxin, 151
- Spider poison, 96
- Spirillosis of fowls, 246
- Spirochete infections, 240
- Standard loop, 8
- Staphylococci, 31, 44, 92, 199, 200, 205  
     vaccine, 209-210
- Staphylohemotoxin, 92-94
- Staphylolysin, 92-94
- Sterilisatio Magna, 240, 245  
     mode of action, 240-241
- Strauss canula, 13
- Street virus, 27
- Streptococci, 32, 173, 206  
     sera, 236-237  
         vaccine, 211-238
- Subcutaneous tuberculin injections, 45-49, 56
- Substance sensibilisatrice, 133; see "amboceptor," "bacteriolysin"
- Summation of antigen, 156, 157, 174
- Swine erysipelas, 30  
     pest, 36, 40, 41, 42, 43
- Syphilis, 123, 124; see "atoxyl," "salvarsan," "complement fixation."
- System of hemolysis tests, 147

## T

- Tabes dorsalis, 164, 165
- Tauruman, 30, 71
- Tebesapin, 72
- Tetanolysin, 86, 87
- Tetanospasmin, 86, 87
- Tetanus, 86, 87  
     antitoxin, 88  
     cerebral, 86  
     sine tetano, 86
- Thermolabile substances in serum, 15, 16, 133
- Thermoresistant substances in serum, 15, 109, 133
- Thyroid, 102
- Titration of antigen, 178, 179, 185  
     of bacteriolytic serum, 137  
     of complement, 150  
     of hemolysin, 146-149  
     of luetic sera, 179-182  
     of virulence of cultures, 34
- Toad poison, 96
- Toxins, 74-104, 147  
     action of, 74, 75  
     definition, 85  
     dilution of, 18  
     obtaining of, 74-75  
     titration of, 76  
     of higher plants and animals, 95-101
- Toxoids, 81, 147
- Toxolipoids, 100
- Toxon, 80
- Toxophore group, 147, 159

Toxozeptids in relation to anaphylaxis, 228  
to tuberculosis, 161  
Transfusion of blood, 116-118  
Transplantation of tumors, 216  
Transudate, 173  
Traumatic tuberculin reaction, 50  
Trichophytin, 58  
Trypan blue, 240  
red, 240  
Trypanosomiasis, 22, 166, 240-241  
trypanocidal agents, 240  
Trypsin, 102-104  
Tubercle bacilli, 60, 61, 62, 172, 196  
emulsion in opsonic determinations, 206  
Tuberculin, 45-73, 155-163, 200, 201-203  
action, 63-64  
as antigen, 162  
Beranek's tuberculin, 63  
bovine tuberculin, 71-72  
"depot" reaction, 47  
diagnosis, 45-49  
cutaneous, v. Pirquet, 49-50  
explanation of, 156, 157, 160, 161  
Hamburger's method, 47  
intracutaneous, 51  
ointment reaction, 50-51  
ophthalmo reaction, 52-54, 57, 58  
subcutaneous reaction, 45-49  
preparations, 62-63, 70  
new tuberculin, 63, 69-71, 211  
obtaining of, 45-46  
old tuberculin, 45-59, 62, 65  
original old tuberculin, 62  
vacuum, 62  
watery, 62  
reaction, 46-54  
theories, of Citron, 160-161  
of Wassermann, 156, 157  
therapy, 60-71, 155-162  
antituberculin production during therapy, 155-162  
contraindications, 68-69  
general principles, 64-65  
with bovine tuberculin, 71-72  
with new tuberculin, 69-71  
with old tuberculin, 65-69  
with sensitized tuberculin, 70-71  
Tuberculosis, 99, 114-115, 233 see under  
"complement fixation"  
prognosis in, from local reaction, 58-59  
sera for, 99, 114, 115, 238  
treatment of, by Friedmann vaccine, 30  
in successive steps, 66  
with nastin, 72  
with tebesapin, 72  
with tuberculin; see "tuberculin  
therapy"  
vaccination in, 30, 212-213

Tuberculosis of kidney, 67  
Tumors, malignant; see under "carcinoma"  
Typhoid bacteria, 32, 106-113, 123, 134, 135,  
171-173, 192-193  
immunization of rabbit with, 32  
titration of virulence of culture, 24  
extract, 44  
Typhoid fever, 141, 185, 239, 255  
protective inoculations, 33-35  
serum, 106, 113, 140, 141, 239  
vaccine, 33-35, 44, 211

U

Ulcus corneæ, 238  
Unit of antitoxin, 80  
Urine, complement fixation test for tuberculosis, 163, 174  
Urticaria, 227, 229

V

Vaccination against rabies, 26-30  
against small-pox, 25-26  
against tuberculosis, 30, 212-213  
against typhoid, 33-35  
Vaccines (Wright), 209-213  
dosage, 211-212  
preparation, 209-211  
Vacuum desiccator, 15, 16  
tuberculin, 62  
Venopuncture, 12, 13  
Viper's poison, 100  
Virulence of bacteria, 24  
method for determining, 134  
virulence of typhoid, 134  
diminishing virulence, 232  
increasing virulence, 135, 233  
Virus fixe, 27  
Vital staining, 198

W

Wall of leucocytes, 233  
Wassermann's reaction; see "complement  
fixation"  
Watery tuberculin, 62  
Weigert's law, 159  
Wet cupping for obtaining blood, 13  
Wet nurse, examination for syphilis, 169  
Whooping cough bacilli, 173  
Widal's test, 106-108  
Wright's pipettes and tubes, 203  
method for obtaining blood, 204

Zootoxins, 96-100





## INDEX OF AUTHORS

---

- Alessandrini, 99  
 Amiradzibi, 218  
 Anderson, 87  
 Apolant, 216  
 Arloing, 114  
 Arndt, 93  
 Aronson, 236, 239  
 Arrhenius, 86  
 Arthus, 222, 223, 227  
 Ascoli, 218, 219  
 Audeoud, 57  
 Auer, 227  
 Aufrecht, 68  
  
 Bail, 23, 31, 36, 38, 40, 42, 44, 46, 139  
 Bamberg, 103  
 Bandelier, 48, 69  
 Bandi, 238  
 Bashford, 216  
 Bassenge, 44  
 Bauer, 99, 189, 190  
 Benario, 247  
 v. Behring, 2, 30, 71, 77, 79, 80, 88, 160, 189, 190  
 Béranek, 63  
 Bergell, 239  
 Berger, 50  
 Berghaus, 82, 83  
 v. Bergmann, 102, 103  
 Bernstein, 220  
 Berthelm, 242  
 Bertrand, 99  
 Besredka, 223, 224, 239  
 Biedl, 227, 228  
 Bier, 14  
 Bitter, 245  
 Blaschko, 165  
 Blumenthal, 166, 242  
 Boas, 168  
 Bockenheimer, 88  
 Boer, 79  
 Bordet, 3, 31, 86, 120, 132, 143, 144, 152, 153, 154, 171  
 Borrel, 86  
 Borelli, 165  
 Brieger, 44, 102, 139, 187, 217  
 Bruck, 64, 92, 93, 155, 156, 164, 165  
 Buchner, 133  
  
 Calmette, 52, 88, 99, 100, 238  
 Castellani, 111, 113  
 Chamberland, 31  
 Chantemesse, 239  
 Christian, 160  
 Citron, 38, 52, 57, 62, 154, 160, 165, 166, 178, 184, 225, 233  
 Coca, 98  
 Cohen, 173  
 Conradi, 90, 109  
  
 Courmont, 114  
 Craig, 166  
 Czaplewski, 8  
  
 Dagonet, 216  
 Denys, 62, 198  
 Deutsch, 239  
 Deycke, 72  
 Diatropoff, 30  
 Doenitz, 83, 86  
 Doerr, 90, 91, 223  
 Doganoff, 50  
 Donath, 100, 101  
 Dopfer, 91  
 Douglas, 198  
 Doyen, 212  
 Dreyer, 245  
 Dunbar, 229  
 v. Dungen, 98, 100, 116, 144  
 Durham, 105  
  
 Ehrlich, 2, 79, 80, 81, 82, 85, 87, 95, 100, 132, 144, 147, 152, 158, 198, 240, 242, 246, 254  
 v. Eisler, 122  
 Ellerman, 55  
 Eppenstein, 52, 57  
 Epstein, 116, 118  
 Erlandsen, 55  
 van Ermenghem, 88  
 Escherich, 237  
  
 Ferran, 29  
 Ficker, 107  
 Fleischmann, 165  
 Fleming, 190  
 Flexner, 90, 97, 234, 235, 236  
 Flu, 245  
 Foix, 173, 236  
 Fornet, 22, 123  
 Fournier, 168  
 Fränkel, C., 77  
 Franz, 56  
 Freeman, 203  
 Freund, 217, 218  
 Friedberger, 11, 12, 16, 136, 161, 223, 225  
 Friedmann, 31  
 Friedman, 116  
 Friedemann, 223, 225  
 Fuld, 103  
  
 Garbat, 178, 187, 192, 239  
 Gay, 227  
 Gengou, 3, 143, 152, 153, 154  
 Ghedini, 170  
 Goetsch, 65  
 Graff, 218  
 Gross, 103  
 Gruber, 105, 233



- Hamburger, 47  
 Hartung, 227  
 Hata, 243, 244, 246  
 Hecht, 189  
 Helmann, 58  
 Hendersen-Smith, 85  
 Herbst, 128  
 Heubner, 83  
 Högyes, 29  
 Hoke, 44  
 Holzmann, 99  
 Hüppe, 44  
  
 Issaëff, 136  
 Iversen, 245, 246  
 Izar, 218  
  
 Jaffé, 142, 143  
 Jenner, 3, 25  
 Jensen, 216  
 Jobling, 234  
 Jochmann, 102, 234  
 Johannides, 246  
 Joseph, 51  
  
 Kaminer, 217, 218  
 Kelning, 58  
 Kempner, 89  
 Keysser, 161, 228, 255  
 Kikuchi, 44  
 Kitasato, 74, 77, 87  
 Kitashima, 77  
 Klausner, 124  
 Kleine, 115  
 Klinkert, 162, 163  
 Koch, R., 30, 45, 47, 48, 56, 60, 64, 71, 114, 242  
 Kolle, 32, 35, 44, 108, 138, 139  
 Koplik, 227  
 Korte, 141, 143  
 Kossel, 82  
 Krämer, 68  
 Kraus, R., 82, 85, 90, 91, 120, 178, 223, 227, 228, 234, 239  
 Kreibich, 124  
 Kruse, 90  
 Kuhn, 35  
 Kutscher, 114  
 Kyes, 97  
  
 Landsteiner, 100, 101, 144, 186, 188  
 Laubry, 170  
 Laveran, 241  
 Leclef, 198  
 Ledermann, 165  
 Leishman, 33, 198  
 Lenhartz, 67  
 Lesourd, 154, 192  
 Leuchs, 173, 192  
 Levaditi, 165, 197  
 Lewin, C., 216  
 Liefmann, 155  
 Liepmann, 216  
 Lignières, 50  
 Loewenstein, 49, 162  
 Löhlein, 199  
 Lorenz, 31  
 Lüdke, 192  
 Lustig, 238  
  
 Mac Fadyen, 239  
 Madsen, 78, 79, 83, 86, 89  
 Mallein, 173, 236  
 Mantoux, 51  
 Manwaring, 228  
 Maragliano, 62, 238  
 Marcus, 102  
 Marie, 30, 165  
 Markl, 238  
 Marmorek, 163, 174, 236, 238  
 Martin, 77  
 Marx, 82  
 Matthews, 213  
 Mayer, 44  
 McNeil, 191  
 Meier, G., 123, 165, 182, 183, 184, 186  
 Meakins, 191  
 Mendel, 51  
 Menzer, 236  
 Mesnil, 241  
 Metallnikoff, 72  
 Metschnikoff, 2, 138, 139, 151, 152, 196, 232  
 Meyer, 87  
 Meyer, F., 70, 83, 162, 165, 178, 236, 239  
 Meyer, K., 102, 103  
 Michaelis, G., 92, 93  
 Micheli, 165  
 Miessner, 128  
 Mita, 226, 228  
 Möller, 68  
 Moreschi, 154, 155  
 Morgenroth, 10, 13, 16, 83, 144, 147, 152, 158, 165, 255  
 Moro, 50  
 Moser, 236  
 Much, 99  
 Müller, 102, 186, 188, 191  
 Munk, 166, 187  
  
 Nakayama, 156, 158, 174  
 Negri, 26  
 Neisser, M., 92, 141-143, 155, 164, 165, 168, 193  
 Netter, 83  
 Neufeld, 166, 199, 213, 214  
 Nichols, 166  
 Nicolle, 153, 246  
 Nocard, 26  
 Noguchi, 26, 72, 97, 100, 187, 189  
  
 Obermeyer, 31, 129  
 Oppenheim, 191  
 Ostertag, 31  
 Ottenberg, 116, 118  
 Otto, 89, 223  
  
 Pane, 237  
 Parvu, 170  
 Pasteur, 3, 26, 27, 41  
 Petit, 57  
 Petruschky, 66  
 Pfeiffer, 35, 44, 108, 131, 134-140, 198, 223, 226, 228  
 Phisalix, 99  
 Pick, 31, 129  
 Pickert, 162  
 v. Pirquet, 48, 52, 55, 223, 224, 226  
 Plato, 58  
 Plaut, 123, 164, 165  
 Porges, 122, 123, 186

- Portiers, 222  
 Pötlz, 186, 188  
 Prausnitz, 230  
  
 Rabinowitsch, 158  
 Radziewsky, 136  
 Ransom, 87  
 Remlinger, 26, 30  
 Reschad, 72  
 Ribbert, 233  
 Richet, 222  
 Rietschel, 169  
 Römer, 51, 82, 95, 237  
 Röpke, 48, 70  
 Rosculet, 92  
 Rosenau, 87, 223  
 Rosenblatt, 160  
 Rosenthal, 90, 91  
 Roux, 26, 31, 74, 77, 82, 86  
 Ruppel, 70, 162, 234  
 Russel, 34  
  
 Sachs, H., 89, 97, 155, 186, 193  
 Sahli, 63  
 Salimbeni, 238  
 Salmon, 31  
 Salomonsen, 78, 79  
 Salus, 44  
 Schaudinn, 242  
 Schenck, 57  
 Schick, 223, 224, 227  
 Schittenhelm, 223, 226  
 Schleissner, 173, 236  
 Schöne, 192  
 Schucht, 164, 165  
 Schulze, 92, 93  
 Schütze, 125, 164, 165  
 Schwartz, 191  
 Schwarz, 82  
 Slavo, 239  
 Seiffert, 57  
 Seligmann, 165  
 Shiga, 44, 98  
 Simons, 220  
 Smith, 31  
 Smith, Theob., 222, 227  
 Sobernheim, 239  
 Southardt, 227  
 Spengler, 62  
 Steinberg, 143  
  
 Steinhardt, 224  
 Stenitzer, 239  
 Stern, Marg., 165, 190  
 Stern, 141, 148  
 Stertz, 165  
 Strauss, 13  
 Szaboky, 99  
  
 Takaki, 86  
 Tallquist, 100  
 Tavel, 238, 239  
 Teague, 191  
 Teichmann, 59  
 Terni, 238  
 Todd, 90, 91  
 Töpfer, 142, 143  
 Torrey, 191  
 Toussaint, 31  
 Traube, 218  
 Trebing, 102  
 Tschernogubow, 189  
 Tschistowitsch, 120  
  
 Uhlenhuth, 9, 11, 125-130, 242  
  
 Vaillard, 84  
 Vannod, 191  
 Vigano, 218  
  
 Wassermann, A., 2, 3, 11, 38, 64, 86, 89, 114,  
     125, 155, 156, 161, 164, 165, 178,  
     232, 255, 256  
 Wassermann, M., 228, 255  
 Wechsberg, 92, 141-143  
 Weichardt, 218, 223, 225, 229  
 Weidanz, 189  
 Weigert, 159  
 Weil, 36, 41, 156, 158, 174  
 Weinberg, 170, 190  
 Werner, 246  
 Widai, 106, 154, 192  
 Wolff-Eisner, 52, 54, 59, 223, 224, 225, 229,  
     231  
 Wollstein, 191  
 Wright, 3, 32, 33, 44, 108, 198, 199, 200,  
     209, etc.  
  
 Yersin, 74, 238  
  
 Zeuner, 72  
 Zupnik, 87











0126-135  
n 352

YC 88631

360085

QR181

C6

1914

cop. 2

ZOOLOGY  
LIBRARY

UNIVERSITY OF CALIFORNIA LIBRARY



